

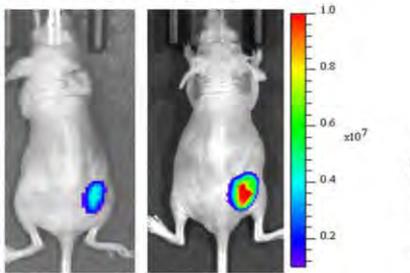
Biomedical Research Journal

OCTOBER 2015 | VOLUME 2 | ISSUE 2

pISSN: 2349-3666; eISSN: 2349-3674

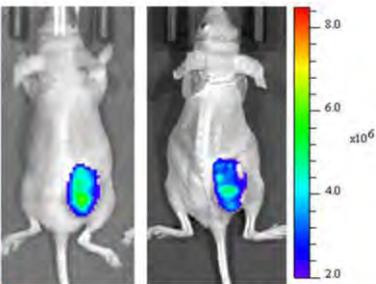
Bioluminescence monitoring of ^{131}I treatment effect on breast cancer tumor xenograft

Untreated control group



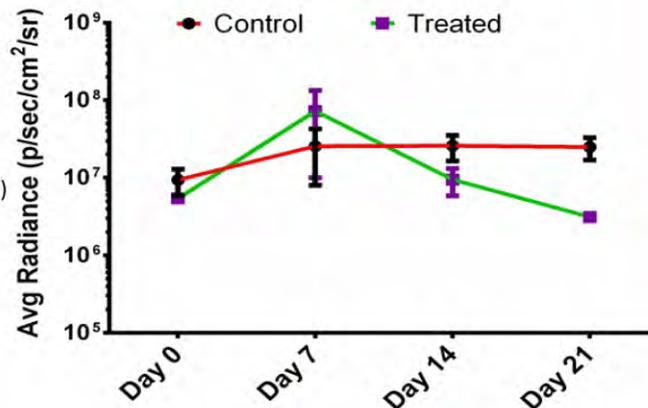
Radiance (P/s/cm²/sr)

^{131}I treated group



Day 0

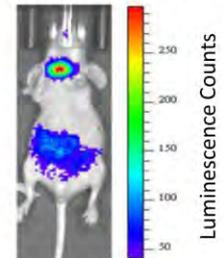
Day 21



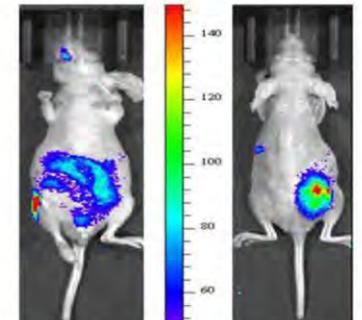
1 mCi ^{131}I treatment per week

Cerenkov luminescence imaging

Natural accumulation of ^{131}I in NIS over-expressing thyroid gland



^{131}I uptake in thyroid tissues can be blocked by T3+Methimazole



Ventral

Dorsal

Biomedical Research Journal

OCTOBER 2015 | VOLUME 2 | ISSUE 2

EDITORS-IN-CHIEF

Dhananjaya Saranath (Mumbai, India)
Aparna Khanna (Mumbai, India)

SECTION EDITORS

Cancer Biology:

Girish Maru (Navi Mumbai, India)

Stem Cell Biology:

Vaijayanti P. Kale (Pune, India)

Nanotechnology:

Vilas G. Gaikar (Mumbai, India)

Phytochemistry:

Lokesh Bhatt (Mumbai, India)

EDITORIAL BOARD

Ali Syed Arbab (Detroit, USA)

Amit Agarwal (Bangalore, India)

Anjali A. Karande (Bangalore, India)

Basuthkar J. Rao (Mumbai, India)

Hemant Malhotra (Jaipur, India)

Kirti S. Laddha (Mumbai, India)

Mohan C. Vemuri (Frederick, USA)

Nancy Pandita (Mumbai, India)

Paul J. Verma (Rosedale, Australia)

Pritish Bhattacharya (New Jersey, USA)

Purvish M. Parikh (Mumbai, India)

Sai Yendamuri (New York, USA)

Sumitra Chanda (Rajkot, India)

Surinder K. Mehta (Chandigarh, India)

Victoria M. Villaflor (Chicago, USA)

Alpana Ray (Missouri, USA)

Anandwardhan Hardikar (Sydney, Australia)

Ashok B. Vaidya (Mumbai, India)

Dhirendra Bahadur (Mumbai, India)

Karuna Shanker (Lucknow, India)

Mayur Yergeri (Mumbai, India)

Naganand Rayapuram (Evry, France)

Partha Basu (Kolkata, India)

Prasad S. Adusumilli (New York, USA)

Pulok Mukherjee (Kolkata, India)

Ramesh Goyal (Ahmedabad, India)

Sukhinder Kaur Cheema (St. John's, Canada)

Sunita Saxena (New Delhi, India)

Tania Fernandez (San Francisco, USA)

EDITORIAL OFFICE

Sunandan Divatia School of Science,
NMIMS (Deemed-to-be University)
Bhaidas Sabhagriha Building, Bhaktivedanta Swami Marg,
Vile Parle (W), Mumbai 400056, India.
Email: brj.sos@nmims.edu

EDITORIAL ASSISTANT

Brijesh S. (Mumbai, India)

General Information

Aims and Scope

“Biomedical Research Journal (BRJ)” is a premier peer reviewed open access journal, published by Sunandan Divatia School of Science, NMIMS (Deemed-to-be) University, for promoting the advancement of ideas in the interdisciplinary realms of Medicine, Science and Technology. The goal is to share new discoveries and translational knowledge with scientists, academicians, clinicians and students in the field of Biomedical and Biology/Chemistry/ Biotechnology/Stem Cell Biology/Cancer Biology in the realm of basic and applied aspects in the different areas.

BRJ aims at creating a platform to help advance the domains and frontiers of inter- and multi-disciplinary research across the various areas of sciences and recent advances in cross pollination across biology, chemistry, and medicine. Integrative science is the present and future of science, and the journal proposes to highlight and emphasize contemporary technology towards understanding various aspects of the sciences.

The initial focus areas of BRJ include review articles and original research papers in cancer biology, stem cell biology, nano-technology and phytochemistry.

A rigorous peer review process is implemented to judge the effectiveness, legitimacy and reliability of the research content. The papers will be published online as

well as provide hard copy of the Journal issues to the authors of the papers on request.

Information for Subscribers

BRJ is planned as a six monthly publication with two issues published in the first year. Currently, there are no subscription charges for the journal and can be accessed online. For submission instructions, subscription and additional information please visit: <http://science.nmims.edu>

Disclaimer

The views and opinions expressed in the articles published in the journal are the sole responsibility of the authors. The Publisher, NMIMS School of Science and the Editors cannot be held responsible for errors or any consequences from the use of information contained in this journal.

Copyright

The Journal grants all users a free, permanent, worldwide, continuous right of access to, and a license to copy, use, distribute, perform and display the work publicly and to make and distribute derivative works in any digital medium for any reasonable non-commercial purpose, subject to proper citation of authorship and ownership rights. The journal also grants the right to make a printed copy for personal non-commercial use only.

Editorial: Right Patient, Right Diagnosis, Right Treatment! Dhananjaya Saranath and Aparna Khanna	134
Areca Nut Use and Cancer in India Prakash C. Gupta and Cecily S. Ray	140
Identification of Therapeutic Targets for Cancer: Proteomic Technologies and Strategies are the Key to Success Rukmini B. Govekar	166
Genetic Markers and Evolution of Targeted Therapy in Cancer Pratibha S. Kadam Amare	179
Cancer Gene Therapy: Prospects of Using Human Sodium Iodide Symporter Gene in Non-thyroidal Cancer Shruti Dutta and Abhijit De	198
Microbiota in Immune Pathogenesis and the Prospects for Pre and Probiotic Dietetics in Psoriasis Garima Pandey, Abhay Kumar Pandey, S. S. Pandey, and B. L. Pandey	220

Right Patient, Right Diagnosis, Right Treatment!

Dhananjaya Saranath and Aparna Khanna

The 2015 Nobel Prize in Medicine for drugs to fight malaria and other tropical diseases, and in Chemistry for fundamental contributions towards understanding DNA repair and maintaining of genomic integrity in cells, highlights the interdisciplinary approach for maximizing benefits of contemporary science to mankind. The Nobel Prize in Medicine was awarded to William Campbell Ph.D., born in Ireland and migrated to US; Satoshi Omura, Ph.D., from Japan, and Youyou Tu, the first Chinese Nobel laureate. The Nobel laureates – Dr. Campbell and Dr. Omura were cited for their discovery of Avermectin, derivatives of the drug responsible for decreasing incidence of river blindness and lymphatic filariasis affecting millions in Asia and Africa. Ms. Tu's intensive efforts led to the active compound from the herbal Chinese sweet wormwood plant, giving us the antimalarial drug artemisinin, currently the first line drug for malaria affecting 50% of global population. The Chemistry

Nobel acknowledged three scientists for their research in DNA repair, for their intensive work on mapping the process at a molecular level and providing insights into cell functioning and maintenance of genomic stability. The Nobel laureates were Dr. Thomas Lindahl, Ph.D., Francis Crick Institute, London, for his discoveries in base excision repair; Dr. Paul Modrich, Ph.D., Howard Hughes Medical Institute, and Duke University School of Medicine, North Carolina, USA, for the mismatch repair pathway; and Dr. Aziz Sancar, M.D., Ph.D., at the University of North Carolina, USA, for nucleotide excision pathway. The understanding of DNA repair mechanisms in the cells is a breakthrough in understanding how cancer develops and furthers treatment of cancer and also several diseases, much needed for better health management.

Despite the tremendous advances in technology, particularly biotechnology, information technology and imaging technology, cancer development,

progression, response to treatment, and recurrence is still an enigma with several glaring lacunae. It was in 1971, Richard M Nixon, 37th President of US, signed a bill, 'The National Cancer Act', to create new research infrastructure with enormous resources devoted to fighting cancer, and the act was known as 'Nixons war on cancer'. By 2005, NCI spent USD 165 billion, and the outcome was better understanding, and applications of the generated data resulted in reduced mortality from cancer in both men and women. The outcome of genome based screening in Cervical Cancer, Breast Cancer, Colon Cancer and other cancers was early detection, better prognosis and better survival. Genome wide association studies gave us 'Predictive markers', and the whole genome expression studies through transcriptomics, proteomics, metabolomics gave us 'Prognostic Biomarkers' and identified 'Drug Targets' with cutting edge technology. The 'Next Generation Sequencing' with massive parallel sequencing over the past several years, is anticipated to be an invaluable part of clinical medicine.

With the current 1.2 billion population in India, preventive medicine often takes a back seat, although the focus of health

management in developed and developing world should be 'Prevention' in monogenic and multigenic complex diseases including 'Cancer'. Despite oral cancer ranked as the number one cancer in Indian males and fourth most common cancer in females, contributing 26% of the global oral cancer burden, preventive measures are slow in implementation. Tobacco has been unequivocally established as a high risk factor in oral and lung cancers; however, the role of areca nut is not commonly acknowledged as an important risk factor for oral cancer. In the current issue Dr. Prakash Gupta and Cecily Ray, Healis Sekhsaria Institute for Public Health, Mumbai, in their article on 'Areca nut use and cancer in India', give us a comprehensive review of the evidence for carcinogenicity of areca nut, detail epidemiological and animal studies, and reveal the mechanistic evidence highlighting the causal biochemical and molecular mechanisms of oral submucous fibrosis – a premalignant condition and oral cancer in areca nut chewers. The review emphasizes the necessity for awareness programs for areca nut hazards and control policies on areca nut per se and areca nut products.

Inherent in all health management

programs is the understanding of the mechanisms of diseases using state of the art technology. The concepts of 'Systems Biology' and 'Omics' was knowledge based and advanced applications in clinical medicine. Dr. Rukmini Govekar, Advanced Centre for Treatment, Research and Cancer, Navi Mumbai has lucidly highlighted proteomic technologies and strategies in her article 'Identification of Therapeutic Technologies and Strategies are the Key to Success'. The 'Omics' research has immensely contributed to

'Targeted therapy' in cancer, and is a rapidly resulting in pathology specific therapeutic molecules relevant to an expanding list of tumor types. Thus, the targeted therapy is useful not only in the initially indicated cancer, but is useful in several cancers with identical or similar molecular pathology. The therapy is tailored to the individual patient, with 'Companion Diagnostics' giving a helping hand in the informed decision for the patient. Off label use of targeted therapies will be indicated as confirmed data such as BRCA1/BRCA2 gene mutations as predictive diagnostic test for breast cancer, and hence use of targeted therapy with the specific molecular pathology, is useful in Pancreatic cancers and Non-Small Cell

Lung Cancer patients. The recent immune checkpoint blockade therapies, Anti-PDL1, may be used effectively in advanced melanoma or metastatic bladder cancer with impressive clinical responses. A massive effort for high incidence and rare cancers is the NCI-Molecular Analysis for Therapy Choice (NCI-MATCH) Trial, world's largest ever molecular oncology trial, screening 3000 patients for comprehensive molecular pathology, and treated with targeted therapy. Biopsy specimens from tumors will be analyzed for more than 4,000 different variants across 143 genes, regardless of tumor origin. Dr. Pratibha Kadam Amare, Tata Memorial Hospital, Mumbai, gives a snap shot of molecular pathology and targeted therapy in her review on 'Genetic markers and evolution of targeted therapy in cancer'. The author elaborates the cell surface antigens and tyrosine kinase targets identified as pathognomonic in several common cancers, and development of effective inhibitor molecules as small molecules or antibodies for therapy, a must read for our oncologists and basic scientists.

The re-emergence of 'Gene Therapy' recently is obvious. The field has matured, gathering steam and is anticipated to join

mainstream therapy in several monogenic diseases as also complex diseases including cancer, after several years of taking a backseat in both the research realm and clinical applications. The falling precipice in gene therapy truly began in 1999 when 18 year old Jesse Gelsinger died from multiple organ failure four days after treatment for ornithine transcarboxylase deficiency, the death triggered by severe response to adenoviral carrier. The disenchantment with gene therapy continued when several children developed leukemia like condition after treatment for X-linked severe combined immunodeficiency disease. The current scenario is quite different with several companies conducting clinical trials with gene based therapy using AAV vector, oncolytic viruses, liposomes coated with Polyethylene Glycol, siRNA, etc. And the clinical trials using gene therapy in varied diseases – Parkinson disease, the drug used is Nerologix (Biotech Company); A successful correction of inherited form of blindness has been launched by Spark Therapeutics; Inhalation of normal Cystic Fibrosis CFTR gene copies for improvement in lung function for Cystic fibrosis patients. Gene therapy with siRNA to degrade particular RNA

sequences is under investigation for Huntington's disease; and lentivirus based gene transfer to improve efficiency of gene transfer in metachromatic leukodystrophy with the pathology being mutated arylsulfatase A enzyme. The lentivirus vector avoids activation of cancer causing genes by loading the vector with self-inactivating promoter sequences that exclusively induce expression of the therapeutic gene. The advances in gene delivery systems will relegate complex diseases such as cancers to a manageable disease without severe disability, disfigurement and death. Dr. Abhijit De and Shruti Dutta in their article 'Gene therapy for sodium iodide symporter in non-thyroidal cancers' focus on potential suicide genes in cancer supporting diagnosis through imaging of the cancer and therapeutic applications, with emphasis on sodium iodide symporter. The authors emphasize the effectiveness of sodium iodide symporter in non-thyroidal cancers, particularly hormone receptor negative patients.

The technology today facilitates a 'Systems Biology' approach for understanding the role of 'microbiome' at different sites of our body, with emergence of functional metagenomics, to enable a

holistic picture of the beneficial normal microbes. Subtle imbalances in our microbial population can cause diseases, influences our response to drugs, and restore balance between microbial normalcy and pathogenesis, leading to effective cures. Global parameters of microbial communities provide valuable information regarding human health status and disease predisposition. Downstream analyses of the functional interaction between host and the microbiome deciphering the metagenome i.e. the human microbiome and its collective genes, may lead to mechanistic insights into the interaction, with new opportunities for next generation diagnosis, prognosis and treatment of various acute, chronic, localized, systemic, simple and complex diseases. *In vitro* cultivation of the microbiota earlier formed the cornerstone in microbiology, though not possible in densely populated microbial communities. DNA based analyses expands the horizons generating enormous new data sets mined for information. The Human Microbiome Project (by NIH), generated 2.3 terabyte 16S rRNA metagenomics dataset of over 35 billion reads from 690 samples of 300 subjects across 15 body sites. The

microbiome and core metagenomics occupies a specific human niche, and varies both by anatomical site and substantial interpersonal variation, with a causal link established between microbiome variation and significant pathology. Dietary changes rapidly cause substantial metagenomic changes. Dr. Abhay Kumar Pandey, Banaras Hindu University, Varanasi, and colleagues succinctly highlight the important role of the microbiome in disease and health in the review 'Microbiota in immune pathogenesis and the prospects for Pre and Probiotic Dietetics in psoriasis'. Psoriasis is a chronic idiopathic, inflammatory dermatologic condition, with the cutaneous microbiome a critical target. The pathogenesis is advanced by genetic predisposition, and a change in the microbiota is observed, with increased ratio of Firmicutes to Actinobacteria. The alterations in commensal microbiome, basis for the changes in disease, and the mechanisms of colonization and host homeostasis is restored, is crucial for manipulating the host microbiota. The authors elaborate the role of dietary prebiotics and probiotics towards healthy host microbiome relationship via dietary probiotics and manipulation of the specific

site and general gut microbiota. It is feasible that the advances in technology facilitating better understanding of disease pathogenesis will prove fruitful and provide optimal benefits and the end of 'Right diagnosis, Right treatment, Right patient,' is in sight, making therapeutics robust and dynamic.

Areca Nut Use and Cancer in India

Prakash C. Gupta and Cecily S. Ray*

Healis Sekhsaria Institute for Public Health, 501 Technocity, Plot X-4/5 TTC Industrial Area, Mahape, Navi Mumbai – 400701, India

Areca nut is widely used in India and the consumption has increased over the past two decades, with availability in new dry packaged forms (*pan masala*, *gutka*, *mawa*). Recent reports of increasing mouth cancer incidence have suggested an association with areca nut consumption. Here we have reviewed the evidence for carcinogenicity of areca nut, including epidemiological studies, several animal studies and mechanistic evidence. Studies primarily from India, providing odds ratios (ORs) or relative risks for precancers or cancer with use of areca nut without inclusion of tobacco is the focus of the review. Six case-control studies on oral submucous fibrosis (OSF) had significantly elevated ORs for use of areca nut in various forms. Six case-control studies on head and neck cancers, primarily oral cancer reported elevated ORs for chewing of betel quid without tobacco. Eight case control studies on oral cancer have reported elevated and significant ORs for betel quid with tobacco. A significant risk in oral cancer was noted in *gutka* users. Animal studies confirmed correlation between development of precancers or cancers and exposure to areca nut or *pan masala* without tobacco. Mechanistic evidence shows a role for areca nut alkaloids, polyphenols and copper in promoting carcinogenesis. Our review emphasizes control policies on areca nut products and appropriate mass communication programs for awareness of hazards of areca nut with emphasis on areca nut *per se*.

INTRODUCTION

The areca nut, fruit of the oriental palm (*Areca catechu*), also called 'betel' nut in English, *supari* in Hindi, *adike* or *betta* in Kannada, *adakka* in Malayalam, and *pakku* in Tamil, is commonly used in India (FRLHT.org, 2015) and needs no introduction. It is used in traditional quids (*beeda*) wrapped in betel leaves (*Piper betle*) or as tobacco and areca nut mixtures. Areca nut is also used *per se* and available in specialised shops and by roadside vendors, in sachets, as *pan masala* and *gutka*. A product containing areca nut chips, slaked lime and tobacco, popularised in Gujarat, is called *mawa* (Gupta, 1998), also sold in Maharashtra as *kharra* (Hazare *et al.*, 1998). *Mainpuri* tobacco containing similar ingredients is

Key words: Areca nut, Prevalence, India, Oral submucous fibrosis, Leukoplakia, Neoplasms, Case control studies, Laboratory studies, Oxidative stress, DNA damage.

***Corresponding Author:** Cecily S. Ray, Healis Sekhsaria Institute for Public Health, 501 Technocity, Plot X-4/5 TTC Industrial Area, Mahape, Navi Mumbai – 400701, India.

Email: raycs@healis.org

consumed in Uttar Pradesh, since the 1960s (Wahi, 1968). These commercial developments resulted in doubling areca nut consumption in India during 1991 to 2010 from 2.5 to 5.2 lakh tons, with about 5% increase each year (Kammardi *et al.*, 2012).

A recent report of the National Cancer Registry Programme (ICMR) showed an increasing incidence of cancer of the buccal mucosa ('mouth cancer') for six to ten years up to 2009 or 2010 in five of the nine population-based cancer registries (NCDIR-NCRP, 2013), reiterated by a similar trend in a single registry (Gupta *et al.*, 2014). The cancer registries located in Bhopal, Mumbai, Delhi, Dibrugarh and Ahmedabad rural and urban, in the states or territory of Madhya Pradesh, Maharashtra, Delhi Union Territory, Assam and Gujarat, respectively have high prevalence of high areca nut use (IIPS & MOHFW, 2010). In addition to the increased incidence, patients of oral cancer are younger than 35 years of age since the mid-1990s as compared to the mid-1980s (Gupta, 1999).

Betel quid has been linked with head and neck cancers including oral cancers since the last century, although at that time tobacco and lime in the betel quid were

viewed as the likely causes of the associated cancer (IARC, 2004; Orr, 1933). In the past decade, oral cancer has been diagnosed with increasing frequency in young users (< 35 years) of packaged areca nut products, bringing the potential carcinogenicity of areca nut into focus (Chaudhry, 1999; Gupta, 1999). An extensive review and evaluation of evidence was undertaken on areca nut and betel quid by the International Agency for Research on Cancer (IARC) reported in 2004. The evidence for carcinogenicity of areca nut, primarily from India and South Asia was from use of betel quid with tobacco. Relatively few epidemiological studies on precancers or cancer, in the past twenty years reported on cancer risks associated with use of betel quid without tobacco or use of industrially manufactured areca nut products. Nevertheless these few studies, along with laboratory evidence, made it possible for the monograph to conclude that areca nut by itself is carcinogenic to humans. The final evaluation by IARC concluded that betel quid without tobacco causes cancer of the oral cavity, and betel quid with tobacco causes cancer in the oral cavity, pharynx and esophagus; and emphasized that areca nut is carcinogenic to humans

(IARC, 2004). In the most recent monograph (Vol. 100E) several additional studies were reviewed and the evidence confirmed carcinogenicity of areca nut in humans and animals. However, the message has apparently not reached the masses, perhaps ignored or discounted, in view of the overwhelming evidence of carcinogenicity of tobacco.

Thus, in view of increasing consumption of areca nut products in India and reports of increasing oral cancer incidence over the past ten years, a review of currently available evidence of the carcinogenicity of areca nut was undertaken. An initial literature survey of the use of areca nut products in India, followed by epidemiological and laboratory evidence for the role of areca nut in causing oral cancer and other head and neck precancers, and an outline of the mechanisms of cancer causation are reviewed.

MATERIALS AND METHODS

Literature on carcinogenicity of areca nut and its products as used in India (areca nut, betel quid or *paan*, *gutka*, *pan masala*, *mawa*) was surveyed. Since use of areca nut without tobacco has been rare, earlier epidemiological studies have generally not

reported separate risks for areca nut. However, since tobacco is widely recognised as carcinogenic, and areca nut has not been associated with cancer we focused on case-control studies that reported ORs for use of areca nut without tobacco. Epidemiological studies reporting on oral precancers or oral and pharyngeal cancers are included. The evaluation monographs of the IARC, volumes 85 (2004) entitled, "Betel-quid and areca-nut chewing and some areca-nut-derived nitrosamines" (2004), and 100E (2009) on "Betel quid and areca nut" were used as the basic resources, along with internet searches in Pubmed for case control studies, cohort studies, animal experiments and mechanistic studies. The more recent studies are emphasized, with a few highly informative earlier studies included. Certain studies on oral submucous fibrosis (OSF) not reviewed in the IARC Monographs are emphasized (Bathi *et al.* 2009; Mehrotra *et al.*, 2013). Research conducted in India is prioritized, and additional studies in other parts of the world cited to provide evidence of areca nut as an important carcinogen globally are included. In addition, basic prevalence data on use of areca nut products were obtained from the Global Adult Tobacco

Survey for India (GATS) (International Institute for Population Sciences and Ministry of Health and Family Welfare, Government of India, 2010).

RESULTS

Prevalence of Areca nut Use in India

The report of the GATS for India showed betel quid with tobacco was used by 7.5% men and 4.9% women, and mixtures of areca nut and tobacco, without betel leaf (*gutka* and *mawa*) used by 13.1% men and 2.9% women. The report did not provide

data for use of areca nut without tobacco (Table 1). The data showed that use of pre-packaged imperishable forms of areca nut have superseded the popularity of betel quid.

In rural areas, the prevalence of betel quid with tobacco was higher in urban areas (6.8% rural vs. 4.8% urban), prevalence of *gutka* and similar products in rural areas was higher than in urban areas (8.6% rural vs. 7.1% urban). The regions with high prevalence of use of areca nut products in India were the

Table 1: Prevalence of use of products containing areca nut and tobacco among persons ≥ 15 years in India, from the GATS (International Institute for Population Sciences and Ministry of Health and Family Welfare, Government of India, 2010).

Substances used	<i>All current users</i>	Males	Females
	N %	N %	N %
<i>Gutka</i> & other areca nut mixtures with tobacco and lime	65,072,000 8.2	13.1	2.9
	Range: 2.8–12.1 North to Central	Range: 4.9–18.4 North to Central	Range: 0.2–5.0 North to Central
Betel quid with tobacco:	49,672,000 6.2	7.5	4.9
	Range: 0.7–17.2 North to Northeast	Range: 1.1–18.9 North to Northeast	Range: 0.3–15.6 North to Northeast
<i>Pan masala</i> or betel quid without tobacco; and/ or use of nasal snuff	35,106,000 4.4	3.5	5.4
	Range: 0.8–10.9 North to East	Range: 1.0–10.5 North to Northeast	Range: 2.3–15.0 West to East

Note: Total prevalence could not be calculated from these values as the categories are not mutually exclusive.

Key to regions:

North-East: Sikkim, Arunachal Pradesh, Nagaland, Manipur, Mizoram, Tripura, Meghalaya and Assam

East: West Bengal, Jharkhand, Odisha and Bihar;

Central: Rajasthan, Uttar Pradesh, Chhattisgarh and Madhya Pradesh;

North: Haryana and northwards including Jammu & Kashmir, Himachal Pradesh, Punjab, Chandigarh, Uttarakhand and Delhi.

Northeast (Sikkim, Arunachal Pradesh, Nagaland, Manipur, Mizoram, Tripura, Meghalaya and Assam), the East (West Bengal, Jharkhand, Odisha and Bihar) and the Central region (Rajasthan, Uttar Pradesh, Chhattisgarh and Madhya Pradesh). Low prevalence was found in the North (Haryana and northwards including Jammu & Kashmir, Himachal Pradesh, Punjab, Chandigarh, Uttarakhand and Delhi). In particular, prevalence of betel quid with tobacco was high in the Northeast (17.2%) and East (9.7%), and lowest in the North (5.5%). On the other hand, prevalence of *gutka* and similar mixtures was high in the Central states (12.1%). Among men, *gutka* use was concentrated among the 15 to 44 year age group, whereas women users tended to be older. Betel quid with tobacco was used mainly among the 45–65 year age groups in both men and women (International Institute for Population Sciences and Ministry of Health and Family Welfare, Government of India, 2010).

Occasional consumption of areca nut without betel leaf, lime and condiments has been a norm and a common culturally accepted practice in India (Reddy and Gupta, 2004). Areca nut consumption without tobacco and by itself has been

occasionally reported as practiced by a small fraction of the population before the 1980s (Mehta *et al.*, 1972). In the last 10–15 years areca nut habits have been observed in children (Chaturvedi *et al.*, 2002; Khandelwal *et al.*, 2012).

Evidence of Carcinogenicity in Humans

In India, a quid containing areca nut is chewed and kept next to the cheek (buccal) mucosa, for hours including overnight. Blanching often appears at the site as an early sign of OSF and squamous cell carcinoma may develop. Various case-control studies on precancers and cancers associated with areca nut use are summarised in the following section.

Oral Precancers

Six case control studies on OSF, five from India in the states of Bihar, Gujarat, Kerala, Karnataka and Uttar Pradesh (Ahmad *et al.*, 2006; Bathi *et al.*, 2009; Jacob *et al.*, 2004; Mehrotra *et al.*, 2013; Sinor *et al.*, 1990) and one from Sindh in Pakistan (Maher *et al.*, 1994), showed significantly elevated ORs for OSF associated with areca nut use without tobacco in various forms (Table 2). ORs for OSF for betel quid without tobacco (BQ) ranged from 1.3, the lowest, which

Table 2. Oral submucous fibrosis and use of areca nut (AN) in various forms, in case control studies conducted in India and Pakistan (both men and women).

Location	Chewing status	OSF Cases	Controls	OR (CI) OSF	Study type, Matching, Adjustments, Stratifications	References Notes
Bhavnagar, Gujarat, India	Non Chewer (currently) ^a BQ or AN (no tobacco)	1	39	1.0 (ref.)	Dental clinic based. Age ≥15 yrs. Matched on age, sex, religion, and occupation.	(Sinor <i>et al.</i> , 1990) Significant dose response for frequency & duration
Karachi, Pakistan	None (includes ex-chewers > 6 m) BQ (no tobacco) AN alone	2 7 64	82 9 17	1 (ref.) 32 (6–177)** 154 (34–693)**	Dental clinic based. Matched on sex and age.	(Maher <i>et al.</i> , 1994) Significant dose response for frequency & duration
Kerala (rural), India	No chewing (currently) BQ (no tobacco) Areca nut only	9 15 0	31884 1100 12	1.0 (ref.) 47.2 (20.2–110.4) -	Population based. Age >35 yrs Adjusted for age, sex, education, smoking and alcohol drinking.	(Jacob <i>et al.</i> , 2004) Significant dose response for frequency
Patna, Bihar, India	No areca nut product use BQ not specified AN alone <i>Pan masala</i>	5 25 8 32	108 13 1 5	1.0 (ref.) 41.5 (13.5–127.2) 172.8 (18.0–1662.6) 138.2 (37.6–507.7)	Dental clinic based. Matched on age, sex, religion & socio-economic status. ORs calculated by review authors	(Ahmad <i>et al.</i> , 2006)
Dharwad, Karnataka, India	No habit (currently) <i>Pan masala</i> /BQ/areca nut BQ (no tobacco)	1 2 23	119 4 43	1.0 (ref.) 59.5 (3.1–2154) 63.6 (8.7–1304) Confidence intervals recalculated by review authors	Hospital based. Matched on age, sex, and socio-economic status.	(Bathi <i>et al.</i> , 2009) Dose response calculation included diverse product users, not informative
Lucknow (urban, rural), Uttar Pradesh, India	Non users (currently) Tobaccoless products BQ <i>Pan masala</i> Both Total visitors to camps	NA NA 448	NA NA 2688	(ref.) 1.3 (0.95–2.7) 3.0 (1.2–7.4) 6.4 (4.3–9.5)	Subjects from urban and rural health camps Matched on age, and socio-econ. status. Males: (89.1% cases); (82% controls).	(Mehrotra, <i>et al.</i> , 2013) Dose response seen for frequency per day for each product

CI= Confidence intervals; AN=areca nut; BQ=Betel quid without tobacco; BQT= Betel quid with tobacco.

^aOccasional chewer of AN**P* < 0.01, ***P* < 0.0001

did not reach significance (Mehrotra *et al.*, 2013) to 78.0 (Sinor *et al.*, 1990). In contrast, ORs for BQ with tobacco ranged from 7.9 (Mehrotra *et al.*, 2013) to 64 (Maher *et al.*, 1994). Use of areca nut alone reported in two studies had ORs of 154 (Maher *et al.*, 1994) and 172 (Ahmad *et al.*, 2006). ORs exclusively for tobacco-less *pan masala* use in two studies were 3.0 (Mehrotra *et al.*, 2013) and 138.2 (Ahmad *et al.*, 2006).

Ors tended to be higher for users of mixtures made with areca nut and tobacco but without betel leaf, such as *mawa* (106.4) (Sinor *et al.*, 1990), or *gutka* (from 10.8 to 1142) (Bathi *et al.*, 2009; Mehrotra, *et al.*, 2013). Additionally, a cross-sectional house-to-house study showed an OR for men with OSF as 75.6 for *mawa* chewing in 11,262 men in Bhavnagar District of Gujarat (Gupta *et al.*, 1998).

Studies were not adjusted for smoking, with one exception that also studied leukoplakia (Jacob *et al.*, 2004). However, the report of the earliest study stated the rate of smoking in cases and controls was similar and also that smoking did not appear play a role in the development of OSF (Sinor *et al.*, 1993). Another study performed multiple logistic regression on smoking and OSF and reported negligible

effect of smoking (Bathi *et al.*, 2009). Few smokers were found among chewers in a study and they were excluded from calculation of ORs (Maher *et al.*, 1994). Mehrotra *et al.* (2013) concluded that tobacco smoking did not affect risk of OSF, whereas alcohol consumption increased the risk in chewers of tobacco-less betel quid or *pan masala* several fold.

A dose response was clearly seen for frequency per day of using areca nut preparations in four of the studies (Jacob *et al.*, 2004; Maher *et al.*, 1994; Mehrotra *et al.*, 2013; Sinor *et al.*, 1990). A clear dose response was also clearly seen for duration (Jacob *et al.*, 2004; Maher *et al.*, 1994; Sinor, *et al.*, 1990).

Two case control studies, one with betel quid and *pan masala* (Shah and Sharma, 1998) and the other with *pan masala*, *kharra*, tobacco-lime and betel quid in different combinations (Hazare *et al.*, 1998), reported significant increasing trends for frequency of use of areca nut containing substances per day ($p < 0.01$), although overall ORs for OSF was not reported. An increasing prevalence of OSF was observed between 2000 and 2004 with 77.8% of OSF patients using multiple areca nut products (Hazare *et al.*, 2007).

Table 3. Selected precancers and use of areca nut without tobacco in Kerala, India (Both men and women).

Location	Chewing status	Cases	Controls	OR (CI)	Study type, Matching, Adjustments, & Stratifications	Reference
Kerala				Leukoplakia:	Population based.	(Jacob <i>et al.</i> , 2004)
	Non chewers	176	31884	1.00 (ref.)	Aged >35 yrs.	
	BQ	27	1100	4.0 (2.7–6.1)	Adjusted for smoking & alcohol drinking.	
	AN only	1	12	12.8 (1.6–101.2)		
				Erythroplakia:		
	Non chewers	8	31884	1.00 (ref.)		
	BQ	4	1100	12.5 (3.70–42.4)		

CI = Confidence intervals. BQ = Betel quid without tobacco; AN = Areca nut. Note: Use of lime may be inferred.

Leukoplakia

Betel quid chewing with or without tobacco has been associated with leukoplakia, a precancerous lesion, as reported in case series, case-control, cross-sectional and cohort studies (IARC, 2004). A case control study from Kerala (Jacob *et al.*, 2004), reported an OR of 4.0 for chewers of betel quid without tobacco, and OR of 12.8 (1.6–101.2) for chewers of areca nut by itself, that may include lime. Both the ORs were adjusted for smoking (Table 3). The trends for both frequency and duration were significant ($p < 0.0001$). The OR for chewers of BQ with tobacco was 10.0 (8.3–12.0) and that for tobacco only was 30.9 (13.7–69.7). The study also showed an OR of 12.5 (3.70–42.4) for erythroplakia, a rarer lesion.

Oral Cancer and Other Head and Neck Cancers

Eight case control studies on oral and other

head and neck cancers, in India are summarised in Table 4, conducted in Madhya Pradesh (Dikshit and Kanhere, 2000), Maharashtra (Jussawala and Deshpande, 1971; Wasnik *et al.*, 1998), and southern Indian states of Kerala, Karnataka and Tamil Nadu (Balaram *et al.*, 2002; Mahapatra, 2015; Muwonge *et al.*, 2008; Nandakumar *et al.*, 1990; Znaor *et al.*, 2003), with two studies being multi-centric, and two studies in men only. Five of the studies adjusted for tobacco smoking, one also for oral dip products (smokeless tobacco) and four adjusted for alcohol.

Six of the studies showed elevated ORs for cancer and chewing of betel quid without tobacco. In the two smallest studies, the ORs were not significant (Dikshit and Kanhere, 2000; Nandakumar *et al.*, 1990). The study from Trivandrum, Kerala, reported an elevated and significant OR on chewing of areca nut

Table 4. Areca nut chewing practices ad risk of oral and other head and neck cancers in case control studies in India.

Sex / Location/	Cancers	Chewing status	Cases	Controls#	Odds ratio (CI) for cancer	Study type, Matching, Adjustments, & Stratifications	References
BOTH MEN & WOMEN							
Mumbai, Maharashtra	Oral cavity, pharynx, esophagus, and larynx: ICD 9 codes 140–148, 150, 161.	Non chewers BQ	129 44	1340 152	1.0 (ref.) 3.0 oral cavity* 1.0 (ref.) 3.0 oropharynx*	Population based. Matched on age, sex, and religion. Not adjusted.	(Jussawala and Deshpande, 1971)
Bangalore, Karnataka	Oral cancer: ICD 9 sites for lip, tongue (excluding base of the tongue), alveolus, and mouth	Never chewers BQ	87 24	233 45	1.0 (ref.) 1.7 (0.9–3.5) NS	Hospital based. Matched on age, sex, and area of residence. Adjusted for smoking.	(Nandakumar <i>et al.</i> , 1990)
Nagpur, Maharashtra	Oro-pharyngeal cancers, ICD 9 codes not specified.	Non chewers Areca nut BQ	33 5 7	185 14 18	1.0 (ref.) 2.6 (0.9–7.7) NS 2.8 (1.1–7.4)	Hospital based. Matched on age and sex. Univariate.	(Wasnik <i>et al.</i> , 1998) Significant trends for frequency and duration of use.
Trivandrum, Kerala	Oral cancer: ICD 10 codes C001–C009	Never chewers BQ	80 13	915 44	1.0 (ref.) 3.5 (1.7–7.1)	Population based. Matched on age & sex. Adjusted for education, religion, smoking and alcohol drinking.	(Mwonge <i>et al.</i> , 2008). Significant trends for frequency and duration of use.
Manipal, Karnataka	Oral cancer: ICD 10 codes Not specified	Supari : No Yes BQ (T): No Yes Incorporation of tobacco not specified	114 20 110 24	261 7 257 17	1.0 (ref) 11.4 (3.4–38.2) 1.0 (ref) 6.4 (2.6–15.5)	Hospital based. Unmatched. Adjusted for age, sex, social class, education level, diet, other tobacco types, dip products, and alcohol.	(Mahapatra, <i>et al.</i> , 2015) No analysis for trends.

Contd...

Table 4. Areca nut chewing practices ad risk of oral and other head and neck cancers in case control studies in India (Contd...)

Sex/ Location	Cancers	Chewing status	Cases	Controls#	Odds ratio (Adj.) & CI for oral cancer	Study type, Matching, Adjustments, & Stratifications	Reference
MEN							
Bangalore, Karnataka	Oral cancer: ICD 9 sites for lip, tongue (excluding base of the tongue), alveolus, and mouth	Never chewers BQ	68 15	89 15	1.0 (ref.) 1.5 (0.6–3.8) NS	Hospital based. Matched on age, sex, and area of residence. Adjusted for smoking.	(Nandakumar <i>et al.</i> , 1990) Significant trends for frequency and duration of chewing in general.
Bhopal, Madhya Pradesh	Oral cavity & oropharynx: ICD 9 codes 140, 141, 143– 145, 146–149	Non chewers BQ	28 4	140 12	1.0 (ref.) 1.7 (0.9–3.3)NS	Population based. Men only. Matched for age. Adjusted for age and smoking.	(Dikshit and Kanhere, 2000) Significant trends for frequency and duration of use
Chennai, Tamil Nadu; Bangalore, Karnataka; Trivandrum, Kerala	Oral cavity: ICD 9 codes not specified	Never chewers BQ	127 15	232 6	1.0 (ref.) 4.2 (1.5–11.8)	Hospital based, at 3 centers. Matched on center, age and sex. Adjusted for age, center, education, smoking and drinking.	(Balaram <i>et al.</i> , 2002) Significant trend for frequency per day
Chennai, Tamil Nadu; Trivandrum, Kerala	Oral cavity: ICD 9 codes 140, 141, 143–145	Non chewers BQ	122 24	1471 83	1.0 (ref) 3.4 (2.0–5.7)	Hospital based at 2 centers. Men only. Stratified: only non- smokers and non-drinkers. Adjusted for age, center, education	(Znaor <i>et al.</i> , 2003) Significant trends for frequency and duration of use.
Trivandrum, Kerala	Oral cavity: ICD 10 codes C001–C009	Never chewers BQ	64 5	561 16	1.0 (ref.) 3.3 (0.9–12.0)	Population based. Matched on age & sex. Adjusted for education, religion, smoking and alcohol drinking (never vs ever).	(Muwonge <i>et al.</i> , 2008) Significant trends for frequency and duration of use.

Contd...

Table 4. Areca nut chewing practices ad risk of oral and other head and neck cancers in case control studies in India (Contd...)

Sex / Location	Chewing status	Cases	Controls#	Odds ratio for oral cancer	Study type. Matching, Adjustments, & Stratifications	Reference
WOMEN						
Bangalore, Karnataka	Never Chewers	19	144	1.0 (ref.)	Hospital based.	(Nandakumar <i>et al.</i> , 1990) Significant trends for frequency and duration of chewing in general.
	BQ	9	30	2.2 (0.7–6.5)	Matched on age, sex and area of residence. Hospital based. Adjusted for smoking.	
Chennai, Bangalore & Trivandrum	Never Chewers	29	251	1.0 (ref.)	Hospital based. Matched on center, age and sex.	(Balaram <i>et al.</i> , 2002) Significant trends for frequency. Starting chewing at an early age has higher risk only for women.
	BQ	14	5	16.4 (4.8–56.5)	Adjusted for age, center, education, smoking and drinking.	
Trivandrum, Kerala	Never chewers	16	354	1.0 (ref.)	Population based. Matched on age & sex. Adjusted for education, religion, smoking and alcohol drinking (never vs. ever)	(Muwonge <i>et al.</i> , 2008) Significant trends for frequency and duration of use.
	BQ	8	28	5.4 (2.1–14.1)		

CI= confidence intervals. BQ=Betel quid without tobacco. Note: Use of lime may be generally inferred even if not mentioned.

BQ= Betel quid; T=tobacco; AN= Areca nut; NS=Not significant; #Controls, from voters' list; † Some smokers among chewers.

* $p < 0.001$

without tobacco for men and women combined (Muwonge *et al.*, 2008). One study reported an OR of 11.4 for *supari* (areca nut) chewing for men and women combined (Mahapatra *et al.*, 2015).

All eight studies had significantly elevated ORs for cancer for chewing of betel quid with tobacco. Trends for frequency were analysed in all but two studies and were significant. Trends for duration analysed in all but three studies and were significant. It is notable that in one study the OR for cancer for past users of any type of betel quid was 11.9 (7.0–20.4), higher than for current users, 4.3 (3.1–6.1) (for men and women combined) (Muwonge *et al.*, 2008), suggesting an accumulation of risk over time before the users quit.

For betel quid with tobacco (BQT), available ORs for men and women combined ranged from 4.8 to 14.6 (Jussawala and Deshpande, 1971; Nandakumar *et al.*, 1990); for men only ORs ranged from 3.4 to 9.3 (Muwonge *et al.*, 2008; Znaor *et al.*, 2003) and for women only ORs ranged from 30.4 to 45.9 (Nandakumar *et al.* 1990; Balaram *et al.*, 2002), all significant. All of the studies, but one, were matched on age and sex (Table 4). Five studies were adjusted

for smoking, and three for alcohol drinking; one study was stratified for smoking and drinking and was of high significance (Znaor *et al.*, 2003).

ORs for areca nut, lime and tobacco use without betel leaf, for men and women combined ranged from a non-significantly elevated 2.4 to a significant 10.2 (Muwonge *et al.*, 2008; Wasnik *et al.*, 1998). For women, the only available OR for areca nut, lime and tobacco was 9.1 (Muwonge *et al.*, 2008). An OR for *gutka* for men and women combined was 5.1 and highly significant (Mahapatra *et al.*, 2015).

Animal Experiments

Studies in animals carried out to investigate the carcinogenicity of areca nut, its constituents and its products and have helped to validate the results of epidemiological studies. Two sets of studies with areca nut (Table 5) and with *pan masala* (Table 6) are reviewed in the following section.

Areca nut studies

Three different animal experiments were designed for simultaneous testing of the carcinogenicity of areca nut, in 2–3 months old inbred Swiss mice (n = 65),

Table 5. Selected animal experiments on the carcinogenicity of areca nut.

Animals	Treatment	Route of administration	Frequency	Durations	Cancerous and other changes observed	Authors, Year, (Country), Notes
25 control Swiss Mice (13M+12F) Aged 2-3 months	Group 1: Pure distilled water	Subcutaneous injections	Once weekly	10 weeks Then allowed to live their full life span up to 27 months	0/25; No local tumour	Ranadive et al., 1976, (India)
40 Experimental Swiss Mice (10 Males & 10 Females) Aged 2-3 months	Aqueous extracts of areca nut: Group 1: cold aqueous extract Group 2: hot aqueous extract	Subcutaneous injections	Once weekly	Life span up to 27 months	Fibrosarcomas at the site of injection: (first after 8 months): Group 1 (cold) : 10/20 Group 2 (hot) : 14/20 (Equal numbers in both sexes)	
30 control Golden Syrian hamsters Aged 2-3 months; Sex not recorded	Untreated	Untreated	NA	Killed in two age groups: 6-12 and 13-21 months (e.g. duration from about 3-9 and 10-18 months)	Cheek pouch: 0 atypia 0 precancers 0 cancer Fore-stomach: 0 atypia, 1 precancer 0 cancer 0 glandular stomach ulcerations (GSU)	Ranadive et al., 1979, SEE P 152 IARC (India) Results for the two age groups are combined
21 Golden Syrian hamsters; Aged 2-3 months	Aqueous extract of areca nut	Cheek pouches painted inside	Tri-weekly	Killed in two age groups: 6-12 and 13-21 months	Cheek pouch: 12 atypia, 2 precancers 1 cancer Fore-stomach: 5 atypia 6 precancers, 4 cancers 5 GSU	
20 Golden Syrian hamsters; Aged 2-3 months	Polyphenol fraction of areca nut (aqueous)	Cheek pouches painted inside	Tri-weekly	Killed in two age groups: 6-12 and 13-21 months	Cheek pouch: 13 atypia 6 precancers 1 cancer Fore-stomach: 6 atypia 2 precancers 4 cancers 5 GSU	

Contd...

Table 5. Selected animal experiments on the carcinogenicity of areca nut. (Contd...)

Animals	Treatment	Route of administration	Frequency	Durations	Cancerous and other changes observed	Authors, Year, (Country), Notes
20 Golden Syrian hamsters; Aged 2–3 months	Whole betel quid aqueous extract	Cheek pouches painted inside	Tri-weekly	Killed in two age groups: 6–12 and 13–21 months	Cheek pouch: 11 atypia 1 precancers 0 cancers	Fore-stomach: 2 atypia 5 precancers 6 cancers 4 GSU Ranadive et al., 1979, SEE P 152 IARC (India)
13 Golden Syrian hamsters; Aged 2–3 months	Areca nut pieces (to induce rauma) + aqueous extract of areca nut	Insertion into cheek pouch followed by painting		Killed in two age groups: 6–12 and 13–21 months	Cheek pouch: 8 atypia 2 precancer 0 cancer	Fore-stomach: 1 atypia 2 precancers 6 cancers 3 GSU Results for the two age groups are combined
27 Golden Syrian hamsters; Aged 2–3 months	Market processed suparis (pieces) of two brands (showing combined results)	Insertion into cheek pouch		Killed in two age groups: 6–12 and 13–21 months	Cheek pouch: 5 atypia 8 precancers 5 cancers	Fore-stomach: 5 atypia 5 precancers 10 cancers 4 GSU
25 Golden Syrian hamsters; Aged 2–3 months	Wax pellet control	Insertion into cheek pouch	Assume every fortnight	Killed in two age groups: 6–12 and 13–21 months	No cancerous changes/lesions	
18 Golden Syrian hamsters; Aged 2–3 months	Wax Pellets containing Betel quid	Insertion into cheek pouch	Every fortnight	Killed in two age groups: 6–12 and 13–21 months	Cheek pouch: 3 atypia 1 precancers 4 cancers	Fore-stomach: 3 atypia 1 precancers 8 cancers 4 GSU

Contd...

Table 5. Selected animal experiments on the carcinogenicity of areca nut. (Contd...)

Animals	Treatment	Route of administration	Frequency	Durations	Cancerous and other changes observed	Authors, Year, (Country), Notes
9 Golden Syrian hamsters; Aged 2–3 months	Gelatine capsule control	Insertion into cheek pouch	Assume every fortnight	Killed in two age groups: 6–12 and 13–21 months	No cancerous changes/lesions	Ranadive et al., 1979, SEE P 152 IARC (India)
19 Golden Syrian hamsters; Aged 2–3 months	Gelatine capsules containing Areca Nut powder	Insertion into cheek pouch	Every fortnight	Killed in two age groups: 6–12 and 13–21 months	Cheek pouch: 5 atypia, 7 precancer, 4 cancers Fore-stomach: 4 atypia, 1 precancer, 6 cancers, 1 GSU	Results for the two age groups are combined
15 Golden Syrian hamsters; Aged 2–3 months	DMBA wax pellet Standard carcinogen control	Cheek pouch	Assume every fortnight	Killed after 6–12 months	Cheek pouch: 4 atypia, 3 precancer, 12 cancer Fore-stomach: 4 atypia, 1 precancer, 6 cancers, 2 GSU	
20 control albino BALB-C mice	Normal saline solution	Buccal mucosa via pipette	Twice daily; 6 days a week	Group 1: 300 days Group 2: 350 days Group 3: 450 days Group 4: 600 days	None (normal status) Mean body weight at 600 days: 49.3 g ± 4.7 g	Perera et al., 2007, (Sri Lanka)
20 albino BALB-C mice in 4 subgroups of 5 mice each; Aged 12 weeks	Aqueous extracts of areca nut	Buccal mucosa via pipette	Twice daily; 6 days a week	Group 1: 300 days Group 2: 350 days Group 3: 450 days Group 4: 600 days	Cellularity, inflammation and muscle atrophy increased from normal to mild by 300 days and remained so up to 600 days. Compared with controls at 600 days the difference was significant (Wilcoxon statistic 15; P = 0.03) Mean body weight at 600 days: 44.5 g ± 2.8 g	The lower average body weight of the exposed mice compared to controls was noted.

DMBA = 7, 12 - dimethylebenz (a) anthracene.

Table 6: Selected animal experiments on the carcinogenicity of *pan masala*, a processed form of areca nut.

Animals	Treatment	Treatment Location	Frequency	Durations	Changes observed	Authors, Year, (Country), Notes
14 Albino Wistar rats (Controls)	No treatment	---	---	8 months	No significant changes in biopsies of buccal mucosa taken at beginning and end of 8 month study period	Khrame et al., 1991 (India)
21 Albino Wistar rats	Paste made of <i>pan masala</i>	Oral cavity	Alternate days	6 months	Dysplasia in 65% of animals; and thickened & condensed submucosal collagen seen in 88% biopsies	
20 Swiss mice (10 Males; 10 Females) of S/RVCri strain, Aged 6-7 weeks (Controls)	Normal diet	Oral consumption	Daily	For intermediate period: killed at 6, 12 and 18 months	No neoplastic lesions	Bhisey et al., 1999 (India)
40 Swiss mice (30 M; 30 F) of S/RVCri strain, Aged 6-7 weeks	2 dose groups: Powdered <i>pan masala</i> mixed in feed at concentrations of 2.5% or 5%	Oral Consumption (10/gender/dose group)	Daily	For intermediate period: killed at 6, 12 and 18 months	2.5% <i>pan masala</i> group: <ul style="list-style-type: none"> No tumors. Forestomach hyperplasia in 10 out of 60 mice 5.0% <i>pan masala</i> group: <ul style="list-style-type: none"> Adenocarcinoma of the lung in 1 male and 1 female. Forestomach hyperplasia in 5 out of 60 mice 	
108 Swiss mice (54 Males, 54 Females) of S/RVCri strain, Aged 6-7 weeks (Controls)	Normal diet	Oral Consumption (54/gender/dose group)	Daily	For lifetime: killed when moribund or at 24 months	No neoplastic lesions	
216 Swiss mice (108 M; 108 F) of the S/RVCri strain, Aged 6-7 weeks	2 does groups: <i>Pan masala</i> finely powdered, mixed with feed at concentrations of 2.5% & 5% and pelleted. (54 mice of each sex allocated to each dose = 108)	Oral Consumption (54/gender/dose group)	Daily	For lifetime: killed when moribund or at 24 months	2.5% <i>pan masala</i> group: <ul style="list-style-type: none"> 5 malignant lesions 5.0% <i>pan masala</i> group: <ul style="list-style-type: none"> 7 malignant lesions Overall 15 benign lesions and 12 malignant lesions Decrease in survival as shown by log rank test ($p = 0.02$). Lung adenocarcinoma, showed a 2-fold increase in the higher dose group compared to lower dose group.	

NA = Not applicable; DMSO= Dimethyl Sulphoxide; DMBA = 7-1 2-Dimethyl-benz(a)anthracene

C17 mice (n = 78), and golden hamsters (n = 45) (Table 5) (Ranadive *et al.*, 1976). Hot and cold aqueous solutions of areca nut were injected subcutaneously in Swiss mice, once a week. Control groups of animals were treated with distilled water. In C17 mice and golden hamsters, dimethyl sulfoxide (DMSO) solutions were used with the aim of enhancing the dermal absorption of the areca nut components from the extract. DMSO areca nut solutions were applied on the skin of the backs of the C17 mice thrice weekly between the shoulder blades. Control groups of animals were treated with 100% DMSO. The hamsters received DMSO areca nut solutions, painted inside the cheek pouch three times a week.

By the end of the lifespan of the Swiss mice (≤ 27 months), ten of 20 mice subcutaneously injected with cold water areca nut extract developed transplantable fibrosarcomas (50%) at the site of injection and 14 of 20 injected with hot water areca nut extract developed fibrosarcomas (Table 5). Tumours were not observed in the internal organs of control and experimental Swiss mice. Skin applications of DMSO areca nut extracts in C17 mice up to 27 months resulted in some mild to moderate hyperplasia but no

skin lesions. Cheek pouches of golden hamsters painted with DMSO extract of areca nut showed some early malignant changes (atypia) up to 24 months. The authors concluded that areca nut demonstrated a carcinogenic principle using aqueous extracts (Ranadive *et al.*, 1976).

With the insights gained, a subsequent set of experiments was conducted in Golden Syrian hamsters. Hamster cheek pouches were painted with aqueous areca nut or betel quid extracts or distilled water for controls. Besides, either wax pellets or gelatin capsules containing betel quid or areca nut powder, or pieces of commercially processed supari were inserted into the cheek pouches and compared to controls with distilled water filled wax pellets or empty gelatine capsules on a triweekly basis. In contrast to the control groups, all treated groups developed numerous malignant changes and cancers (Table 5), a majority occurring in the forestomach (Ranadive *et al.*, 1979).

In a recent study in Sri Lanka, 20 BALB-C mice treated with aqueous extract of fresh areca nut for a maximum of 600 days, developed OSF-like condition in the buccal mucosa with 20 mice treated with normal saline solution as control

groups (Table 5). The changes observed in oral tissues of the mice included proliferation of fibroblasts (increased cellularity), abundance of collagen fibres, increased thickness of the *lamina propria*, infiltration of inflammatory cells (mainly lymphocytes and plasma cells) in the connective tissue, and atrophic epithelium and muscle atrophy in the submucosal layer. These changes closely resembled the human oral mucosa affected with OSF (Perera *et al.*, 2007).

Pan masala

The histopathological changes due to *pan masala* were depicted in a study on painting a paste of a well-known brand of *pan masala* in the oral cavity of 21 albino rats on alternate days for six months. Mild to moderate loss of nuclear polarity and increased keratosis and parakeratosis, inflammatory cell infiltration and vascularity were observed (Table 6). Nearly eight out of nine biopsies showed thickened and condensed sub-mucosal collagen. Thus, histopathological changes observed were similar to OSF in humans (Khrime *et al.*, 1991). Further, carcinogenicity of *pan masala* was studied in six

groups each of 54 Swiss mice (three groups of males and three groups of females, 6–7 weeks of age). The mice were fed diet containing either dry finely powdered *pan masala* (2.5% or 5%), or normal diet (control group) either for life or an intermediate period. The animals were sacrificed when moribund or after 24 months, whichever was earlier. In the intermediate period group, no tumours were seen in the group fed with 2.5% *pan masala*, but two mice in the 5% *pan masala* group developed adenocarcinoma of the lung. In the lifetime group, a total of 15 benign and 12 malignant tumours were observed in the treated mice, while no tumours were found in controls. Most of the malignant tumours occurred in the liver (n = 13), lung (n = 8) and stomach (n = 3). The most common lung neoplasm was lung adenocarcinoma. The mice fed *pan masala* also lost weight after six months and lived a significantly shorter life span compared to the control mice. Thus, the authors have demonstrated evidence of the carcinogenicity of *pan masala* in different mouse tissues, indicating that *pan masala* should be considered a potential human carcinogen (Bhisey *et al.*, 1999).

Mechanistic Evidence of Carcinogenicity

The causal biochemical and molecular mechanisms of oral submucous fibrosis and oral cancer in areca nut chewers are broadly summarized here. During chewing, certain areca nut components, including the alkaloids (mainly arecoline and arecaidine) and polyphenols (tannins, flavonols and catechins) are absorbed through the oral mucosa into the tissues and blood stream (IARC, 2004). These components promote simultaneous abnormal changes in the two main layers of the oral mucosa. A role of in areca nut metabolites in stimulating collagen synthesis in oral mucosa was suggested by tissue culture studies on human fibroblasts from the oral mucosa (Canniff and Harvey, 1981; Harvey *et al.*, 1986; Murti *et al.*, 1995). In the presence of slaked lime (aqueous calcium hydroxide), arecoline, the principal alkaloid, is hydrolysed into arecaidine resulting in irritation and induction of inflammatory mediators (Feller *et al.*, 2013), followed by inflammation. This inflammation stimulates fibroblast proliferation in the *lamina propria*, the connective tissue layer of the mucosa. The stimulated fibroblasts then synthesize excess collagen fibres,

resulting in dense fibrosis, leading to stiffening of the mucosa and eventually to palpable fibrous bands. The increasing atrophy of the overlying epithelium, leads to a burning sensation, impaired vasculature and ulcerations (Angadi and Rao, 2011; Khan *et al.*, 2012). Leukoplakia caused by areca nut may cause atrophy (Borle, 2014). Impaired vasculature is initially responsible for the whitish appearance or blanching of the mucosa due to reduced blood supply, occurring from an early stage of the disease prior to fibrous bands appearance (Ekanayaka and Tilakaratne, 2013). The polyphenols and arecoline react in the presence of slaked lime, forming reactive oxygen species, such as the hydroxyl radical (Nair *et al.*, 1995), resulting in inhibition of collagenase enzymes and phagocytosis, preventing collagen degradation and increasing fibrosis. The high copper content of areca nut participates in promoting fibrogenesis (Angadi and Rao, 2011; Khan *et al.*, 2012).

Genetic damage is observed in the oral mucosa of areca nut chewers. Areca nut-specific nitrosamines, or their precursors, and reactive oxygen species generated in the saliva during betel quid chewing are implicated in causing various forms of

genetic damage in the keratinocytes of the basal layer. The copper content promotes formation of cross linkages between the fibrous bands (Angadi and Rao, 2011; Khan *et al.*, 2012). Betel leaf contains substances, including beta carotene that functions as scavenger of reactive oxygen species and help prevents DNA breakage, thus lowering the risk of cancer among *pan* chewers, compared to those who chew areca nut or its products without betel leaf (Jeng *et al.*, 2002). Genetic damage is indicated by micronucleated cells in the exfoliated oral epithelial cells of chewers of areca nut products and OSF patients (Desai *et al.*, 1996). Micronucleated cells in chewers are in excess ($p < 0.0001$) of those in non-chewers (Joshi *et al.*, 2011). Further genetic alterations in the keratinocytes followed by increased proliferation may lead to malignant phenotypes. A higher percentage of cells with karyolysis (dissolution of chromatin or nuclear contents) has demonstrated in OSF ($p < 0.05$) compared to non-chewers (Joshi *et al.*, 2011). Interactions between the fibroblasts and the keratinocytes appear to promote malignant transformation in OSF (Ekanayaka *et al.*, 2013).

Nitrosation of the areca nut alkaloids

occurs in saliva in the presence of bacterial enzymes, particularly in individuals with poor oral hygiene. The resulting areca nut specific nitrosamines are mutagenic and form DNA adducts in experimental systems, indicating cancer risk (IARC, 2004). Aflatoxins, in areca nut due to fungus infection, form DNA adducts (IARC, 2004). The various genetic lesions (adducts, breaks, etc.) that form with the use of areca nut may progress to cancer over longer time periods (Shah *et al.*, 2012).

DISCUSSION

There is convincing evidence that betel quid or areca nut chewing without tobacco is a cause of oral cancer. A meta-analysis of case-control studies over the last 50 years, on oral/oropharyngeal cancers concluded that overall estimate of relative risk (RR) for use of betel quid without tobacco in the Indian subcontinent was 2.6 (95%CI: 2.0–3.3) (Guha *et al.*, 2014). The frequency of use per day was a more important factor than duration of the habit was unequivocally shown in OSF. The chewing of betel quid containing tobacco confers a greater risk than chewing betel quid without tobacco, besides the added carcinogenicity of tobacco.

Comparing ORs for use of different products and duration of use showed significant differences in risk for OSF. The betel quid chewers were diagnosed after 6–10 years of chewing, whereas *pan masala* and or *gutka* chewers presented with OSF after 2–3 years of use. Thus, chewing of *pan masala* and/or *gutka* causes progression to OSF faster than betel quid. The possible reasons considered were absence of betel leaf and higher consumption by weight of areca nut (Babu *et al.*, 1996).

In the Mumbai Cohort Study the RRs for mortality due to oral and pharyngeal cancers for areca nut or betel quid chewing without or with tobacco did not show significantly elevated RRs, although an RR was significant for other forms of smokeless tobacco use (Gupta *et al.*, 2005). The analysis of incident cancers in the Mumbai Cohort Study (Pednekar *et al.*, 2011), RRs reported for all cancers combined were elevated but not significantly for use of betel quid or areca nut; while RR for cancer of the oral cavity and pharynx for all smokeless tobacco use combined was significant (RR 1.48, 95% CI: 1.03–2.13). These results may in part be due to the rare use of areca nut and tobacco without betel leaf, protective

effect of betel leaf among betel quid users, and number of person years in the cohort yielding a small number of cancer cases during the study period. In contrast, in case-control studies, cancer patients come to specialised treatment centres from very wide geographical areas, home to very large populations.

Risk estimates for precancerous lesions and cancer among the exposed are significantly elevated in case control studies, showing strength of association and a temporal relationship. Most case control studies on OSF or cancer show a dose response relationship with higher frequency per day and greater duration of use. The observed changes in exposed animals and humans fit broadly within known pathways for carcinogenesis, including chronic inflammation and genetic damage, showing plausibility and coherence of findings. Changes in the cheek mucosa occur where the quid is kept by areca nut chewers, implying a direct association. OSF shows specificity to areca nut use, almost always preceding mouth cancer in areca nut users. Alternate explanations, such as the consumption of chillies alcohol or tobacco are not causally related to OSF or cancer. Data showing a positive correlation in OSF and current

users of only tobacco are not verified and past use of areca nut not known. Thus the evidence described in this review is abundantly clear and unequivocally fits the Bradford Hill criteria for causality (Hill, 1965). Policy decisions by the Indian government to control the use of areca nut for the benefit of public health are the need of the hour.

The increasing prevalence of use of areca nut products containing tobacco such as *gutka*, *mawa*, and *pan masala* coincides with rise in OSF and oral cancer primarily at the site of placement in the buccal mucosa. Hospitals in India have noticed increase in admissions for OSF and oral cancer from patients using areca nut products. Thus, convincing evidence on the carcinogenicity of areca nut and tobacco, common use and consequent hazards are obvious in the Indian context. Besides, it is alarming that areca nut products are increasingly exported (40 countries and more), with official quantities of export tripling since 1991 (Kammardi *et al.*, 2012). While tobacco

has been widely recognised as a carcinogen, carcinogenicity of areca nut has not been widely communicated or acknowledged. It is mandatory to dispel ignorance of the hazards of areca nut and recognize the importance of increasing awareness of the carcinogenic potential of areca nut.

CONCLUSIONS

In view of the elevated risk of cancer posed by use of areca nut and the rising incidence of OSF and oral cancer in India, control of areca nut and its products, through banning, is justified in order to contain the adverse health effects on the population and improve public health in the affected individuals. In addition, appropriate communications programmes on the harmfulness of areca nut are strongly recommended.

CONFLICT OF INTEREST

The authors acknowledge no conflict of interest.

REFERENCES

- Ahmad MS, Ali SA, Ali AS, Chaubey KK. Epidemiological and etiological study of oral submucous fibrosis among gutkha chewers of Patna, Bihar, India. *J Indian Soc Pedod Prev Dent* 2006;24(2):84–89.
- Angadi PV, Rao SS. Areca nut in pathogenesis of oral submucous fibrosis: revisited. *Oral Maxillofac Surg* 2011;15(1):1–9.
- Areca catechu L. Indian Medicinal Plants Nomenclature Database. FRLHT.org

- [homepage on the internet]; Bangalore: FRLHT's ENVIS Centre on Medicinal Plants, Last Updated on: 3 February 2015. Available from: http://envis.frlht.org/trade_search.php?txtpart=Not+recorded&lst_part=Not+recorded&txtrade=&lst_trade=ADAKKA+PANJAYADAKKA
- Babu S, Bhat RV, Kumar PU, Sesikaran B, Rao KV, Aruna P, *et al.* A comparative clinico-pathological study of oral submucous fibrosis in habitual chewers of *pan masala* and betelquid. *J Toxicol Clin Toxicol* 1996;34:317–322.
- Balaram P, Sridhar H, Rajkumar T, Vaccarella S, Herrero R, Nandakumar A, *et al.* Oral cancer in southern India: the influence of smoking, drinking, paan-chewing and oral hygiene. *Int J Cancer* 2002;98(3):440–445.
- Bathi R, Parveen S, Burde K. The role of gutka chewing in oral submucous fibrosis: A case-control study. *Quintessence Int* 2009;40(6):e19–25.
- Bhisey RA, Ramchandani AG, D'Souza AV, Borges AM, Notani, PN. Long-term carcinogenicity of *pan masala* in Swiss mice. *Int J Cancer* 1999;83:679–684.
- Borle, Rajiv M. *Textbook of Oral and Maxillofacial Surgery*. New Delhi: Jaypee Brothers Medical Publishers Ltd. 2014;672.
- Canniff JP, Harvey W. The aetiology of oral submucous fibrosis: the stimulation of collagen synthesis by extracts of areca nut. *Int J Oral Surg* 1981;10(Suppl):163–167.
- Chaturvedi P, Chaturvedi U, Sanyai B. Prevalence of tobacco consumption in school children in rural India – an epidemic of tobacco-associated cancers looming ahead in the third world. *J Cancer Educ* 2002;17:6. Letter to the editor.
- Chaudhry K. Is *pan masala*-containing tobacco carcinogenic? *Natl Med J India* 1999;12(1):21–27.
- Desai SS, Ghaisas SD, Jakhi SD, Bhide SV. Cytogenetic damage in exfoliated oral mucosal cells and circulating lymphocytes of patients suffering from precancerous oral lesions. *Cancer Lett* 1996;109:9–14.
- Dikshit RP, Kanhere S: Tobacco habits and risk of lung, oropharyngeal and oral cavity cancer: a population-based case-control study in Bhopal, India. *Int J Epidemiol* 2000;29:609–614.
- Ekanayaka RP, Tilakaratne WM. Oral Submucous Fibrosis: Review on Mechanisms of Pathogenesis and Malignant Transformation. *J Carcinogene Mutagene* 2013;S5:002.
- Feller L, Altini M, Lemmer J. Inflammation in the context of oral cancer. *Oral Oncol* 2013;49(9):887–892.
- Guha N, Warnakulasuriya S, Vlaanderen J, Straif K. Betel quid chewing and the risk of oral and oropharyngeal cancers: a meta-analysis with implications for cancer control. *Int J Cancer* 2014;135(6):1433–1443.
- Gupta PC, Hebert JR, Bhonsle RB, Sinor PN, Mehta H, Mehta FS. Dietary factors in oral leukoplakia and submucous fibrosis in a population-based case control study in Gujarat, India. *Oral Dis* 1998;4(3):200–206.
- Gupta PC, Pednekar MS, Parkin DM, Sankaranarayanan R. A cohort study of 99,570 individuals in Mumbai, India for tobacco-associated mortality. *Int J Epidemiol* 2005;34(6):1395–1402.
- Gupta PC, Ray CS, Murti PR, Sinha DN. Rising

- incidence of oral cancer in Ahmedabad city. *Indian J Cancer* 2014;51(Suppl 1):S67–72.
- Gupta PC, Warnakulasuriya S. Global epidemiology of areca nut usage. *Addict Biol* 2002;7:77–83.
- Gupta PC. Mouth cancer in India: a new epidemic? *J Indian Med Assoc.* 1999;97(9):370–373.
- Harvey W, Scutt A, Meghji S, Cannif JP. Stimulation of human buccal mucosa fibroblasts in vitro by betel-nut alkaloids. *Arch Oral Biol* 1986;31:45–49.
- Hazare VK, Goel RR, Gupta PC. Oral submucous fibrosis, areca nut and *pan masala* use: A case control study. *Nat Med J India* 1998;11(6):299.
- Hazare VK, Erlewad DM, Mundhe KA, Ughade SN. Oral submucous fibrosis: study of 1000 cases from central India. *J Oral Pathol Med* 2007;36(1):12–17.
- Hill AB. *The Environment and Disease: Association or Causation?* Proceedings of the Royal Society of Medicine. 1965;58:295–300. Available from: http://www.drabruzzi.com/hills_criteria_of_causation.htm (accessed on 7 April, 2015)
- International Agency for Research on Cancer, Monographs on the evaluation of carcinogenic risks to humans. *Betel-quid and areca-nut chewing and some areca-nut-derived nitrosamines*. Vol 85. Lyon: IARC 2004; 44,45,140–142,160–167,227–229.
- International Agency for Research on Cancer, Monographs on the evaluation of carcinogenic risks to humans. *Personal habits and indoor combustions*. Vol. 100E. *Betel quid and areca nut*. Lyon: IARC 2009;337–372.
- International Institute for Population Sciences and Ministry of Health and Family Welfare, Government of India. *Global Adult Tobacco Survey (GATS) India, 2009–10*. New Delhi: Ministry of Health and Family Welfare (MOHFW), Government of India; Indian Institute for Population Sciences (IIPS); 2010. Available from: <http://mohfw.nic.in/WriteReadData/1892s/1455618937GATS%20India.pdf> (accessed on 15 January, 2015)
- Jacob BJ, Straif K, Thomas G, Ramadas K, Mathew B, Zhang ZF, *et al.* Betel quid without tobacco as a risk factor for oral precancers. *Oral Oncol* 2004;40(7):697–704.
- Jayalekshmi PA, Gangadharan P, Akiba S, Koriyama C, Nair RR. Oral cavity cancer risk in relation to tobacco chewing and bidi smoking among men in Karunagappally, Kerala, India: Karunagappally cohort study. *Cancer Sci.* 2011;102(2):460–467.
- Jayalekshmi PA, Gangadharan P, Akiba S, Nair RR, Tsuji M, Rajan B. Tobacco chewing and female oral cavity cancer risk in Karunagappally cohort, India. *Br J Cancer* 2009;100(5):848–852.
- Jeng JH, Chen SY, Liao CH, Tung YY, Lin BR, Hahn LJ, Chang MC. Modulation of platelet aggregation by areca nut and betel leaf ingredients: roles of reactive oxygen species and cyclooxygenase. *Free Radic Biol Med* 2002;32(9):860–871.
- Joshi MS, Verma Y, Gautam AK, Parmar G, Lakkad BC, Kumar S. Cytogenetic alterations in buccal mucosa cells of chewers of areca nut and tobacco. *Arch Oral Biol* 2011;56(1):63–67.
- Jussawala DJ, Deshpande VA. Evaluation of cancer risk in tobacco chewers and smokers:

- an epidemiologic assessment. *Cancer* 1971;28(1):244–252.
- Kammardi TNP, Ranganath L, Ashok Kumar HA, Rajkumar BJ, Umesh CP, Ranjith Kumar PS, et al. *Areca Nut Economy at The Crossroads. Special Scheme on Cost of Cultivation Areca nut in Karnataka*. Bengaluru: Department of Agricultural Economics, University of Agricultural Sciences (UAS); 2012. Available from: <http://www.costofcultivationkarnataka.in/downloads/Areca>
- Khan S, Chatra L, Prashanth SK, Veena KM, Rao PK. Pathogenesis of oral submucous fibrosis. *J Cancer Res Ther* 2012;8(2):199–203.
- Khandelwal A, Khandelwal V, Saha MK, Khandelwal S, Prasad S, Saha SG. Prevalence of areca nut chewing in the middle school-going children of Indore, India. *Contemp Clin Dent* 2012;3(2):155–157.
- Khrime RD, Mehra YN, Mann SBS, Mehta SK, Chakraborti RN. Effects of instant preparation of betel nut (*pan masala*) on the oral mucosa of albino rats. *Indian J Med Res* 1991;94:119–124.
- Mahapatra S, Kamath R, Shetty BK, Binu V S. Risk of oral cancer associated with gutka and other tobacco products: A hospital-based case-control study. *J Can Res Ther* [serial online] 2015 [cited 2015 May 11];11:199–203. Available from: <http://www.cancerjournal.net/text.asp?2015/11/1/199/143332>
- Maher R, Lee AJ, Warnakulasuriya KA, Lewis JA, Johnson NW. Role of areca nut in the causation of oral submucous fibrosis: a case-control study in Pakistan. *J Oral Pathol Med* 1994; 23(2):65–69.
- Mehrotra D, Kumar S, Agarwal GG, Asthana A, Kumar S. Odds ratio of risk factors for oral submucous fibrosis in a case control model. *Br J Oral Maxillofac Surg* 2013;51(7):e169–173.
- Mehta FS, Gupta PC, Daftary DK, Pindborg JJ, Choksi SK. An epidemiologic study of oral cancer and precancerous conditions among 101,761 villagers in Maharashtra, India. *Int J Cancer* 1972;10(1):134–141.
- Murti PR, Bhonsle RB, Gupta PC, Daftary DK, Pindborg JJ, Mehta FS. Etiology of oral submucous fibrosis with special reference to the role of areca nut chewing. *J Oral Pathol Med* 1995;24:145–152.
- Muwonge R, Ramadas K, Sankila R, Thara S, Thomas G, Vinoda J, Sankaranarayanan R. Role of tobacco smoking, chewing and alcohol drinking in the risk of oral cancer in Trivandrum, India: a nested case-control design using incident cancer cases. *Oral Oncol* 2008;44:446–454.
- Nair UJ, Nair J, Friesen MD, Bartsch H, Ohshima H. ortho- and meta-Tyrosine formation from phenylalanine in human saliva as a marker of hydroxyl radical generation during betel quid chewing. *Carcinogenesis* 1995;16:1195–1198.
- Nandakumar A, Thimmasetty KT, Sreeramareddy NM, Venugopal TC, Rajanna, Vinutha AT et al. A population-based case-control investigation on cancers of the oral cavity in Bangalore, India. *Br J Cancer* 1990;82:847–851.
- National Centre for Disease Informatics and Research (NCDIR), National Cancer Registry Programme (NCRP). *Time Trends in Cancer Incidence Rates, 1982–2010*. Bangalore: NCDIR-NCRP (ICMR); 2013;316.
- Orr IM, Glasg MB. Oral cancer in betel nut

- chewers in Travancore. Its etiology, Pathology, and treatment. *Lancet* 1933;222(5741):575–580.
- Pednekar MS, Gupta PC, Yeole BB, Hebert JR. Association of Tobacco Habits, including Bidi Smoking, with Overall and Site-Specific Cancer Incidence: Results from the Mumbai Cohort Study. *Cancer Causes and Control* 2011;22(6):859–868.
- Perera MWS, Gunasinghe D, Perera PAJ, Ranasinghe A, Amaratunga P, Warnakulasuriya S, Kaluarachchi K. Development of an in vivo mouse model to study oral submucous fibrosis. *J Oral Pathol Med* 2007;36(5):273–280.
- Ranadive KJ, Gothoskara V, Rao R, Tezabwalla BU, Ambaye RY. Experimental studies on betel nut and tobacco carcinogenicity. *Int J Cancer* 1976;17:469–476.
- Ranadive KJ, Ranadive SN, Shivapurkar NM, Gothoskar SV. Betel quid chewing and oral cancer: experimental studies on hamsters. *Int J Cancer* 1979;24(6):835–843.
- Reddy KS and Gupta PC (eds). *Report on Tobacco Control in India*. Ministry of Health and Family Welfare, Government of India, New Delhi, 2004.
- Shah G, Chaturvedi P, Vaishampayan S. Areca nut as an emerging etiology of oral cancers in India. *Indian J Med Paediatr Oncol* 2012;33(2):71–79.
- Shah N, Sharma PP. Role of chewing and smoking habits in the etiology of oral submucous fibrosis (OSF): a case-control study. *J Oral Pathol Med* 1998;27(10):475–479.
- Sinor PN, Gupta PC, Murti PR, Bhonsle RB, Daftary DK, Mehta FS, *et al*. A case-control study of oral submucous fibrosis with special reference to the etiologic role of areca nut. *J Oral Pathol Med* 1990;19:94–98.
- Wahi PN. The epidemiology of oral and oropharyngeal cancer. A report of the study in Mainpuri District, Uttar Pradesh, India. *Bull World Health Organ* 1968;495–521.
- Wasnik KS, Ughade SN, Zodpey SP, Ingole DL. Tobacco consumption practices and risk of oropharyngeal cancer: a case-control study in central India. *S E Asian J Trop Med Public Health* 1998;29:827–834.
- Winstock AR, Trivedy CR, Warnakulasuriya KAAS, Peters TJ. A dependency syndrome related to areca nut use: Some medical and psychological aspects among areca nut users in the Gujarat community in the UK. *Addict Biol* 2000;5:173–179.
- Znaor A, Brennan P, Gajalakshmi V, Mathew A, Shanta V, Varghese C, *et al*. Independent and combined effects of tobacco smoking, chewing and alcohol drinking on the risk of oral, pharyngeal and esophageal cancers in Indian men. *Int J Cancer* 2003;105(5):681–686.

Identification of Therapeutic Targets for Cancer: Proteomic Technologies and Strategies are the Key to Success

Rukmini B. Govekar

Advanced Centre for Treatment, Research and Education in Cancer (ACTREC), Tata Memorial Centre, Kharghar, Navi Mumbai, India

With the emergence of the field of 'omics' a new era of systematic global profiling of cellular molecules has been initiated in biology. Different 'omics' approaches have been extensively used to identify biomarkers for better diagnosis and prognosis, therapeutic strategies and monitoring response to therapy in diverse types of cancers. Proteomics is the approach of choice for identification of therapeutic targets. This is because therapeutic modulation of expression, post-translational modification and activity of a protein can directly rectify the derangement in the disease-causing cellular pathway. The current review scans literature on tumor proteomics to understand the influence of developments in proteomics technology and study approaches on identification of targets for therapy. Diversity of tumor types, molecular heterogeneity in pathologically indistinguishable tumors provides ample challenge to assess the strength of proteomics in identification of drug targets. The review highlights comparative proteomic profiling by gel-based or gel free approach, in tumor and normal tissues or chemo-resistant/sensitive tumor tissues have identified differentiator proteins, with potential as targets as therapeutic targets. Further, along with evolution in proteomic technologies for identification and quantification of proteins, various tools for functional analysis of proteins have contributed to strategies for target identification. It also suggests that future advances in quantitative, functional and structural proteomics isare necessary to widen the search for therapeutic targets.

INTRODUCTION

The pace of development of technology is a rate determining factor in the rapid progress of basic and applied sciences. This is exemplified in the healthcare sector by the translational advances of the technological inventions in the healthcare sector which include computerized

tomography (CT) and magnetic resonance imaging (MRI) for diagnosis; laparoscopes and cardiopulmonary bypass pumps for surgery; and systems for therapies such as radiotherapy and dialysis. Apart from these technologies which are directly used in clinics,

Key words: Proteomic, Therapeutic target, Cancer, Two dimensional gel electrophoresis, Liquid chromatography, Mass spectrometry.

***Corresponding Author:** Rukmini B Govekar, Advanced Centre for Treatment, Research and Education in Cancer (ACTREC), Tata Memorial Centre, Kharghar, Navi Mumbai, India.

Email: rgovekar@actrec.gov.in

advances in optical and analytical instruments have revolutionized biomedical research contributing to the progress in understanding pathobiology of diseases and their management. The current review focuses on proteomic technology-driven advances in the identification of therapeutic targets in various cancers, as a milestone in development of targeted therapeutics.

Serendipitous discoveries of drugs and drug targets

Identification of novel therapeutic targets remains an area of interest to clinicians and biotechnology and pharmaceutical industries. Several drugs have been discovered earlier by extraction of the active principle from natural sources such as plants traditionally used to treat a disease (Cragg, 2013). A few drugs were discovered by serendipity, a classic example being penicillin from a contaminating fungus (Bennet, 2001) and a cancer chemotherapeutic vincristine by its undesired myelosuppressive effect when used to treat diabetes (Johnson, 1963). With improved understanding of molecular machinery of the cell and aberrations associated with disease conditions, efforts were made to design drugs to target the disease-causing molecule. This approach was favoured for diseases with an established causal association with a molecular alteration. For example, in chronic myeloid leukemia (CML), inhibition of the transforming

tyrosine kinase has been an excellent targeted therapy in CML patients (Freireich, 2014). However, for a long time, serendipity remained the basis for discoveries of drugs as well as for the identification of targets as in CML. Such discoveries require fortuitous co-occurrence of the phenomenon which led to the discovery and an alert analyst. A structured approach is necessary for identification of therapeutic targets and discovery of new drugs.

'Omics' for systematic identification of molecular alterations in tumors

With emergence of 'Omics', biology began a new era of planned and systematic search for global molecular alterations in diseases. The strength of this approach is evident from its potential to unravel key molecular events from the numerous molecular alterations seen in cancer (Bertrand, 2015; Castro-Vega, 2015). Multifactorial and multigenic origin of cancer is reflected in the diverse molecular alterations in pathologically indistinguishable tumors, which progress and respond differentially to drugs and therefore require an individualized therapeutic approach. Interrogation of differences in genome, transcriptome, proteome, metabolome, lipidome have led to the identification of several cancer biomarkers useful in diagnosis (Du, 2014; Liu, 2015), predict disease prognosis (Minca, 2014; Shipitsin, 2014), assist in choice of therapy (Fènichel, 2014; Sjøholt,

2006) and monitor response to therapy (Rebecca, 2014).

Proteomics: The 'Omics' of choice for identification of drug targets

Proteins are functional molecules in a cell and alterations in their function can affect cell phenotype. Proteomics, a global study of proteins, is therefore an approach of choice to identify drug targets. The study of a cell proteome is challenging due to the complexity of protein structure; effect of post-translational processing i.e., cleavage or modification of function, sub-cellular localization and changes in interacting molecules including substrates (Parker, 2014); dynamic range of proteins expressed in a cell (Corthals, 2000) and the temporal variations in the proteins.all the above variables. In a disease state, alterations in any of these parameters may occur in one to several proteins. Challenges posed by these compounding factors to proteomic profiling have been addressed at every step of development of proteomic technology and supported by advances in the fields of genomics (Wang, 2014), bioinformatics (Boguski, 2003) and computational biology (Dowsey, 2003).

Technological advances meet the challenges of profiling complex proteomes

Protein identification by mass spectrometry

Emergence of the field of proteomics began with the changes improvisations in

mass spectrometry (MS) instrumentation and techniques in late 1990s. Proteins and other biomolecules fragmented by the previously used ionization methods, were preserved intact by the softer ionizations – electrospray ionization (ESI) (Karas, 1988) and matrix assisted laser desorption ionization (MALDI) (Fenn, 1989) promoting MS as a tool for biologists. The ionizers were coupled to an analyzer, such as a quadrupole, time of flight, ion trap, etc. which separated the ionized molecules on the basis of their mass/charge (m/z) ratio. MS-based identification of proteins was achieved by comparing the masses of peptides generated by cleavage of a protein using specific protease and those generated 'in silico' by digestion of sequences available in public databases with the same protease, an application called peptide mass fingerprinting (PMF). In tandem mass spectrometry platforms (MS-MS) with more than one analyzer, peptides detected in the first analyzer are put through controlled fragmentation by collision induced dissociation (CID). The accurate masses of the peptide fragments obtained from analysis in the second analyzer, when processed by appropriate software designed generated the sequence of the peptide. The sequence is used for protein identification. Thus mass spectrometry circumvented the need for a probe to detect the protein of interest from a complex mixture of proteins (Abersold, 2003). Further, deeper interrogation of the proteome, essential for biomarker

discovery, was made possible by the features in mass spectrometer which allowed data-independent fragment analysis (Sajic, 2014). Further advancement in the fragmentation mechanisms, introduction of electron transfer dissociation (ETD), improved detectability of labile post-translational modifications in proteins (Kim, 2012). Advances in mass spectrometry therefore improved the detection, identification and knowledge of the post-translational modifications in proteins.

Reduction in protein complexity before mass spectrometry

The strength of mass spectrometry for protein identification is compromised in biological samples due to the complexity of proteome and dynamic range of protein expression. High or medium abundant species in a sample may interfere with the detection of low abundant species, called “suppression effect”. Therefore to enable identification of less abundant proteins and to enrich less concentrated species, clinical proteomics studies require fractionation of proteins from complex mixtures, prior to proceeding for identification by mass spectrometry analysis. The strategy which can optimally reduce the complexity would differ in each sample (Camerini, 2015).

Among gel based separation methods, two dimensional gel electrophoresis (2DGE) has been the method of choice. Separated proteins are subject to in-gel

digestion with specific proteases, peptides are extracted from the gel and subjected to MS for protein identification. The technique of 2DGE has metamorphosed due to availability of immobilized pH gradients (IPG), multi-gel electrophoresis apparatus improving the reproducibility of profiles, and staining protocols using fluorescent dyes with improved sensitivity and linearity over a wider dynamic concentration range. IPG strips are available in micro pH ranges for wider resolution, and thereby improving detection of proteins (Gorg, 2000). Apart from 2DGE, capillary electrophoresis and agarose gel isoelectric focusing are used to separate proteins from biological samples (Manabe, 2000).

Liquid chromatography (LC) is a versatile method for protein separation as different column chemistries allow separation of proteins based on distinct characteristics (Di Palma, 2012). Since ESI can ionize samples introduced in liquid phase, LC-MS platforms were designed, wherein sample separated on LC can be introduced directly into the ionizer of MS. However, as biological samples are obtained in limiting amounts, efforts have been made to increase the sensitivity of detection by tapping measures beyond the improvisations in the hardware of MS. In LC-MS systems, reducing the flow rate contributes to higher overall sensitivities due to the higher efficiencies in ESI and reduced ion suppression effects (Köcher, 2014). Nano LC-MS-MS is the

configuration of choice for biological samples analysis.

Separation and identification tools for differential quantification

Distinction between expression of proteins in disease and normal state requires quantitative evaluation of the expression. Several tools are available to achieve the same. Quantitative proteomics can be categorized into absolute and relative types. Absolute quantitation determines changes in protein expression in terms of an exact amount or concentration of each the protein present; whereas relative quantitation determines the up- or down-regulation of a protein relative to the control sample, generally used in clinical proteomics. In MS based quantitation, the relative concentration can be obtained by: Label free methods are based on less rigorous mass spectrometry, with more reliance on bioinformatics and separation techniques. Chemical labeling is applicable to a wider range of biological samples, and methods such as isotope coded affinity tags (iCAT) and isobaric tags for relative and absolute quantitation (iTRAQ), are favoured in quantitative proteomics. The control and test samples are labeled with separate tags, and intensity of the same ion with distinct tags indicates the relative quantitation (Elliott, 2009). Protein quantitation can be done by two dimensional difference gel electrophoresis (2D-DIGE) wherein proteins from different samples are labeled

using dyes that provide different fluorescence wavelengths for detection. The labeled samples to be quantitated are mixed in equal proportion and separated on the same gel. The gels are scanned and the relative fluorescence of distinct dyes is recorded for quantitation (Timms, 2008). The quantitative differences obtained from the above mentioned studies are the first scan of the differentiators, which are the storehouse of potential biomarkers and therapeutic targets. .

Selection of appropriate proteomic technology

Advocates of gel free and gel-based proteomics favour the method of choice due to certain advantages. Shot-gun proteomics carried out using LC-MS platforms are less laborious, more reproducible and capable of generating a larger profile (Wilkins, 2009). On the other hand, gel-based proteomics (Rogowska-Wrzesinska, 2013) aids identification and sequencing of proteins from organisms with minimal genomic information, efficiently identifies protein isoforms and proteins modified by glycosylation, proteolytic cleavage, etc.

Selection of comparison groups for identification of therapeutic targets

Identification of differentiators has been carried out with different aims and therefore differentiators are derived from diverse comparison groups. However, they inadvertently point at the key

molecules as potential therapeutic targets.

Comparison of tumor and normal in retrospective or prospective studies

For understanding cancer biology, the profile of differentiators are generated to reveal molecular mechanisms responsible for disease progression. Potential markers for progression of pulmonary squamous cell carcinoma were identified by examining samples of lung SCC and adjacent normal tissues using 2D-DIGE (Lihong, 2014). Markers of progression of oral squamous cell carcinoma from premalignant lesion to carcinoma have been similarly demonstrated (Wang, 2009). Using gel free approach, insight into the underlying mechanisms of formation of polyploidy giant cancer cells (PGCC) and the relationship between PGCCs and cancer stem cells in patients with ovarian cancers has been established (Zhang, 2013).

Differential molecular profile is often generated for better stratification of tumors in order to improve diagnosis and management of the disease. Diagnostic markers have been identified by 2D-DIGE for cervical cancer (Canales, 2014; Guo, 2014). Similarly, using gel-based approach it has been shown that high expression level of Galectin-1 may correlate with development of nasopharyngeal carcinoma (NPC), and Galectin-1 as a potential diagnostic marker or therapeutic target for NPC (Tang, 2010). Further, a significant proportion of

primary breast cancers are negative for estrogen receptors (ER), progesterone receptor (PgR), and Her2, comprising the triple negative breast cancer (TNBC) group. Women with TNBC have poor prognosis because of the aggressive nature of the tumors and current lack of suitable targeted therapies. The increased expression of Mage-A4 in the tumors enabled the detection of the protein in the tumor interstitial fluids and in sera. Immunotherapeutics approaches specifically targeted Mage-A4 protein, or Mage-A protein family members represents novel management options for TNBC (Cabezon, 2013).

An alternative aim for profiling of tumors is to predict prognosis. Using gel based proteomics approach, a subgroup of breast tumors with overexpressed C7 or f24 showed poor clinical outcome (Gromov, 2010). Similarly, LC-MS approach identified RBBP6 as prognostic marker for gastric cancer stem cell (Morisaki, 2014) and WD repeat containing protein 1 was identified as a diagnostic marker in the interstitial fluid from ovarian cancer (Haslene-Hox, 2013).

Comparison groups with focus on therapeutics

Several differentiators identified earlier may be useful potential therapeutic targets. Additionally, proteomic analyses aimed at understanding mechanism of drug resistance or drug action have greater probability to identify as therapeutic

targets.

Analysis of cell lines or samples from patients, untreated or treated with a drug, provides insight into the molecular mechanism of action of drugs/chemopreventive agents. Further identification of the drug modulated pathway may indicate therapeutic targets for further exploration. Curcumin, a natural anticancer agent, inhibits cell growth in a number of tumor cell lines and animal models. Molecular mechanism of curcumin induced apoptosis in different gastric cancer cell lines was studied by 2DGE (Cai, 2013). Similarly, using gel free approach, *Bifidobacterium infantis* thymidine kinase/ nucleoside analogue ganciclovir (BI-TK/ GCV) exhibited sustainable anti-tumor growth activity and induced apoptosis in bladder cancer, via peroxiredoxin I and NF κ B pathway (Jiang, 2014).

Comparison of chemo-sensitive and -resistant tumors/cell lines provides information of the molecular basis of resistance and hence molecules to be considered as alternate therapeutic targets. In mantle cell lymphoma, a rare aggressive type of B cell non-Hodgkin's lymphoma, wherein response to chemotherapy tends to be short and patients relapse, the tumor necrosis factor related apoptosis inducing ligand (TRAIL) is a novel molecule with antitumor effects. In TRAIL resistant cases, 2DGE analysis demonstrated downregulation of the key enzymes of purine metabolism with profound effects

on nucleotide homeostasis and can render cells vulnerable to further disruption of purine nucleotide metabolism. Thus proteins in this pathway identified are potential therapeutic targets for selective elimination of resistant cells (Pospisilova, 2013). Chemo-resistance hinders effective treatment in several human cancers. HSP27 is as an alternate target for anticancer drug development in gemcitabine therapy resistant pancreatic cancer (Liu, 2012). Histone deacetylase inhibitors (HDACi) demonstrates anticancer activities and used in combination therapy. In lymphoid cell lines, 2DGE analysis has identified HSPA1A as an overexpression with resistance to valproic acid HDAC inhibitor. *In vitro* experiments demonstrate that treatment with KNK-437, an inhibitor of HSPA1A, enhanced cytotoxic effects of valproic acid, thereby identifying HSPA1A as a possible therapeutic target, in combination with HDACi, for lymphoid neoplasms (Fuji, 2012).

Authentication of the potential of identified therapeutic target

Differentiators have been identified for several cancers, however, differentiators as a therapeutic targets needs further investigations. Bioinformatic tools for pathway identification are extensively used to find a functional link between the differentiators. A molecule in a pathway associated with hallmarks of cancer

(Hanahan, 2011), qualify as potential therapeutic targets. However, the potential needs to be authenticated experimentally. In several studies, over expression or activation as well as down-regulation or inhibition of the identified potential drug target is used to demonstrate the effect on tumor promotion or progression.

Tamoxifen (Tam) is a widely used selective estrogen receptor modulator (SERM) for treatment of hormone-responsive breast cancer and acts via inhibition of E6AP expression identified as a differentiator by 2DGE. Authentication of E6AP as a therapeutic target was achieved by demonstration of Tam- and siE6AP-mediated inhibition of E6AP with subsequent enhanced G0-G1 growth arrest and apoptosis (Lochab, 2012). Small interfering RNA (siRNA)-mediated knockdown confirmed a functional role for MDA-9 and GRP78 in promoting cell invasion in A375 cells (Guan, 2014). Similarly in liposarcoma, an aggressive cancer with poor outcome, gankyrin oncoprotein showed a significantly high expression. Inhibition of gankyrin led to reduction of *in vitro* cell proliferation, colony-formation and migration, besides *in vivo* dedifferentiated liposarcoma tumorigenesis (Hwang, 2014). KvLQT1 channel blockade was showed efficient reduction of A549 and H460 cell proliferation and migration. Moreover, KvLQT1 overexpression in AD samples suggested it to be a potential therapeutic target in lung cancer (Girault,

2014). In an *ex vivo* model, siRNA mediated inhibition of HSP70, showed dose-dependent inhibition of cell growth and burst formation unit erythroid (BFU-E), increased apoptosis in the erythroid lineage and decreased pJAK2 signaling. Thus HSP70 as a potential therapeutic target in myeloproliferative neoplasms especially polycythemia vera was confirmed (Gallardo, 2013). Similarly inhibition of Apg-2 showed decreased cell proliferation and induced apoptosis in BCR/ABL positive cells, indicating an additional therapeutic target for chronic myeloid leukemia (Li, 2013). Whereas, in chronic lymphocytic leukemia (CLL) a clonal malignancy with immense clinical heterogeneity with variable prognosis, hyper reactivity of the B cell receptor (BCR) to unknown antigen ligation plays a pivotal role in CLL survival. Proteomic analysis revealed that kininogen, a critical protein of kinin-kallikrein system, was upregulated upon BCR stimulation and may provide a therapeutic target in CLL (Kashuba, 2013). Further, in MLL-rearranged leukemia, TET1 was identified as a potential therapeutic target (Huang, 2013). Similarly in endometrial cancer, overexpression of bone marrow stromal antigen 2 (BST2) was detected in LC-MS analysis and confirmed by immunohistochemistry using clinical samples. In an *in vivo* xenograft model, BST2 antibody treatment inhibited tumor growth of BST2-positive endometrial cancer cells in an NK cell-dependent manner,

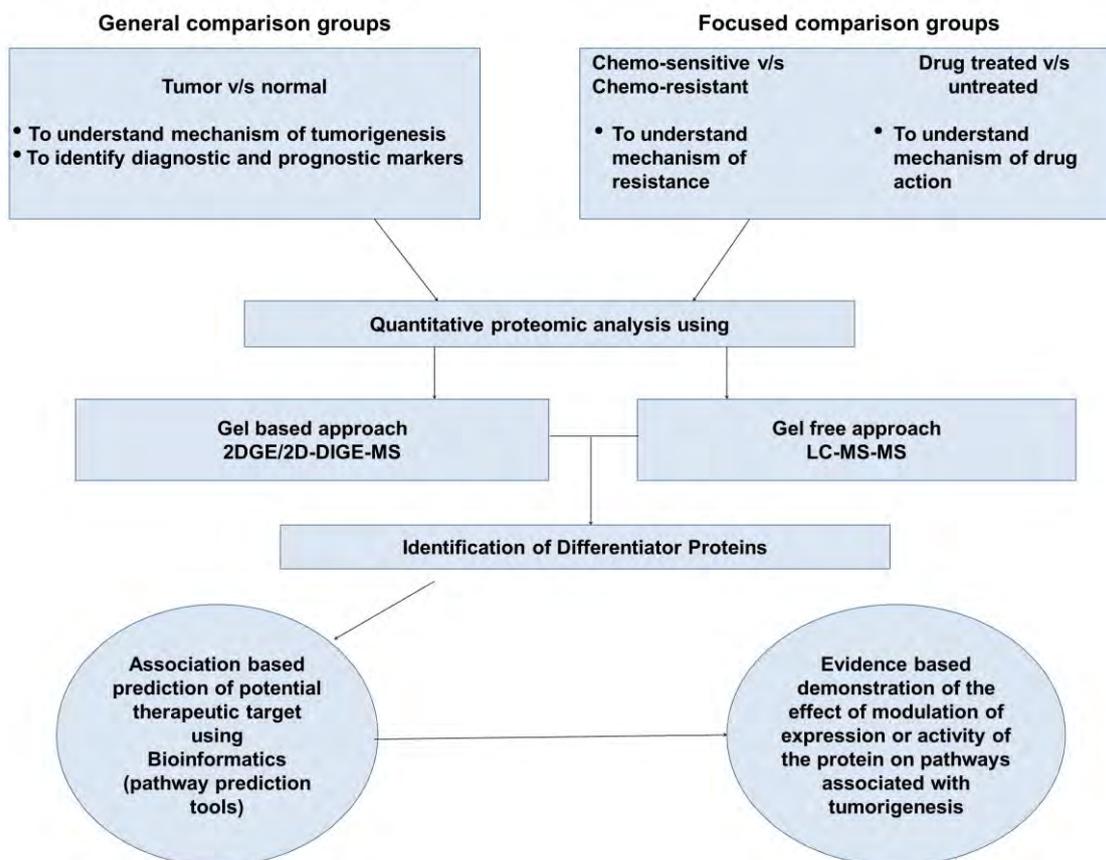


Figure 1: Various strategies and technologies for proteomic identification of therapeutic targets for cancer. (2D-DIGE: two dimensional difference gel electrophoresis; MS- mass spectrometry).

advocating candidacy of BST2 as a therapeutic target (Yokoyama, 2013). These studies provide an initial authentication of the potential of identified therapeutic targets.

To summarize, the review highlights usefulness of proteomic technology in identification of therapeutic targets as outlined in figure 1. The review reveals that differentiators, identified by both gel-based and gel free approaches qualify therapeutic targets. It appears that comparative proteomic analysis of chemosensitive and chemoresistant cells as well as that of drug treated and untreated cells, are useful in identification of

therapeutic targets. The search strategy for therapeutic targets has evolved from association based approaches wherein a differentiator protein with known role in key functional pathways qualified as a potential target. Evidence based selection of therapeutic targets necessitated experimental demonstration of the ability of the differentiator to affect the hallmarks of cancer (Guo, 2013). Thus, we conclude that advances in proteomic technology and refinements in experimental strategies have contributed to identification of therapeutic targets in tumors and in turn to the field of targeted therapeutics.

REFERENCES

- Abersold R. A mass spectrometric journey into protein and proteome research. *J Am Soc Mass Spectrometry*. 2003;14:685–695.
- Bennet J, Chung K. Alexander Fleming and the discovery of penicillin. *Adv Appl Microbiol* 2001;49:163–184.
- Bertrand D, Chng K, Sherbaf F, Kiesel A, Chia B, Sia Y, *et al.* Patient-specific driver gene prediction and risk assessment through integrated network analysis of cancer omics profiles. *Nucleic Acids Res* 2015;8.
- Boguski M, McIntosh M. Biomedical informatics for proteomics. *Nature* 2003;422:233–237.
- Cabezón T, Gromova I, Gromov P, Serizawa R, timmermans Wielenga V, Kroman N, *et al.* Proteomic profiling of triple-negative breast carcinomas in combination with a three-tier orthogonal technology approach identifies Mage-A4 as potential therapeutic target in estrogen receptor negative breast cancer. *Mol Cell Proteomics*. 2013;12(2):381–394.
- Cai X, Huang W, Qiao Y, Du S, Chen Y, Chen D, *et al.* Inhibitory effects of curcumin on gastric cancer cells: a proteomic study of molecular targets. *Phytomedicine* 2013;20(6):495–505.
- Camerini S, Mauri P. The role of protein and peptide separation before mass spectrometry analysis in clinical proteomics. *J Chromatography A* 2015;1381:1–12.
- Canales N, Marina V, Castro J, Jimenez A, Mendoza-Hernandez G, McCarron E, *et al.* A1BG and C3 are overexpressed in patients with cervical intraepithelial neoplasia III. *Oncology Letters* 2014;8(2):939–947.
- Castro-Vega L, Letouzé E, Burnichon N, Buffet A, Disderot P, Khalifa E, *et al.* Multi-omics analysis defines core genomic alterations in pheochromocytomas and paragangliomas. *Nature Communications* 2015;27(6):6044.
- Corthals G, Wasinger V, Hochstrasser D, Sanchez J. The dynamic range of protein expression: A challenge for proteomic research. *Electrophoresis* 2000;21:1104–1115.
- Cragg G, Newman D. Natural products: a continuing source of novel drug leads. *Biochim Biophys Acta* 2013;1830(6):3670–3695.
- Di Palma S, Hennrich M, Heck A, Mohammed S. Recent advances in peptide separation by multidimensional liquid chromatography for proteome analysis. *J Proteomics* 2012;75(13):3791–3813.
- Dowsey A, Dunn M, Yang Q. The role of bioinformatics in two-dimensional gel electrophoresis. *Proteomics* 2003;3:1567–1596.
- Du M, Liu S, Gu D, Wang Q, Zhu L, Kang M, *et al.* Clinical potential role of circulating microRNAs in early diagnosis of colorectal cancer patients. *Carcinogenesis* 2014;35(12):2723–2730.
- Elliott M, Smith D, Parker C, Borchers C. Current trends in quantitative proteomics. *J Mass Spectrom*. 2009;44(12):1637–1660.
- Fénichel P, Rouzier C, Butori C, Chevallier P, Poullot A, Thyss A, *et al.* Extragestational β HCG secretion due to an isolated lung epithelioid trophoblastic tumor: microsatellite genotyping of tumoral cells confirmed their placental origin and oriented specific chemotherapy. *Clin Endocrinol Metab*. 2014;99(10):3515–3520.

- Fenn J, Mann M, Meng C, Wong S, Whitehouse CM. Electrospray ionization for mass spectrometry of large molecules. *Science* 1989;246:64–71.
- Freireich E, Wiernik P, Steensma D. The leukemias: a half-century of discovery. *J Clin Oncol* 2014;32(31):3463–3469.
- Fuji K, Suzuki N, Ikeda K, Hamada T, Yamamoto T, Kondo T, *et al.* Proteomic study identified HSP 70 kDa protein 1A as a possible therapeutic target, in combination with histone deacetylase inhibitors, for lymphoid neoplasms. *Proteomics* 2012;75(4):1401–1410.
- Gallardo M, Barrio S, Fernandez M, Paradela A, Arenas A, Toldos O, *et al.* Proteomic analysis reveals heat shock protein 70 has a key role in polycythemia Vera. *Mol Cancer* 2013;12:142.
- Girault A, Privé A, Trinh N, Bardou O, Ferraro P, Joubert P, *et al.* Identification of KvLQT1 K⁺ channels as new regulators of non-small cell lung cancer cell proliferation and migration. *International J Oncol* 2014;44(3):838–848.
- Gorg A, Obermaier C, Boguth G, Harder A, Scheibe B, Wildgruber R, Weiss W. The current state of two-dimensional electrophoresis with immobilized pH gradients. *Electrophoresis* 2000;21:1037–1053.
- Gromov P, Gromova I, Friis E, Timmermans-Wielenga V, Rank F, Simon R, *et al.* Proteomic profiling of mammary carcinomas identifies C7orf24, a gamma-glutamyl cyclotransferase, as a potential cancer biomarker. *J Proteome Res* 2010;9(8):3941–3953.
- Guan M, Chen X, Ma Y, Tang L, Guan L, Ren X, *et al.* MDA-9 and GRP78 as potential diagnostic biomarkers for early detection of melanoma metastasis. *Tumor Biology*. 201; PMID: 25480418
- Guo S, Zou J, Wang G. Advances in the proteomic discovery of novel therapeutic targets in cancer. *Drug Design Dev Ther* 2013;7:1259–1271.
- Guo X, Hao Y, Kamilijiang M, Hasimu A, Yuan J, Wu G, *et al.* Potential predictive plasma biomarkers for cervical cancer by 2D-DIGE proteomics and Ingenuity Pathway Analysis. *Tumour Biol* 2014; PMID: 25427637
- Hanahan D, Weinberg R. Hallmarks of cancer: The next generation. *Cell* 2011;144:646–674.
- Haslene-Hox H, Oveland E, Woie K, Salvesen H, Wiig H, Tenstad O. Increased WD-repeat containing protein 1 in interstitial fluid from ovarian carcinomas shown by comparative proteomic analysis of malignant and healthy gynecological tissue. *Biochemical Biophysics Acta* 2013;1834(11):2347–2359.
- Huang H, Jiang X, Li Z, Li Y, Song C, He C, *et al.* TET1 plays an essential oncogenic role in MLL-rearranged leukemia. *PNAS USA* 2013; 110(29):11994–11999.
- Hwang J, Yang H, Hong D, Joo S, Choi Y, Park J, *et al.* Gankyrin is a predictive and oncogenic factor in well-differentiated and dedifferentiated liposarcoma. *Oncotarget* 2014;5(19):9065–9078.
- Jiang L, Xiao X, Ren J, Tang Y, Weng H, Yang Q, *et al.* Proteomic analysis of bladder cancer indicates Prx-I as a key molecule in BI-TK/GCV treatment system. *PLoS One* 2014;9(6):e98764.
- Johnson I, Armstrong J, Gorman M, Burnet J. The Vinca Alkaloids: A New Class of Oncolytic

- Agents. *Cancer Res* 1963;23(8 Part 1):1390–1427.
- Karas M, Hillenkamp F. Laser desorption ionization of proteins with molecular mass exceeding 10,000 daltons. *Anal Chem* 1988;60:2299–2301.
- Kashuba E, Eagle G, Bailey J, Evans P, Welham K, Allsup D, *et al.* Proteomic analysis of B-cell receptor signaling in chronic lymphocytic leukaemia reveals a possible role for kininogen. *J Proteomics* 2013;91:478–485.
- Kim M, Pandey A. Electron transfer dissociation mass spectrometry in proteomics. *Proteomics* 2012;12(4–5):530–542.
- Köcher T, Pichler P, De Pra M, Rieux L, Swart R, Mechtler K. Development and performance evaluation of an ultralow flow nanoliquid chromatography-tandem mass spectrometry set-up. *Proteomics* 2014;14(17–18):1999–2007.
- Li Y, Chen X, Shi M, Wang H, Cao W, Wang X, *et al.* Proteomic-based identification of Apg-2 as a therapeutic target for chronic myeloid leukemia. *Cell Signalling* 2013;25(12):2604–2612.
- Lihong H, Linlin G, Yiping G, Yang S, Xiaoyu Q, Zhuzhu G, *et al.* Proteomics approaches for identification of tumor relevant protein targets in pulmonary squamous cell carcinoma by 2D-DIGE-MS. *PLoS One*. 2014;16;9(4):e95121.
- Liu P, Chen C, Wang C, Wu Y, Hsu C, Lee C, *et al.* In-depth Proteomic Analysis of Six Types of Exudative Pleural Effusions for Non-Small Cell Lung Cancer Biomarker Discovery. *Mol Cell Proteomics* 2015; pii: mcp. M114. 045914
- Liu Q, Zhao C, Zhang J, Chen Y, Gao L, Ni C, *et al.* Role of heat shock protein 27 in gemcitabine-resistant human pancreatic cancer: comparative proteomic analyses. *Mol Med Reports* 2012;6(4):767–773.
- Lochab S, Pal P, Kanaujiya J, Tripathi S, Kapoor I, Bhatt M, *et al.* Proteomic identification of E6AP as a molecular target of tamoxifen in MCF7 cells. *Proteomics* 2012;12(9):1363–1377.
- Manabe T. Combination of electrophoretic techniques for comprehensive analysis of complex protein systems. *Electrophoresis* 2000;21:1116–1122.
- Minca E, Tubbs R, Portier B, Wang Z, Lanigan C, Aronow M, *et al.* Genomic microarray analysis on formalin-fixed paraffin-embedded material for uveal melanoma prognostication. *Cancer Genet* 2014;207(7–8):306–315.
- Morisaki T, Yashiro M, Kakehashi A, Inagaki A, Kinoshita H, Fukuoka T, *et al.* Comparative proteomics analysis of gastric cancer stem cells. *PLoS One* 2014;9(11):e110736.
- Parker S, Raedschelders K, Van Eyk J. Emerging proteomic technologies for elucidating context-dependent cellular signaling events: A big challenge of tiny proportions. *Proteomics* 2014; doi: 10.1002/pmic.201400448
- Pospisilova J, Vit O, Lorkova L, Klanova M, Zivny J, Klener P, *et al.* Resistance to TRAIL in mantle cell lymphoma cells is associated with the decreased expression of purine metabolism enzymes. *Int J Mol Med* 2013; 31(5):1273–1279.
- Rebecca V, Wood E, Fedorenko I, Paraiso K, Haarberg H, Chen Y, *et al.* Evaluating melanoma drug response and therapeutic escape with quantitative proteomics. *Mol Cell*

- Proteomics* 2014;13(7):1844–1854.
- Rogowska-Wrzesinska A, Le Bihan M, Thaysen-Andersen M, Roepstorff P. 2D gels still have a niche in proteomics. *J Proteomics* 2013; 2(88):4–13.
- Sajic T, Liu Y, Aebersold R. Using data-independent, high resolution mass spectrometry in protein biomarker research: Perspectives and clinical applications. *Proteomics Clin Appl* 2014; doi: 10.1002/prca.201400117
- Sharma G, Mirza S, Parshad R, Gupta S, Ralhan R. DNA methylation of circulating DNA: a marker for monitoring efficacy of neoadjuvant chemotherapy in breast cancer patients. *Tumour Biol* 2012;33(6):1837–1843.
- Shipitsin M, Small C, Choudhury S, Giladi E, Friedlander S, Nardone J, *et al.*. Identification of proteomic biomarkers predicting prostate cancer aggressiveness and lethality despite biopsy-sampling error. *British J Cancer* 2014;111(6):1201–1212.
- Sjøholt G, Bedringaas S, Døskeland A, Gjertsen B. Proteomic strategies for individualizing therapy of acute myeloid leukemia (AML). *Curr Pharm Biotechnology* 2006;7(3): 159–170.
- Tang C, Tan T, Li C, Chen Z, Ruan L, Wang H, *et al.* Identification of Galectin-1 as a novel biomarker in nasopharyngeal carcinoma by proteomic analysis. *Oncology Rep* 2010; 24(2):495–500.
- Timms J, Cramer R. Difference gel electrophoresis. *Proteomics* 2008;8(23–24): 4886–4897.
- Wang L, Wheeler D. Genomic sequencing for cancer diagnosis and therapy. *Annual Rev Med* 2014;65:33–48.
- Wang Z, Feng X, Liu X, Jiang L, Zeng X, Ji N, *et al.* Involvement of potential pathways in malignant transformation from oral leukoplakia to oral squamous cell carcinoma revealed by proteomic analysis. *BMC Genomics* 2009;10:383.
- Wilkins M. Hares and tortoises: the high-versus low-throughput proteomic race. *Electrophoresis* 2009;30(Suppl 1):S150–S155.
- Yokoyama T, Enomoto T, Serada S, Morimoti A, Matsuzaki S, Ueda Y, *et al.* Plasma membrane proteomics identifies bone marrow stromal antigen 2 as a potential therapeutic target in endometrial cancer. *Int J Cancer* 2013;132(2): 472–484.
- Zhang S, Mercado-Uribe I, Hanash S, Liu J. iTRAQ-based proteomic analysis of polyploid giant cancer cells and budding progeny cells reveals several distinct pathways for ovarian cancer development. *PLoS One* 2013;8(11): e80120.

Genetic Markers and Evolution of Targeted Therapy in Cancer

Pratibha S. Kadam Amare

Cancer Cytogenetics Department, Tata Memorial Hospital, Annex Building, Room No. 726 B, Dr. Ernest Borges Road, Parel, Mumbai – 400012, India

The advances in biotechnology including high throughput platforms, and bioinformatics has resulted in detailing molecular pathology of various cancers, identifying targets such as fusion genes, chimeric RNA, fusion proteins, amplified gene, genes with point mutation, overexpression or down regulation of RNA, microRNA (miRNA) and aberrant DNA methylation. The genetic markers provide diagnostic, prognostic and therapeutic markers, and may also provide predictive markers. Several targeted molecules have been identified as cell surface antigens and tyrosine kinases e. g. FLT3, NPM1, CEBPA and PRAM1 in acute myeloid leukemia (AML); BCR-ABL1 in chronic myeloid leukemia; JAK2 in chronic myeloproliferative disorders; ALK, EGFR, K-RAS and BRAF in lung cancer; BRAF, KIT in melanoma; HER2 in breast cancer. The driver molecules and their mechanism of actions revealed various oncogenic pathways in the development of effective inhibitor molecules/proteins as targeted therapy, and novel mutations in the genes associated with the inhibitor protein. Targeted cancer therapy aimed to antagonize the deregulated molecule/s, commonly comprises therapeutic monoclonal antibodies and small molecule inhibitors. *In vitro* studies and clinical trials of the inhibitory molecules showed promising results as single drug therapy or in combination with conventional chemotherapy. Further, multiple mutations associated with resistance to targeted therapy were identified, leading to treatment with second line drugs and consequent better prognosis. Further advancements of biotechnology with identification of genetic variation, multiple resistant mutations which help discovery of a cascade of genetic markers with deeper understanding of biology of disease that offers hopes towards identification of development of more efficient targeted therapy with reduced toxicity and resistance.

Genetic Markers and Evolution of Targeted Therapy in Cancer

Advances in genomic technologies have resulted in remarkable progress in molecular diagnosis of cancer with identification of various unique genetic markers of pathogenic significance as

targeted molecules. The targeted molecules comprise fusion genes, chimeric RNA, fusion/chimeric proteins, amplified genes, genes with point mutation, overexpressed/down regulated RNA and miRNA (Ali *et al.*, 2010; Pavlovi *et al.*, 2014; Shtivelman *et al.*, 1985,

Key words: Genetic markers, Targeted molecule, Targeted Therapy, Tyrosine kinase.

***Corresponding Author:** Pratibha S. Amare Kadam, Cancer Cytogenetics Department, Tata Memorial Hospital, Annex Building, Room No. 726 B, Dr. Ernest Borges Road, Parel, Mumbai, India.

Email: pratibha.amare@gmail.com

Sjogren *et al.*, 1998). The genomic alterations have led to precise WHO classification of hematological malignancies resulting in differential diagnosis and stratification of patients for appropriate treatment protocols. The routine methods used in cancer include FISH, PCR, ARMS-PCR, RFLP, Real Time PCR, capillary electrophoresis, Sanger sequencing/pyrosequencing, microarrays for whole genome/transcriptome/protein analysis, mRNA and methylotype analysis (Ku *et al.*, 2013; Sethi *et al.*, 2013; Staehler *et al.*, 2012).

Several target molecules in cancer are tyrosine kinases, as the tyrosine kinase signaling initiates molecular cascades leading to cell proliferation, differentiation, apoptosis, migration, invasion, and angiogenesis in the malignant tissues. Hence, identification and development of tyrosine kinase inhibitors as therapeutic agents has revolutionized cancer therapy (Sawyers, 2002). Epidermal growth factor receptor (EGFR) is the first receptor tyrosine kinase (RTK) played an important role in the identification of significance of tyrosine kinases in cancer (Carpenter *et al.*, 1978). The tyrosine kinases are primarily RTKs e. g. EGFRs (EGFR-1, EGFR-2, EGFR-3), platelet-derived

growth factor receptor (PDGFR), fibroblast growth factor receptor (FGFR), vascular endothelial growth factor (VEGF) receptor, and non-receptor tyrosine kinases (NRTK), e. g. SRC, ABL1, Janus kinase. The RTKs are activated by ligands, epidermal growth factor (EGF), fibroblast growth factor (FGF), platelet-derived growth factor (PDGF) by binding to the extracellular domain of the receptors (Fig. 1).

The identification of the pathogenic molecules led to development of inhibitors as targeted drugs, impacting pharmacogenomics and personalized medicine. Targeted therapy directly interacts with pathognomic molecule, as against the cytotoxic drugs that primarily kill mitotic cells by interfering with cell cycle.

Targeted cancer drugs are generally monoclonal antibodies and small molecule inhibitors. Therapeutic monoclonal antibodies target specific antigens on the cell surface, such as transmembrane receptors, or extracellular growth factors, CD20, CD33, and CD52, present on leukemic and lymphoproliferative cells. Molecules associated with the immune mechanisms led to monoclonal antibodies – Rituximab, against CD20 (Table 1) in non-Hodgkin

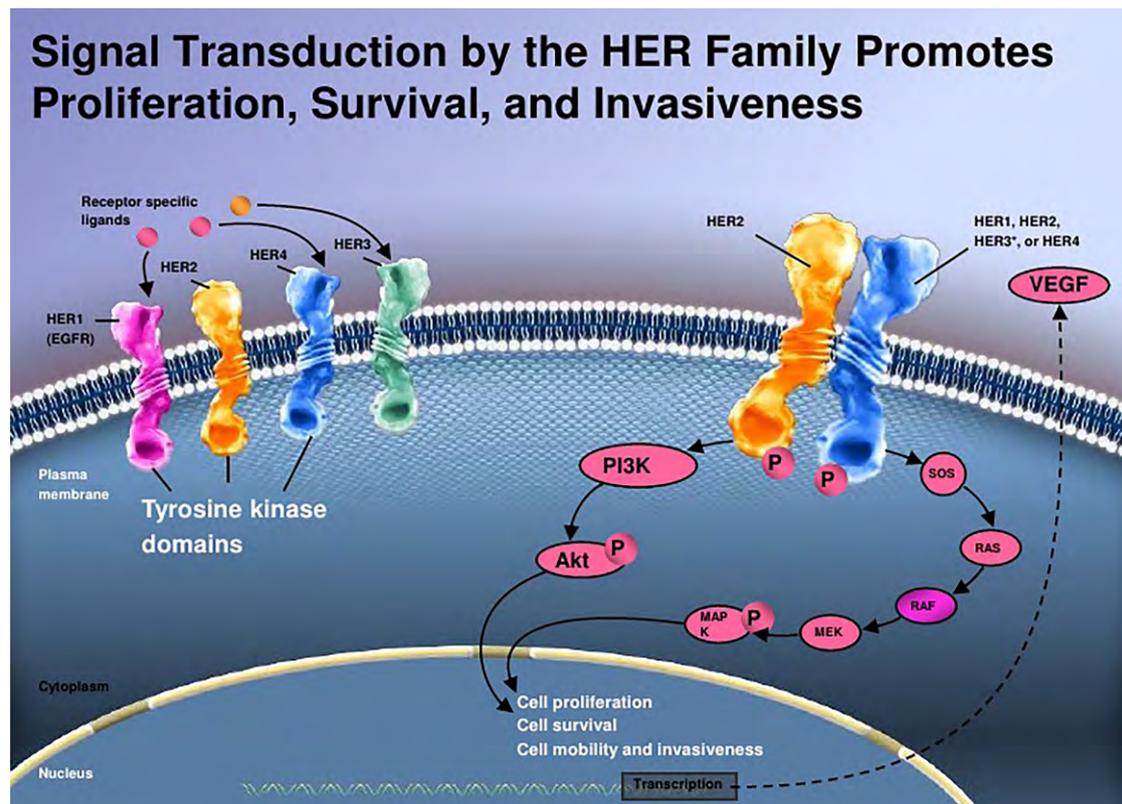


Figure 1: Activation of HER family receptors and signaling pathways (Adapted from: Hudis, 2007).

lymphoma (Silverman, 2007), and several monoclonal antibodies used in cancer treatment resulting in better prognosis (Fig. 1; Table 1). The monoclonal antibodies also target extracellular components of signaling pathways, including ligands and receptor binding domains blocking receptor signaling and downstream intracellular proteins involved in cellular proliferation, angiogenesis and invasion.

Small molecule inhibitors penetrate the cell membrane interacting with enzymatic activity of proteins, thereby blocking receptor signaling and interfering

with downstream intracellular molecules (Fig. 2). Several growth factor receptors with intrinsic tyrosine kinase activity are constitutively active in cancers and inhibition of the kinases using small molecule inhibitors sensitizes the tumor cells to apoptosis.

RTKs are preferred key targets for anti-cancer drugs as aberrant activation of the RTKs usually result in downstream signaling with activation of pivotal cytoplasmic serine/threonine kinases (STKs). Small molecule cancer inhibitors targeting extracellular RTKs and cytoplasmic STKs are extensively studied

Table1. Molecular targets and targeted therapies in cancer (Abramson, 2015)

Agent	Target (s)	Malignancy
Ado-trastuzumab emtansine(Kadcyla)	HER2 (ERBB2/neu)	Breast cancer (HER2+)
Lapatinib (Tykerb)	HER2 (ERBB2/neu), EGFR(HER1/ERBB1)	Breast cancer (HER2+)
Trastuzumab (Herceptin)	HER2 (ERBB2/neu)	Breast cancer (HER2+), Gastric cancer (HER2+)
Afatinib (Gilotrif)	EGFR(HER1/ERBB1), HER2(ERBB2/neu)	Non-small cell lung cancer (NSCLC)(with EGFR exon 19 deletions or exon 21 substitution)
Ceritinib (Zykadia)	ALK	Non-small cell lung cancer
Crizotinib (Xalkori)	ALK, MET	Non-small cell lung cancer
Gefitinib (Iressa)	EGFR (HER1/ERBB1)	Non-small cell lung cancer with known prior benefit from Gefitinib
Ramucirumab (Cyramza)	VEGFR2	Gastric cancer, adenocarcinoma, Non-small cell lung cancer
Axitinib (Inlyta)	KIT, PDGFR β , VEGFR1/2/3	Renal cell carcinoma (RCC)
Bevacizumab (Avastin)	VEGF ligand	Cervical cancer, Colorectal cancer, Glioblastoma, NSCLC, Ovarian cancer, RCC
Pazopanib (Votrient)	VEGFR, PDGFR, KIT	Renal cell carcinoma
Temsirolimus (Torisel)	mTOR	Renal cell carcinoma
Panitumumab (Vectibix)	EGFR(HER1/ERBB1)	Colorectal cancer (KRAS wild type)
Cetuximab (Erbix)	EGFR (HER1/ERBB1)	Colorectal cancer (KRAS wild type), Squamous cell cancer of head and neck
Everolimus (Afinitor)	mTOR	Pancreatic neuroendo tumor, RCC, Breast cancer (HR+, HER2-)
Erlotinib (Tarceva)	EGFR (HER1/ERBB1)	NSCLC, Pancreatic cancer
Bortezomib (Velcade)	Proteasome	Multiple myeloma, Mantle cell lymphoma
Carfilzomib (Kyprolis)	Proteasome	Multiple myeloma
Brentuximab vedotin (Adcetris)	CD30	Hodgkin lymphoma, Anaplastic large cell lymphoma
Alemtuzumab (Campath)	CD52	B-cell Chronic lymphocytic leukemia (CLL)
Dabrafenib (Tafinlar)	BRAF	Melanoma (with BRAF V600 mutation)
Ibritumomab tiuxetan (Zevalin)	CD20	Non-Hodgkin's lymphoma
Idelalisib (Zydelig)	PI3K δ	CLL, Follicular B-cell NHL, Small lymphocytic lymphoma
Rituximab (Rituxan, Mabthera)	CD20	Non-Hodgkin's lymphoma, Chronic lymphocytic leukemia
Imatinib (Gleevec)	KIT, PDGFR, ABL	GI stromal tumor (KIT+), Hematologic malignancies including, Ph +ve ALL and CML
Nilotinib (Tasigna)	ABL1	CML.
Ponatinib (Iclusig)	ABL1, FGFR1-3, FLT3, VEGFR2	CML, ALL- Ph positive
Ruxolitinib (Jakafi)	JAK1/2	Myelofibrosis

(Arora *et al.*, 2005)) Deregulated activation of RTKs results in increased cell growth and survival, and contributes to progression of cancer.

Targeted cancer drugs are designated as per the content of basic compound like monoclonal antibodies that end with "-mab", e.g., Rituximab, whereas small

molecules end with the stem "-ib" indicating protein inhibitory action of targeted drug. For example, the small molecule STI-571 known as Imatinib (generic name) in which tinib indicated tyrosine kinase inhibitor (TKI). Drug with stem "-zom-" indicates proteasome inhibitors, e.g., Bortezomib. Small

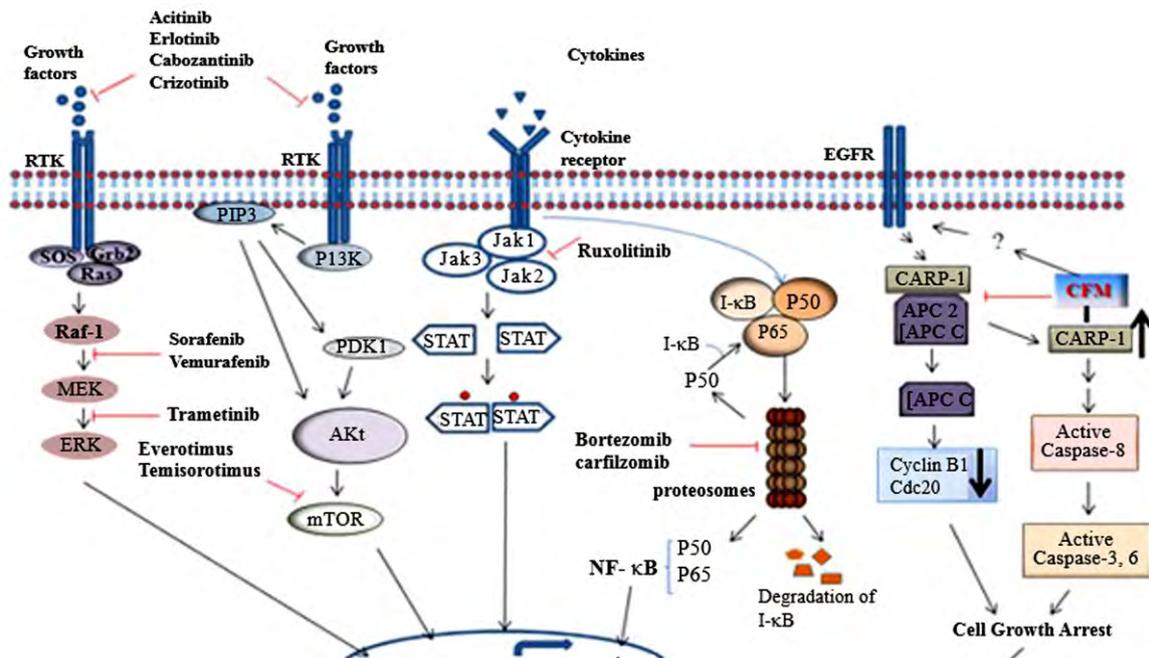


Figure 2: Schematic representation of activated cellular pathways in cancer and mechanism of small molecule inhibitors (Source: Lavanya *et al.*, 2014).

molecule inhibitors, tyrosine kinase inhibitors interrupt various intracellular signaling pathways of tyrosine kinases (Table 1).

Tyrosine Kinase Deregulation and Targeted Therapy in Hematolymphoid Malignancies

Chronic myeloid leukemia (CML) is a hematopoietic stem cell disorder associated with reciprocal translocation between chromosomes 9 (*BCR*) and 22 (*ABL1*) juxtaposing *BCR* sequences to *c-ABL*. *c-ABL* is a tyrosine kinase located at chromosome 9q34, resulting in constitutive production of fusion chimeric protein p210 with increased tyrosine

kinase activity. The deregulated kinase activity usurps the physiologic functions of normal ABL enzyme by interacting with a variety of effector proteins, resulting in deregulated cellular proliferation, decreased adherence of leukemic cells to the bone marrow stroma and a reduced apoptotic stimuli (Deininger *et al.*, 2000).

In acute lymphoblastic leukemia (ALL), TEL-ABL protein is constitutively phosphorylated due to reciprocal translocation t(9;12) (Hannemann *et al.*, 1998). Chronic myelomonocytic leukemia (CMML) with t(5;12) produces TEL-PDGFRB fusion protein, leading to tyrosine kinase activation (Golub *et al.*, 1994). NPM1-ALK fusion product of

t(2;5) is constitutively activated in anaplastic large cell lymphoma (Shiota *et al.*, 1995).

Imatinib mesylate, a tyrosine kinase inhibitor in CML (Druker *et al.*, 2001), acts via competitive inhibition at the ATP-binding site of the BCR-ABL1 protein, resulting in inhibition of phosphorylation of the downstream cascade of proteins in signal transduction pathways. Imatinib mesylate prevents BCR-ABL enzyme from permanent deactivation, thus inhibiting proliferation of leukemic cells and leading to apoptosis (Table 1) (Deshmukh *et al.*, 2005; Druker *et al.*, 2001). Imatinib mesylate efficiently inhibits BCR-ABL kinase, blocks platelet-derived growth factor receptor, and c-kit tyrosine kinase (Druker *et al.*, 2000). However, about 90 kinase domain mutations have been identified in *ABL1*, which prevents binding of the drug and thus induce resistance to the drug. Consequently, second generation tyrosine kinase inhibitors, Nilotinib, Dasatinib (Jabbour *et al.*, 2014; Kantarjian *et al.*, 2010) and Bosutinib (Khoury *et al.*, 2012) were developed to overcome resistance to Imatinib mesylate due to kinase domain mutations. Second generation TKIs overcome resistance of Imatinib.

However T3151, “gatekeeper” mutation, displays resistance to all second generation TKIs. Ponatinib, a third generation TKI, has overcome resistance due to kinase mutation T3151 (Jabbour *et al.*, 2014; O'Hare *et al.*, 2009) (Table 1).

In acute promyelocytic leukemia (APL), fusion gene *PML-RARA* of t(15;17) leads to a differentiation block in the abnormal promyelocytes. The targeted drug all-trans-retinoic acid (ATRA) leads to conformational change of PML-RARA protein followed by activation and regulation of RARA-responsive genes leading to differentiation of promyelocytes to granulocytes (Advani *et al.*, 1999; Grignani *et al.*, 1998). The remission rates were significantly high in APL patients treated by ATRA. However, resistance to ATRA was observed in 25–30% of APL patients (Estey *et al.*, 2006), and arsenic trioxide (ATO) was found to be more efficient than ATRA as it induced apoptosis in addition to differentiation.

Besides, translocations, epigenetic silencing is an important genetic alteration leading abnormal expression of genes involved in cell cycle control and differentiation in AML. The replacement of cytosine by 5-aza-cytidine, a cytidine analogue, acts as a block to DNA methyl

transferases, causing demethylation of DNA and consequent differentiation (Egger *et al.*, 2004). Histone deacetylase (HDAC) inhibitors Vorinostat (Zolinza) and Panobinostat are additional agents for modulation of transcriptional repression of tumor suppressor proteins (Bolden *et al.*, 2006).

The two most prominent mechanisms in Myelodysplastic syndromes (MDS), DNA methylation and histone acetylation play a role in hematopoiesis. Methylation is focally increased around tumor suppressors and other mitogen inhibitors. DNA methyl transferases (DNMTs) play a role in increased methylation and hence a key target for treatment of MDS (Shih *et al.*, 2012). In high risk MDS, a number of genes associated with DNA repair, cell-cycle control, regulation of development, differentiation and apoptosis are hypermethylated in 70% of patients. The critical hypermethylated genes are *ALOX12*, *GSTM1*, *HIC1*, *FZD9*, *TET2* and *HS3ST2* (Jiang *et al.*, 2009). These hypermethylated genes are potential targets for demethylating agents. Patients with hypermethylated *TET2* showed better response rates (82%) on treatment with demethylating drug azacytidine than those with wild-type *TET2* (45%) (Itzykson *et*

al., 2011).

JAK2 mutation has been reported in myeloproliferative disorders Polycythemia Vera, Primary Myelofibrosis and Essential Thrombocythemia. *JAK2* encodes an on-receptor tyrosine kinase associated with signal relays for hemopoietic cell growth, development and differentiation (Neubauer *et al.*, 1998). Ruxolitinib, a JAK inhibitor showed promising results in patients with Myelofibrosis (Harrison *et al.*, 2012).

BRAF is a potent activator of MAP/ERK kinase pathway associated with regulation of cell cycle, differentiation and cell survival. *BRAF* mutations have been reported in solid cancers and hematopoietic cancers (Davies *et al.*, 2002; Holderfield *et al.*, 2014). The most common *BRAF* mutation is the V600E mutation (Holderfield *et al.*, 2014). Vemurafenib, a small molecule inhibitor showed anti-melanoma activity against the *BRAF* V600E mutant protein (Tsai *et al.*, 2008). Hematolymphoid malignancies including hairy cell leukemia and multiple myeloma with *BRAF* V600E mutation, showed favourable clinical response on treatment with Vemurafenib (Machnicki *et al.*, 2014) (Table 1).

Fms-like tyrosine kinase 3, CD135 (FLT3) a tyrosine kinase receptor is activated when bound by the FLT3 ligand (FL), subsequently promoting homodimerization. This switches tyrosine kinase activity of FLT3 followed by recruitment and phosphorylation of intracellular proteins SHC, GRB2, SHIP, CBL, CBLB-related protein domain, further leading to activation of MAP kinase, STAT and AKT/PI3 kinase signal transduction pathways. The proteins are transported to the nucleus regulating cellular proliferation, differentiation and apoptosis (Zhang *et al.*, 1999). *FLT3-ITD* (Internal tandem duplication) is a common mutation in 15–35% AML (Stirewalt *et al.*, 2006) and 5–10% MDS. *FLT3-ITD* and allelic variation in patients influences prognosis of AML patients (Meshinchi *et al.*, 2006). *FLT3-TKD* (Tyrosine kinase domain) mutation occurs in codon 835 (D835). Sorafenib, a tyrosine kinase inhibitor specifically targets the leukemic blasts in AML (Williams *et al.*, 2012) (Table 1).

Upregulation of JAK2 in AML cells results in resistance to *FLT3-TKI* inhibition (Ikezoe *et al.*, 2011). Second generation drug, Quizartinib (AC220) was potent in *FLT3-TKI* resistant cases due to

upregulation of JAK2 (Cortes *et al.*, 2011). Pacritinib (SB 1518) is another potent JAK2/FLT3 inhibitor, in combination with Pracinostat (SB939), an oral HDAC inhibitor, showed synergy in inducing remission and better survival in the patients (Novotny-Diermayr *et al.*, 2012).

Nucleophosmin (NPM1) mutations result in overexpression of the phosphoprotein in 27–35% of adult AML and 40–60% of adult AML with normal karyotype (Falini *et al.*, 2005). *NPM1* mutation occurs due to four base sequence TCTG duplication at position 956–959 in *NPM1* gene (Falini *et al.*, 2005). Inhibitors of *NPM1* oligomerization such as NSC348884 increase apoptosis when exposed to the ATRA plus cytarabine combination (Balusu *et al.*, 2011).

CCAAT/enhancer binding protein alpha (*CEBPA*) protein is a key regulator of granulocytic differentiation (Rosenbauer *et al.*, 2007). Hence, *CEBPA* mutations induce proliferation and block differentiation of myeloid lineage. *CEBPA* mutation occurs due to N-terminal frameshift mutations and secondly due to C-terminal in-frame insertions or deletions. *CEBPA* mutations frequently (70%) occur in AML patients exhibiting a normal karyotype. AML patients with a normal

karyotype and *CEBPA* mutation in the absence of *FLT3* show favorable prognosis (Green *et al.*, 2010).

C-KIT, a stem cell gene, encoding tyrosine kinase, demonstrated c-KIT mutations in AML patients with core binding factor rearrangement. Upon binding of the ligand stem cell factor, to c-kit, phosphorylation of several cytoplasmic proteins occur followed by activation of downstream MAP kinase, JAK/STAT, and PI-3 kinase pathways (Linnekin, 1999). Mutations in *c-KIT* receptor result in constitutive phosphorylation and activation of the receptor in absence of the ligand. Mutations in *c-KIT* and *FLT3* genes are associated with unfavorable prognosis in patients with t(8;21). In particular, patients with *c-KIT* mutation have been reported to have a higher incidence of relapse (80% versus 13.5%) (Pascka *et al.*, 2004). *In vitro* studies have shown sensitivity to Imatinib for a mutation in exon 8 and exon 17. APcK110, with potent proapoptotic and antiproliferative activities has shown promising results in AML cell lines and primary samples (Faderl *et al.*, 2011).

BCL2, an anti-apoptotic protein, is overexpressed in hematological malignancies and is a possible molecule

for targeted therapy. AML patients treated with Bcl-2 antisense oligonucleotide based therapy inhibit Bcl-2 overexpression, promote apoptosis and reduce drug resistance (Marcucci *et al.*, 2003).

Targeted Therapy in Solid Tumors

According to National Comprehensive Cancer Network (NCCN) guidelines, several molecular markers have been identified as targets for therapy in solid tumors. The molecular markers include *HER2 (ERBB2)* amplification in breast cancer, *K-RAS* and *BRAF* mutations in colorectal cancer, and *BRAF* v600 mutation in melanoma, *EGFR* mutation/*ALK*/rearrangement in non-small-cell-lung-cancer (NSCLC), and *c-KIT* in gastrointestinal stromal cancer. The following section discusses the markers in specific cancers.

Molecular Markers and Targeted Therapy in Lung Cancer

Lung cancer is the most common cancer in men globally with about 15% five year survival rates. Based upon various driver mutations, NSCLC is stratified based on the molecular lesions as NSCLC with *K-RAS* mutation, *EGFR* mutation, echinoderm microtubule-associated

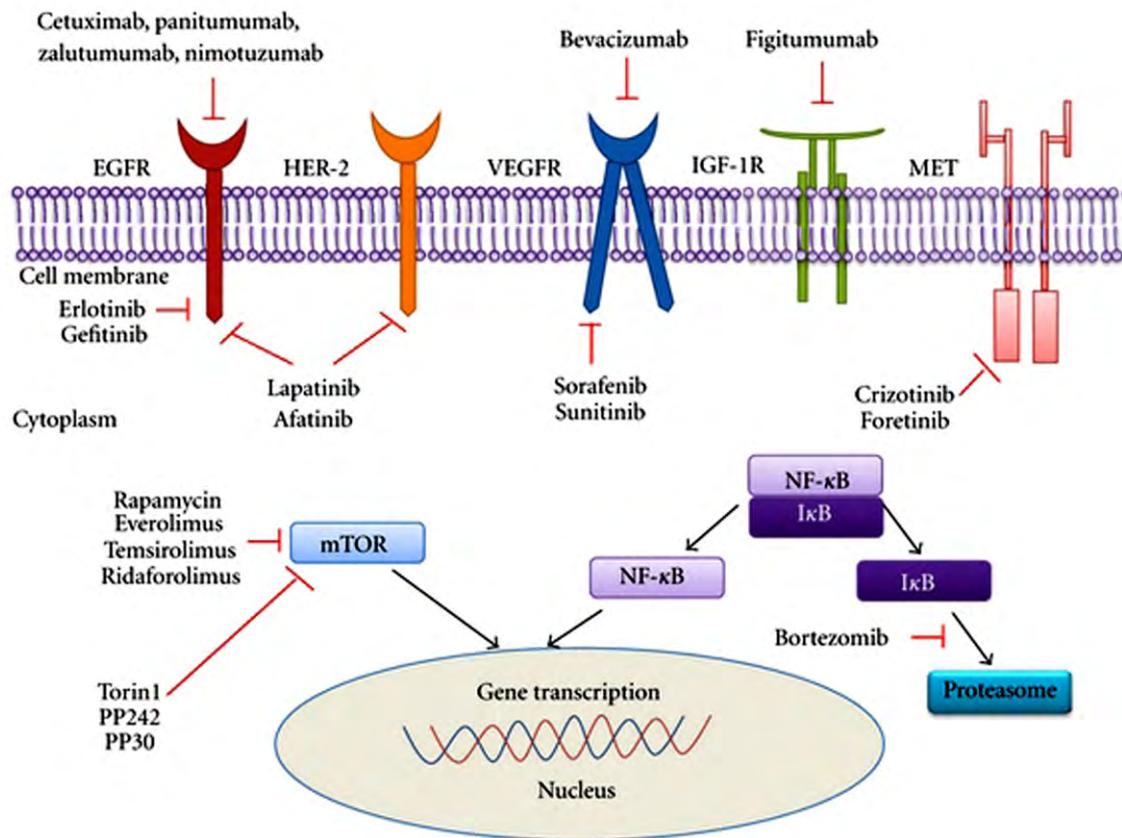


Figure 3: Mechanism of targeted therapies in patients with HER2 and EGFR, VEGFR and IGF-1R mutation (Source: Gao *et al.*, 2012).

protein like 4-anaplastic lymphoma kinase (*EML4-ALK*) mutation, *herceptin 2* (*HER2*) mutation, *v-raf* murine sarcoma (*BRAF*) mutation, mesenchymal epithelial transcription factor (*Met*) mutation, protein kinase B (*PKB/AKT1*), *phosphatidylinositide 3 kinase catalytic subunit* (*PI3KCA*) mutation (Pao *et al.*, 2011).

EGFR plays a critical role in cell proliferation, angiogenesis, and inhibition of apoptosis. *EGFR* mutation is reported in 10% of NSCLC in US, and 35% in Asian population (Pao *et al.*, 2011). The *EGFR*

mutation is observed in less than 5% squamous cell cancer patients and 15–20% adenocarcinomas including females (never smokers) (Pao *et al.*, 2010). *EGFR* mutations are located in the kinase domain at exons 18–21 (Kosaka *et al.*, 2009). *EGFR* amplification has also been reported in NSCLC patients and associated with bad prognosis. Patients stratified as NSCLC with *EGFR* mutation are effectively treated with targeted therapy Erlotinib or Gefitinib targeted to the deregulated *EGFR* (Lazarus *et al.*, 2013) (Figs. 3 and 4) (Table 1). *EGFR* TKI,

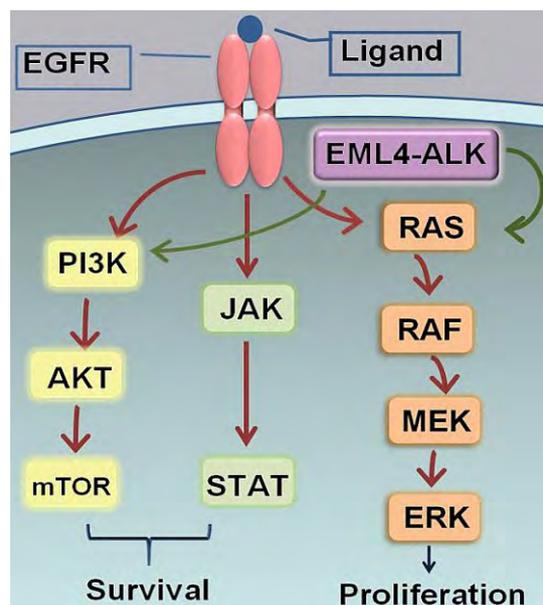


Figure 4: EGFR mutation, EML4-ALK translocation and Signaling (Source: Wu *et al.*, 2012).

a small molecule inhibitor, therapy also shows better response to patients with *EGFR* amplification as compared with *EGFR* mutation.

An additional molecular lesion in lung adenocarcinomas is the point mutation in *K-RAS* gene, codon 12 or 13 (Knickelbein and Zang, 2015). *EGFR* mutation activates *RAS* signaling pathway downstream, hence patients with *K-RAS* mutation are resistant to *EGFR* TKI (Raponi, 2008).

ALK encodes a tyrosine kinase receptor normally expressed in selected neuronal cell types. *ALK-EML4* rearrangement translocation and balanced translocations retain *ALK* kinase domain with constitutive activation of tyrosine kinase, leading to transformation of cells

(Soda, *et al.*, 2007) (Fig. 4). In Lung cancer, *ALK* rearrangement is detected by FISH with an *ALK* break-apart probe (Soda *et al.*, 2007). Lung cancer patients with *EML4-ALK* translocation show sensitivity to TKI inhibitor Crizotinib (Shaw *et al.*, 2011). However, resistance to the targeted therapy has been reported in patients with secondary mutations in *ALK* (Ettinger *et al.*, 2012; Sasaki *et al.*, 2011).

FGFR1, Fibroblast growth factor receptor 1 encodes a member of the *FGFR* tyrosine kinase family, with a critical role in cell development. *FGFR1* is deregulated either by point mutation, translocation or amplification (Turner *et al.*, 2011). Preclinical trials with *FGFR1* inhibitors have shown encouraging results in lung cancer (Weiss *et al.*, 2010). *FGFR1* amplifications are also observed in 20% in smokers with squamous cell sarcoma.

***K-RAS* Mutations and Targeted Therapy in Colorectal Cancer**

KRAS is a membrane bound GTPase, active in the GTP-bound form and inactive when GDP-bound. *KRAS* activity mediates a cascade of intracellular signaling events initiated by the ligand-receptor binding of RTKs, including *EGFR* (Downward *et al.*, 2003). *EGFR*

upon binding to its ligand is auto-phosphorylated creating a docking site for the adaptor protein growth factor receptor bound protein 2 (GRB2), resulting in activation of KRAS GTP, which further stimulates downstream signaling pathways, RAF/MEK and PI3K (and phosphoinositide-3 kinase)/AKT controlling cell growth and survival (Downward *et al.*, 2003) (Fig. 4). *K-RAS* mutations resulting in constitutive activation of RAS with expression of RAS proteins are reported in 20–25% of several human tumors including pancreatic cancer with *K-RAS* mutation in 90% (Downward *et al.*, 2003). The potent transforming mutations are detected in codons 12 (82% of *K-RAS* mutations) and 13 (17%) in exon 2 of the *K-RAS* gene (Wang *et al.*, 2010). *K-RAS* gene mutations predict outcome of treatment with anti-EGFR antibodies in advanced colorectal cancer (CRC).

Cetuximab, a human–mouse chimeric IgG1 monoclonal antibody, EGFR-targeted agent approved for the treatment of colorectal cancer (Jonker *et al.*, 2007) (Fig. 3), and Panitumumab are commonly used in CRC therapy (Heinemann *et al.*, 2013). Bevacizumab (Avastin), Ramucirumab (Cyramza), and Ziv-aflibercept (Zaltrap) are drugs used for

colon cancer that target VEGF (Douillard *et al.*, 2014). These drugs are combined with chemotherapy to treat advanced colon cancer (Table 1). Farnesyl transferase inhibitors (FTIs) are small molecule inhibitors that selectively inhibit farnesylation of a number of intracellular substrate proteins such as RAS, an additional approach to target *K-RAS* mutations (Gysin *et al.*, 2013). However, a comprehensive understanding of RAS mediated signal transduction feedback loops, tumor heterogeneity and mechanisms of downstream targets of *K-RAS* gene on CRC is needed for optimal use of the monoclonal antibodies, small molecular inhibitors to *K-RAS* aberrations.

***HER2* Marker and Targeted Therapy in Breast Cancer**

HER2 amplification has been observed in 20% invasive breast carcinomas, and is a poor prognostic marker with an increased risk of disease progression, recurrence of disease with poor survival (Andrulis *et al.*, 1998). FISH is an efficient tool for detection of *HER2* amplification. *HER2* encodes a transmembrane tyrosine kinase receptor in the EGFR family. *HER2* stimulates growth factor signaling

pathways such as PI3K–AKT–mTOR pathway (Fig. 1). Trastuzumab (Herceptin), a humanized, recombinant monoclonal antibody that binds to the extracellular domain of HER2 is an efficient targeted therapy (Vogel *et al.*, 2002) (Fig. 3). Trastuzumab selectively blocks ligand independent HER2–HER3 dimerization and proteolytic cleavage of the extracellular domain of HER2 resulting in downregulation of PI3K pathway signaling and downstream cell cycle protein cyclin D1 (Junttila *et al.*, 2009). Herceptin resistance is seen in several breast cancer patients with mutational activation of P13K pathway through loss of *PTEN*, indicating PI3K-based treatment options. Lapatinib, an ATP-competitive inhibitor of HER2 and EGFR tyrosine kinases, have shown efficacy in Trastuzumab resistant patients (Konecny *et al.*, 2006). Pertuzumab monoclonal antibody binding to a distinct epitope on the extracellular domain of HER2 blocks ligand induced dimerization of HER2 and HER3 (Junttila *et al.*, 2009) (Table 1).

BRAF V600E mutation occurs in 60% melanoma patients. The mutation constitutively activates mitogen activated protein kinase (MAPK) pathway,

promoting cell proliferation and preventing apoptosis (Gray-Schopfer *et al.*, 2007). Hence, *BRAF* V600E mutation is considered as a promising therapeutic target in metastatic melanoma. Vemurafenib treatment in patients with *BRAF* V600mutant metastatic melanoma indicated that inhibition of MAPK pathway promoted cell proliferation and prevented apoptosis (Flaherty *et al.*, 2010). Vemurafenib induces clinical responses in 50% patients with *BRAF* V600 mutant metastatic melanoma. Vemurafenib and Dabrafenib are effective targeted drugs for melanomas with *BRAF* V600Emutation (Kim *et al.*, 2014) (Table 1).

CONCLUSION

A continuous research efforts by various genomic technologies made remarkable progress in the discovery of genetic markers which have diagnostic as well as prognostic significance in hemato-lymphoid malignancies and solid tumors as well. Driver mutations and their mechanism of actions disclosed role of various oncogenic pathways that contributed significantly in the development of effective inhibitor molecules/proteins as targeted therapy.

Clinical trials of the inhibitor molecules have shown promising results in comparison with traditional cytotoxic chemotherapy. Further advancement in genomics is expected to identify cascade of genetic markers help understanding biology of disease that offers hopes towards development of more efficient targeted therapy with reduced toxicity and resistance.

REFERENCES

- Abramson RG. Overview of Targeted Therapies for Cancer. *My Cancer Genome*. 2015. <http://www.mycancergenome.org/content/molecular-medicine/overview-of-targeted-therapies-for-cancer>. Accessed on: September 25, 2015.
- Advani S, Nair R, Bafna A, Amare-Kadam P, Gladstone B, Saikia T, *et al*. Acute promyelocytic leukemia: All trans retinoic acid (ATRA) along with chemotherapy is superior to ATRA alone. *Am J Hematol* 1999; 60:87–93.
- Ali S, Almhanna K, Chen W, Philip A, Sarkar F. Differentially expressed miRNAs in the plasma may provide a molecular signature for aggressive pancreatic cancer. *Am J Trans Res* 2010;3:28–47.
- Andrulis I, Bull S, Blackstein M, Sutherland D, Mac C, Sidlofsky S, *et al*. Neu/erbB-2 amplification identifies a poor-prognosis group of women with node-negative breast cancer. Toronto Breast Cancer Study Group. *J Clin Oncol* 1998;16:1340–1349.
- Arora A, Scholar E. Role of tyrosine kinase inhibitors in cancer therapy. *J Pharmacol Exp Ther* 2005;315:971–979.
- Balusu R, Fiskus W, Rao R, Chong D, Nalluri S, Madunuru U, *et al*. Targeting levels or oligomerization of nucleophosmin induces differentiation and loss of survival of human AML cells with mutant NPM1. *Blood* 2011;118:3096–3106.
- Bolden J, Peart M, Johnstone R. Anticancer activities of histone deacetylase inhibitors. *Nat Rev Drug Discov* 2006;5:769–84.
- Carpenter, G, King, Land Cohen S. Epidermal growth factor stimulates phosphorylation in membrane preparations in vitro. *Nature* 1978;276:409–410.
- Cortes J, Perl A, Smith C, Kovacsovic T, Dombret H, Dohner H, *et al*. A phase II open-label, Ac220 monotherapy. Efficacy study in patients with refractory/relapsed FLT3-ITD positive acute myeloid leukemia: updated interim results. *ASH Annual Meeting Abstracts* 2011;118:2576.
- Davies H, Bignell GR, Cox C, Stephens P, Edkins S, Cleqq S, *et al*. Mutations of the BRAF gene in human cancer. *Nature* 2002;417:949–954.
- Deininger MW, Goldman JM, Melo JV. The molecular biology of chronic myeloid leukemia. *Blood* 2000;96:3343–3356.
- Deshmukh C, Saikia T, Bakshi A, Amare-Kadam P, Baisane C, Parikh P. Imatinib Mesylate in chronic myeloid leukemia: A prospective single arm nonrandomized study. *JAPI* 2005;53:291–295.
- Douillard J, Siena S, Cassidy J, Taberero R,

- Burkes M, Barugel Y, *et al.* Final results from PRIME: randomized phase III study of panitumumab with FOLFOX4 for first-line treatment of metastatic colorectal cancer. *Ann Oncol* 2014;25(7):1346–1355.
- Downward J. Targeting RAS signaling pathways in cancer therapy. *Nat Rev Cancer* 2003;3:11–22.
- Druker B, Lydon N. Lessons learned from the development of an abl tyrosine kinase inhibitor for chronic myelogenous leukemia. *J Clin Invest* 2000;105:3–7.
- Druker B, Talpaz M, Resta D, Peng B, Buchdunger E, Ford J, *et al.* Efficacy and safety of a specific inhibitor of the BCR-ABL tyrosine kinase in chronic myeloid leukemia. *N Engl J Med* 2001;344:1031–1037.
- Egger G, Liang G, Aparicio A, Jones P. Epigenetics in human disease and prospects for epigenetic therapy. *Nature* 2004;429:457–463.
- Estey E, Garcia-Manero G, Ferrajoli A, Faderi S, Verstovsek S, Jones D, *et al.* Use of all-trans retinoic acid plus arsenic trioxide as an alternative to chemotherapy in untreated acute promyelocytic leukemia. *Blood* 2006;107:3469–3473.
- Ettinger D, Akerly W, Borghaei H, Chang A, Cheney R, Chirieac L, *et al.* National Comprehensive Cancer Network Guidelines: Non-Small Cell Lung Cancer v. 1. 2013. *Clinical Practice Guidelines* 2012.
- Faderl S, Bueso-Ramos C, Liu Z, Pal A, Bommann W, Ciurea D, *et al.* Kit inhibitor APcK110 extends survival in an AML xenograft mouse model. *Invest New Drugs* 2011;29:1094–1097.
- Falini B, Mecucci C, Tiacci E, Alcalay M, Rosati R, Pasqualucci L, *et al.* Cytoplasmic nucleophosmin in acute myelogenous leukemia with a normal karyotype. *N Engl J Med* 2005;352:254–266.
- Flaherty KT, Puzanov I, Kim KB, Ribas A, McArthur GA, Sosman JA, *et al.* Inhibition of mutated, activated *BRAF* in metastatic melanoma. *N Engl J Med* 2010;363:809–819.
- Gao W, Li JZH, Chan JYW, Ho WK, Wong TS. mTOR pathway and mTOR inhibitors in head and neck cancer. 2012;2012(953089).
- Gray-Schopfer V, Wellbrock C, Marais R. Melanoma biology and new targeted therapy. *Nature* 2007;445:851–857.
- Golub, T, Barker, G, Lovett, M, Gilliland D. Fusion of *PDGF* receptor beta to a novel ets-like gene, *tel*, in chronic myelomonocytic leukemia with t (5; 12) chromosomal translocation. *Cell* 1994;77:307–316.
- Green C, Koo K, Hills R, Burnett A, Linch D, Gale R. Prognostic significance of *CEBPA* mutations in a large cohort of younger adult patients with acute myeloid leukemia: impact of double *CEBPA* mutations and the interaction with *FLT3* and *NPM1* mutations. *J Clin Oncol* 2010;28:2739–2747.
- Grignani F, De Matteis S, Nervi C, Tomassoni L, Gelmetti V, Cioce M, *et al.* Fusion proteins of the retinoic acid receptor-alpha recruit histone deacetylase in promyelocytic leukaemia. *Nature* 1998;391:815–818.
- Gysin S, Salt M, Young A, and McCormic F. Therapeutic strategies for targeting ras proteins. *Genes Cancer* 2011;2(3):359–372.
- Hannemann J, McManus D, Kabarowski J and Wiedemann L. Haematopoietic transformation by the TEL-ABL *Oncogene* 1998;102:475–485.

- Harrison C, Kiladjan J, Al-Ali H. JAK inhibition with ruxolitinib versus best available therapy for myelofibrosis. *N Engl J Med* 2012;366:787–798.
- Heinemann V, Douillard J, Ducreux M, Peeters M. Targeted therapy in metastatic colorectal cancer – An example of personalized medicine in action. *Cancer Treat Rev* 2013;39(6):592–601.
- Holderfield M, Deuker M, McCormick F, McMahon M. Targeting RAF kinases for cancer therapy: *BRAF*-mutated melanoma and beyond. *Nat Rev Cancer* 2014;14:455–467.
- Hudis CA. Trastuzumab – Mechanism of action and use in clinical practice. *N Engl J Med* 2007;357:39–51.
- Ikezoe T, Kojima S, Furihata M, Yang J, Nishioka C, Takeuchi A, *et al.* Expression of p-JAK2 predicts clinical outcome and is a potential molecular target of acute myelogenous leukemia. *Int J Cancer* 2011;129:2512–2521.
- Itzykson R, Kosmider O, Cluzeau T, Mansat-De Mas V, Dreyfus F, Beyne-Rauzy O, *et al.* Groupe Francophone des Myélodysplasies (GFM) : Impact of TET2 mutations on response rate to azacitidine in myelodysplastic syndromes and lowblast count acute myeloid leukemias. *Leukemia* 2011;25:1147e1152.
- Jabbour E, Kantarjian H. Chronic myeloid leukemia: 2014 update on diagnosis, monitoring, and management. *Am J Hematol* 2014;89:548–556.
- Jiang Y, Dunbar A, Gondek L, Gondek L, Mohan S, Rataul M, *et al.* Aberrant DNA methylation is a dominant mechanism in MDS progression to AML. *Blood* 2009;113:1315e1325.
- Jonker D, O'Callaghan C, Karapetis C, Zalcberg J, Tu D, Au H, *et al.* Cetuximab for the treatment of colorectal cancer. *N Engl J Med* 2007;357:2040–2048.
- Junttila T, Akita R, Parsons K, Fields C, Lewis P, Friedman L, *et al.* Ligand-independent HER2/HER3/PI3K complex is disrupted by trastuzumab and is effectively inhibited by the PI3K inhibitor GDC-0941. *Cancer Cell* 2009;15:429–440.
- Kantarjian H, Shah N, Hochhaus A, Cortes J, Shah S, Ayala M, *et al.* Dasatinib versus imatinib in newly diagnosed chronic-phase chronic myeloid leukemia. *N Engl J Med* 2010;362:2260–2270.
- Khoury H, Cortes J, Kantarjian H, Gambacorti-Passerini C, Baccarani M, Kim DW, *et al.* Bosutinib is active in chronic phase chronic myeloid leukemia after imatinib and dasatinib and/or nilotinib therapy failure. *Blood* 2012;119:3403–3412.
- Kim G, McKee A, Ning Y, Hazarika M, Theoret M, Johnson J, *et al.* FDA approval summary: vemurafenib for treatment of unresectable or metastatic melanoma with the *BRAF*V600E mutation. *Clin Cancer Res* 2014;20:4994–5000.
- Knickerbein K, and Zang L. Mutant KRAS as a critical determinant of the therapeutic response of colorectal cancer. *Genes Disease* 2015;2:4–12.
- Konecny G, Pegram M, Venkatesan N, Finn R, Yang G, Rahmeh M, *et al.* Activity of the dual kinase inhibitor lapatinib (GW572016) against HER-2-overexpressing and trastuzumab-treated breast cancer cells. *Cancer Res.* 2006;66:1630–1639.
- Kosaka T, Yatabe Y, Onozato, H, Kuwano H,

- Mitsudomi T. Prognostic implication of *EGFR*, *KRAS*, and *TP53* gene mutations in a large cohort of Japanese patients with surgically treated lung adenocarcinoma. *J Thorac Oncol* 2009;4:22–29.
- Ku CS, Cooper DN, Roukos DH. Clinical relevance of cancer genome sequencing. *World J Gastroenterol* 2013;19:2011–2018.
- Lavanya V, Mohamed Adil AA, Ahmed N, Rishi AK, Jamal S. Small molecule inhibitors as emerging cancer therapeutics. *Integr Cancer Sci Therap* 2014. 1:doi:10. 15761/ICST.1000109
- Lazarus R, David E. How and when to use genetic markers for non-small cell lung Cancer. *Curr Opin Pulm Med* 2013;19(4):331–339.
- Linnekin D. Early signaling pathways activated by c-Kit in hematopoietic cells. *Int J Biochem Cell Biol* 1999;31: 1053–1074.
- Machnicki M, Stoklosa T. BRAF, a new player in hematological neoplasms. *Blood Cells Mol Dis* 2014;53:77–83.
- Marcucci G, Byrd JC, Dai G, Klisovic P, Young D, Cataland S, *et al.* Phase I and pharmacokinetic studies of G3139, a Bcl-2 antisense oligonucleotide, in combination with chemotherapy in refractory or relapsed acute leukemia. *Blood* 2003;101:425–432.
- Meshinchi S, Alonzo T, Stirewalt D, Zwaan M, Zimmerman M, Reinhardt D, *et al.* Clinical implications of *FLT3* mutations in pediatric AML. *Blood* 2006;108:3654–3661.
- Muller M, Cortes J, Kim D, Druker B, Erben P, Pasquini R, *et al.* Dasatinib treatment of chronic-phase chronic myeloid leukemia: analysis of responses according to preexisting *BCR-ABL* mutations. *Blood* 2009;114:4944–4953.
- Neubauer H, Cumano A, Muller M, Wu H, Huffstadt U, Pfeffer K. Jak2 deficiency defines an essential developmental checkpoint in definitive hematopoiesis. *Cell* 1998;93:397–409.
- Novotny-Diermayr V, Hart S, Goh KC, Cheong A, Ong L, Hentze H, *et al.* The oral HDAC inhibitor pracinostat (SB939) is efficacious and synergistic with the JAK2 inhibitor pacritinib (SB1518) in preclinical models of AML. *Blood Cancer J* 2012;2:e69.
- O'Hare T, Shakespeare W, Zhu X, Eide C, Rivera V, Wang F, *et al.* AP24534, a pan-BCR-ABL inhibitor for chronic myeloid leukemia, potently inhibits the T315I mutant and overcomes mutation-based resistance. *Cancer Cell* 2009;16:401–412.
- Pao W, Chmielecki J. Rational, biologically based treatment of *EGFR*-mutant non-small-cell lung cancer. *Nat Rev Cancer* 2010;10:760–774.
- Pao W, Girard N. New driver mutations in non-small cell lung cancer. *The Lancet Oncology* 2011;12:175–180.
- Paschka P, Marcucci G, Ruppert A, Mrozek K, Chen H, Kittles R, *et al.* Adverse prognostic significance of KIT mutations in adult acute myeloid leukemia with inv (16) and t (8;21): A Cancer and Leukemia Group B study. *J Clin Oncol* 2006;24:3904–3911
- Pavlovi S, Zuki B, Petrovi M. Molecular genetic markers as a basis for personalized medicine. *Med Biochem* 2014; 33:8–21.
- Raponi M, Winkler H, Dracopoli N. K-RAS mutations predict response to *EGFR* inhibitors. *Curr Opin Pharmacol* 2008;8:413–

- 418.
- Rosenbauer F, Tenen D. Transcription factors in myeloid development: balancing differentiation with transformation. *Nat Rev Immunol* 2007;7:105–117.
- Sasaki T, Koivunen J, Ogino A, Yanagita M, Nikiforow S, Zheng W, *et al.* A novel ALK secondary mutation and EGFR signaling cause resistance to ALK kinase inhibitors. *Cancer Res* 2011;6051–6060.
- Sawyers CL. Rational therapeutic intervention in cancer: kinases as drug targets. *Curr Opin Gen Develop* 2002;12:111–115.
- Sethi, S, Kong D, Land, S, Dyson G, Sakr W, Sarkar F. Comprehensive molecular oncogenomic profiling and miRNA analysis of prostate cancer. *Am J Trans Res* 2013;5:200–211.
- Shaw A, Yeap B, Solomon B, Riely G, Gainor J, Engelman J, *et al.* Effect of crizotinib on overall survival in patients with advanced non-small-cell lung cancer harboring *ALK* gene rearrangement: a retrospective analysis. *Lancet Oncol* 2011;12:1004–1012.
- Shih AH, Abdel-Wahab O, Patel JP, Levine RL: The role of mutations in epigenetic regulators in myeloid malignancies. *Nat Rev Cancer* 2012;12:599e612.
- Shiota, M, Nakamura, S, Ichinohasama R, Ichinohasama R, Akagi T, Takeshita M, *et al.* Anaplastic large cell lymphomas expressing the novel chimeric protein p80NPM/ALK: a distinct clinicopathologic entity. *Blood* 1995;86:1954–1960.
- Shtivelman E, Lifshitz B, Gale RP, Canaani E. Fused transcript of *ABL1* and *BCR* genes in chronic myelogenous leukaemia. *Nature* 1985;4:550–554.
- Silverman GJ. Anti-CD20 therapy and autoimmune disease: therapeutic opportunities and evolving insights. *Front Biosci* 2007;12:2194–2206.
- Sjogren S, Inganas M, Lindgren A, Holmberg L, Bergh J. Prognostic and predictive value of c-erbB-2 overexpression in primary breast cancer, alone and in combination with other prognostic markers. *J Clin Oncol* 1998;16:462–469
- Soda M, Choi Y, Enomoto M, Takada S, Yamashita Y, Ishikawa S, *et al.* Identification of the transforming EML4-ALK fusion gene in non-small-cell lung cancer. *Nature* 2007;448(7153):561–566.
- Soverini S, Hochhaus A, Nicolini F, Gruber F, Lange T, Saglio G, *et al.* BCR-ABL kinase domain mutation analysis in chronic myeloid leukemia patients treated with tyrosine kinase inhibitors: recommendations from an expert panel on behalf of European Leukemia Net. *Blood* 2011;118:1208–1215.
- Staeher CF, Keller A, Leidinger P, Backes C, Chandran A, Wischhusen J, *et al.* Whole miRNome-wide differential co-expression of micro RNAs. *Genomics Proteomics Bioinformatics* 2012;10:285–294.
- Stirewalt D, Kopecky K, Meshinchi S, Engel J, Pogossova –Agadianvan E, Linsley J, *et al.* Size of FLT3 internal tandem duplication has prognostic significance in patients with acute myeloid leukemia. *Blood* 2006;107:3724–3726.
- Tsai J, Lee J, Wang W, Zhang J, Cho H, Mamo S, *et al.* Discovery of a selective inhibitor of oncogenic B-Raf kinase with potent antimelanoma activity. *Proc Natl Acad Sci*

- USA 2008;105:3041–3046.
- Turner N and Grose R. Fibroblast growth factor signaling from development to cancer. *Nat Rev Cancer* 2010;10:116–129.
- Vogel C, Cobleigh M, Tripathy D, Gutheil J, Harris L, Fehrenbacher L, *et al.* Efficacy and safety of trastuzumab as a single agent in first-line treatment of HER2-overexpressing metastatic breast cancer. *J Clin Oncol* 2002;20:719–726.
- Wang H, Lopategui J, Amin M, and Patterson S. KRAS mutation testing in human cancers: The pathologist's role in the era of personalized medicine. *Adv Anat Pathol* 2010;17(1):23–32.
- Weiss J, Sos M, Seidel D, Peifer M, Zander T, Heuckmann J, *et al.* Frequent and focal *FGFR1* amplification associates with therapeutically tractable *FGFR1* dependency in squamous cell lung cancer. *Science Translational Medicine* 2010;2:ID62ra93.
- Williams A, Li L, Nguyen B, Brown P, Levis M, Small D. Fluvastatin inhibits *FLT3* glycosylation in human and murine cells and prolongs survival of mice with *FLT3*-ITD leukemia. *Blood* 2012;120:3069–3079.
- Wu K, House L, Liu W, Cho WCS. Personalized targeted therapy for lung cancer. *Int J Mol Sci* 2012;13(9):11471–11496.
- Zhang S, Mantel C and Broxmeyer H. *FLT3* signaling involves tyrosyl-phosphorylation of SHP-2 and SHIP and their association with Grb2 and Shc in Baf3/*Flt3* cells. *J Leukoc Biol* 1999;65:372–380.

Cancer gene therapy: Prospects of using human sodium iodide symporter gene in non-thyroidal cancer

Shruti Dutta and Abhijit De*

Molecular Functional Imaging lab, Advanced Centre for Training, Research and Education in Cancer (ACTREC), Tata Memorial Centre, Navi Mumbai, India

Gene therapy is one of the promising therapeutic strategies evolved rapidly in the frontier of translational biology in cancer. To overcome the off target effect of conventional cancer therapies it is the most flourishing approach in present epoch. Various researches in this context are ongoing to eradicate devastating cancer cells with minimal or no side effects. Of the various gene therapy protocols developed, a set of genes called suicide genes, are being actively pursued as potential strategy. Briefly, this strategy involves tumor targeted delivery of a therapy/reporter gene to convert a systematically administered pro-drug into a cytotoxic drug which in turn induces tumor cell death. Additionally, advancement in small animal imaging modalities facilitates real-time monitoring of the delivered transgene by using appropriate imaging probe developed against the transgene. Non-invasive monitoring helps to realize precise transgene delivery and also aid to understand therapy response. In this background, we have reviewed potential suicide genes frequently explored for cancer treatment, which supports both diagnostic and therapeutic applications with special emphasis on sodium iodide symporter (NIS). Apart from its natural expression in thyroid, NIS protein expression has raised the possibility of using radioiodide therapy and diagnosis in few non-thyroidal cancers as well. In this review, we also covered various challenges to get NIS gene therapeutics from bench to bedside in various non-thyroidal cancers.

Gene Therapy for Cancer

With rapid advances in cellular and molecular understanding in the genome era, gene therapy holds great potential in treating various human diseases including cancer. The science behind gene therapy relies on introducing genes to cure or retard the progression of the disease. Theoretically, by introducing necessary

modifications for the mutated part(s) of a gene or by replacing the defective gene as a whole, one can potentially cure or retard the severity of a disease caused by the effect of a single gene. However, in reality, cancer is mostly considered as a multi-gene disorder (LaDuca *et al.*, 2014). Though many cancers have a genetic predisposition, a majority of them have

Key words: Sodium iodide symporter, Breast cancer, Histone deacetylase inhibitors, Radioiodine therapy, Molecular imaging.

***Corresponding Author:** Abhijit De, Molecular Functional Imaging lab, Advanced Centre for Training, Research and Education in Cancer (ACTREC), Tata Memorial Centre, Navi Mumbai, India.
Email: ade@actrec.gov.in

acquired mutations and chromosomal abnormalities. As the disease progress, the cells become less differentiated and more heterogeneous with respect to the mutations they carry. The range of genes involved or the mutations they carry have grown into a long-winded task for gene therapy field to solve. The ability to image the location(s), magnitude, and real-time variation of therapeutic gene expression has become a key force in the rapid evolution of gene therapy. Another concern in gene therapy application is the need to achieve controlled and effective gene expression in the target cells, with minimal or no off-target effect in neighboring normal tissue locations. To address this issue, *ex vivo* strategies help to ensure that gene transfer is limited to cells of a particular organ. For example, gene transfer into bone marrow cells provides a means to introduce genes selectively into various types of blood cells, including hematopoietic stem cells (Pan, 2009). However, ability of direct gene transfer to the appropriate cells via systemic delivery of a vector is often complicated, but appreciably enhances gene therapy approaches.

Coming to gene therapy of cancer, diverse applications reported in literatures

can be broadly categorized into five subgroups based on their mechanism of actions: (i) suicide gene therapy which uses introduction of drug-sensitive genes for selective induction of cellular sensitivity to a prodrug, (ii) protection of sensitive tissues like bone marrow from otherwise toxic doses of a cancer drug by use of multidrug resistance genes, (iii) replacement of lost or loss-of-function tumor suppressor genes, (iv) compensating down-regulated oncogene or gain-of function oncogenic mutation, and (v) insertion of a cytokine gene into tumor cells *ex vivo*. In this review, however, we want to focus on the first approach, where the gene or transgene signature can be utilized for killing the same cells where it is being expressed. We will continue our discussion analyzing gene therapy applications using sodium iodide symporter (NIS) in various non-thyroidal cancers.

Suicide Gene Therapy

Eventually the need of a cancer therapy is for complete remission of the cancer cells causing minimal damage to the surrounding normal tissues. In this regard the most promising approach is the targeted suicide gene therapy. With recent

advances in vector design, improvements in transgene (a new or altered gene that is being introduced) and prodrug activation strategies, suicide gene therapy is being applied to a wide variety of cancers (Zarogoulidis *et al.*, 2013). Till date various prokaryotic or eukaryotic genes have been tested as suicide gene therapy candidate, several of them also support diagnostic imaging to identify *in vivo* localization of the gene in action. For a particular therapeutic gene, diagnosis was done mostly by using gamma-ray emitting radionuclide probes while beta-ray emitting radionuclide probes are used for cancer cell diminution. In order to understand the consequences of the delivered transgene, non-invasive and real-time monitoring by using appropriate imaging modality is crucial. In this context reporter gene that supports radionuclide-based imaging approach gains attention as these imaging procedures are clinically relevant. With improved optical imaging instrumentations, the radionuclide based imaging applications has expanded scope now. Based on the phenomenon known as Cerenkov luminescence, emits visible luminescence photons when the charged radioactive particles travel through a dielectric media (such as tissue) at a speed higher than the speed of light, can be

captured in real time to understand the tissue bio-distribution of radiotracers (reviewed in Thorek *et al.*, 2012, Tanha *et al.*, 2015). There are at least three different types of interactions between the reporter proteins with their probes, which include an enzyme-based (e.g. Thymidine kinase and Cytosine deaminase), receptor-based (e.g. Somatostatin receptor) and transporter-based (e.g. NIS), used frequently for cancer gene therapy. We will now discuss ongoing research efforts utilizing these genes.

Herpes Simplex Virus type 1 Thymidine kinase (HSV-1 TK)

Among different suicide genes HSV-1 TK is the most frequently studied classical suicide gene target which converts non-toxic prodrug into a toxic drug. This viral TK phosphorylates various nucleoside analogues like aciclovir, ganciclovir, penciclovir much more efficiently than its mammalian homolog. Thus, mechanistically these cell-permeable substrates first get monophosphorylated by HSV-1 TK and subsequently phosphorylations are carried out by host kinase to generate the triphosphate form, which (deoxy-thymidine triphosphate) is basically a purine analog that inhibits DNA polymerase and therefore creates

toxicity to cause cell death eventually. Being highly proliferative in nature, cancer cells actively synthesize DNA, so the purine analog competes with guanosine triphosphates (GTPs) and get incorporated into the nascent DNA chain. As a result the nuclear and mitochondrial DNA synthesis terminates and cells are forced towards apoptosis. However, in transgenic mice the use of HSV-1 TK for tissue specific sensitization through ganciclovir treatment showed limitations. Due to high nuclear localization, the enzyme creates spermatozoal toxicity which in turn renders the male transgenic mice sterile (Cohen *et al.*, 1998). In another study investigators have revealed that due to the presence of putative cryptic testis-specific promoter within the coding sequence, the HSV-1 TK gene exhibited such outcome (Salomon *et al.*, 1995). Therefore to address this issue various studies have been carried out by generating different mutated version of HSV-1 TK with improved enzymatic activity and varying nuclear clearance (Ponomarev *et al.*, 2003). Among those HSV-1 sr39TK (also termed as HSV-1 tTK) was the most successful mutant (Black *et al.*, 2001) used recurrently for radionuclide imaging as well as suicide

gene therapy purpose. Imaging of HSV-1 tTK using ^{18}F -9-(4-[^{18}F] fluoro-3-hydroxymethylbutyl) guanine (^{18}F -FHBG) and ^{18}F - or ^{124}I -2'-deoxy-2'-fluoro-5-iodo-1- $[\beta]$ -D-arabino-furanosyluracil (^{18}F -FIAU and ^{124}I -FIAU) positron emission tomography (PET) turned it into a imaging reporter for clinical use. Later successful combining strategies by fusing HSV-1 sr39TK to other reporters made it suitable for multimode imaging (De *et al.*, 2003; Ray *et al.*, 2003; Ray *et al.*, 2004; Ruggiero *et al.*, 2010; Serganova *et al.*, 2008). In another study, a triple fusion gene construct using NIS, HSV1-sr39tk, and EGFP was developed and demonstrated its use as a suicide therapy agent in hepatocellular carcinoma (Lee *et al.*, 2010). However majority of such fusion gene using HSV-1 TK are used as imaging reporter rather than suicide therapy purpose. Another therapeutic study noteworthy here is the first-in-man use of engineered T-cells with HSV-1 TK-truncated CD34 fusion. This work highlights the suitability of tCD34 as a GMP compliant selection marker and demonstrates the feasibility, safety and immunological potential of HSVTK-tCD34 suicide gene modified donor T cells (Zhan *et al.*, 2013).

Cytosine deaminase (CD)

Cytosine deaminase protein is mainly synthesized in some bacteria and fungi, which deaminates cytosine to uracil. CD can also convert a non-toxic compound 5-fluorocytosine (5-FC) into the toxic compound 5-fluorouracil (5-FU) (Ramnaraine *et al.*, 2003). It exerts toxic effect by replacing for uracil in cellular RNA and therefore interferes with DNA and protein synthesis. Basically the absence of CD in mammalian tissues allows its use as a drug for suicide gene therapy for various cancer treatments. In an *in vitro* study genetically engineered stem cells to produce CD converts non-toxic 5-FC to a cytotoxic agent 5-FU and after migrating towards tumor site exhibited reduction in tumor growth (Kim *et al.*, 2010). Deamination of 5-FC prodrug produces two toxic metabolites such as 5-fluorodeoxyuridine monophosphate (FdUMP) and 5-fluorouridine triphosphate (FURTP). FdUMP being a potent inhibitor of thymidylate synthetase, an enzyme required for DNA synthesis, inhibits DNA synthesis and endorses apoptosis in tumor cells (Chen *et al.*, 2007; Myers, 1981). In the first attempt suicide gene therapy using CD was demonstrated significant tumor reduction in rat glioma

cells using 5-FC (Nishiyama *et al.*, 1985). Several other studies have also demonstrated positive effect of CD/5-FC as an effective anti-tumorigenic system for therapy in different other cancers (Li *et al.*, 2003; Lv *et al.*, 2009; Yang *et al.*, 2015). Improved tumor regression was obtained when radiation was combined with adenoviral mediated delivery of a mutated CD gene (Ad bCD-D314A) in pancreatic cancer (Kaliberova *et al.*, 2008).

Nitro-reductase (NTR)

Another promising, but relatively less used, prodrug activation enzyme used in cancer therapy is nitro-reductase (NTR). NTR is a flavoprotein synthesized by *Escherichia coli*. One of its substrates is CB1954 (5-[aziridin-1-yl]-2, 4-dinitrobenzamide), which reacts with cellular thioesters and get converted into a potent DNA cross-linking agent by NTR resulting in inhibition of DNA synthesis. Therefore viral mediated delivery of NTR in tumor cells get sensitized upon administration of CB1954, demonstrated the basis of cancer gene therapy (Searle *et al.*, 2004). An *in vitro* study showed that upon CB1954 treatment, NTR expressing clones (retro virus mediated delivery) of pancreatic and colorectal cancer cell lines

became 500 and 50 fold more sensitive than the parental cell line respectively (Green *et al.*, 1997). In preclinical tumor xenograft model, tumor regression using NTR/CD1954 was also reported (McNeish *et al.*, 1998; Weedon *et al.*, 2000).

Somatostatin receptor (SSTR)

Somatostatin (SST) is a peptide hormone involved in various biological processes in normal human tissues. When SST interacts with SSTRs, it exhibits strong anti-proliferative effect in normal and tumor cells. SSTRs are consisting of G-protein coupled receptor subtypes (1-5), differentially expressed in various tumor types. Among these subtypes SSTR1 and SSTR2 has been reported to have pivotal role in anti-proliferative effect. However, SSTR2 is majorly studied in gene therapy for cancer treatment (Schaer *et al.*, 1997). In most of the prostate cancer cases SSTR2 expression is found to be inactivated, so when the full length cDNA of SSTR2 was introduced using non-viral gene delivery system in the PC-3 cells, marked anti-angiogenic effect was reported (Kumar *et al.*, 2004). An *in vitro* study in pancreatic carcinoma also showed significant inhibition in cell proliferation upon

delivery of an adenoviral mediated MUC1-promoter expressing SSTR2, although there was no AdMUC1-SSTR2-induced apoptosis (Chen *et al.*, 2005). *In vivo* study also showed impairment of tumor progression upon delivery of SSTR2 gene in pancreatic cancer (Carrere *et al.*, 2005). Interestingly neuro-endocrine tumors (NETs) have natural over expression of SSTRs, so detection and treatment of NETs become easier by using SSTRs. ⁶⁸Gallium (⁶⁸Ga) labeled somatostatin analogue such as 1,4,7,10-tetraazacyclo-dodecane-1,4,7,10-tetraacetic acid (DOTA) was used to image NETs through PET/CT which was found to be more efficient than the conventional SSTR scintigraphy (Gabriel *et al.*, 2007; Sollini *et al.*, 2014). Moreover, other somatostatin analogues like ¹⁷⁷lutetium (¹⁷⁷Lu) or ⁹⁰yttrium (⁹⁰Y) are also found to have potential use in therapy for NET patients (Kam *et al.*, 2012; Sowa-Staszczak *et al.*, 2011).

Sodium iodide symporter (NIS)

Human NIS expression in various cancers like thyroidal and several non-thyroidal malignancies allow its worldwide use for theranostic application. Being a member of the solute carrier transporter (SLC5A5),

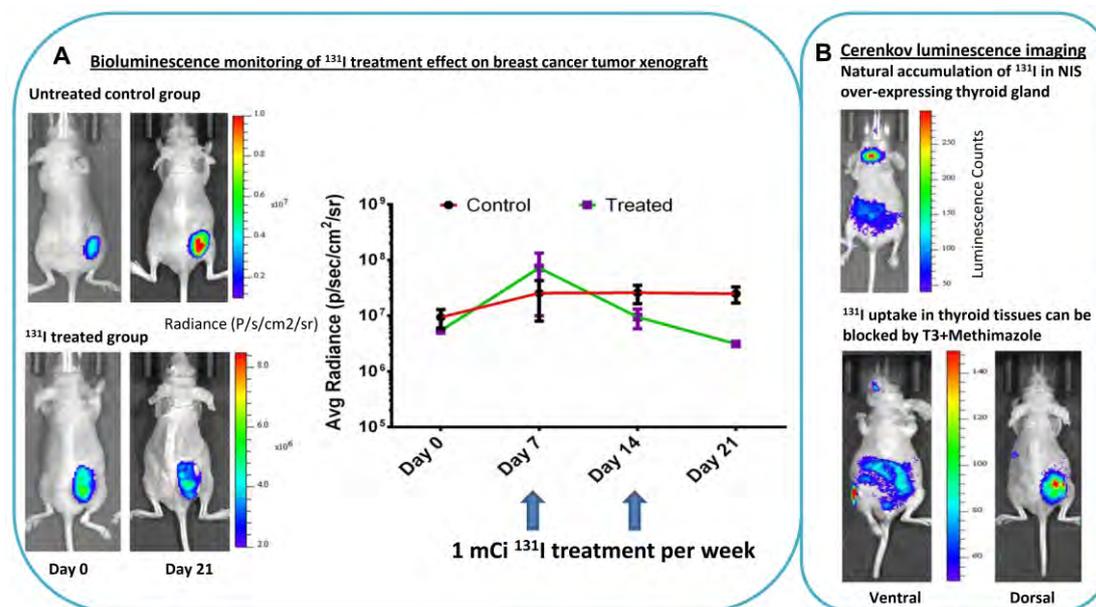


Figure 2: Optical luminescence imaging of breast cancer xenograft in mice undergoing ^{131}I treatment. A. Bioluminescence imaging to evaluate ^{131}I therapy in mice. These sets of mice were implanted with Zr75-1 breast cancer cells co-expressing human NIS and Firefly Luciferase reporter gene. After a good size of tumor develops, mice in the treatment group received weekly dose of 1 milliCurie/mouse ^{131}I on day 7 and 14 as indicated and monitored for attenuation in luciferase signal by injecting D-luciferin substrate. In comparison to the untreated control group, treated group shows significant diminution in tumor luciferase signal indicating NIS expressing tumor cell death. B. ^{131}I being β^- -emitter generates Cerenkov luminescence signal. This signal can be monitored using optical CCD camera. Representative mouse image on top shows natural accumulation of ^{131}I in thyroid gland 24 hours after 1 milliCurie radioiodine was injected intra-peritoneal. Bottom image shows ventral and dorsal view of a tumor bearing mouse in which breast cancer xenograft was placed dorsally on the right flank. This mouse received 14 days pre-treatment with triiodothyroxin (T3) and methimazole (an iodine organification inhibitor) daily, showing possibility of thyroid blocking without affecting the radioiodine uptake in breast tumor xenograft.

luminescence imaging (Thorek *et al.*, 2012; Xu *et al.*, 2012) from most of these particulate emitters (except for $^{99\text{m}}\text{TcO}_4$) supported by optical imaging modalities can be exploited (Fig. 2B). The ability of various radioisotopes producing Cerenkov radiation definitely add advantages such as low scan time requirement providing higher throughput and quantitative measurement of radioisotope distribution in the body, however the modality also suffers from issues such as lack of absolute

quantitative ability and significant signal attenuation with greater tissue depths.

Gene therapy applications using NIS can be broadly categorized into two streams, transgene-mediated and endogenous and we will now discuss various cancer application aspects in detail in the following sections.

Sodium Iodide Symporter as a Transgene Target for Cancer Treatment

Since, human NIS protein functions in a

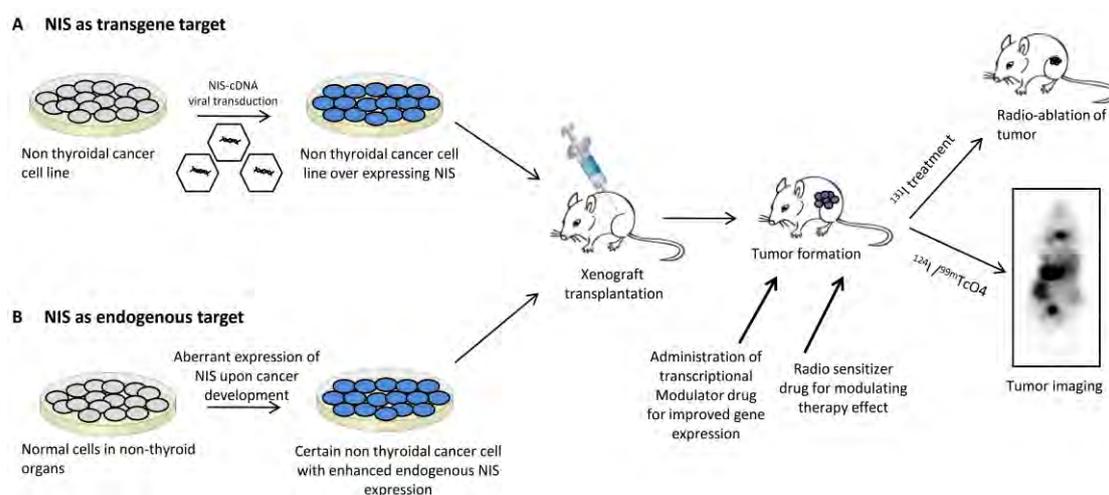


Figure 3: Schematic explaining various preclinical experimental set up for testing NIS gene mediated diagnosis and therapy in non-thyroidal cancers. A. NIS as transgene target is generally achieved by viral transduction methods in cancer cells. Such NIS over-expressing cells can be used for xenograft transplantation. Tumor formation can be visualized through appropriate imaging modality by administering $^{124}\text{I} / ^{99\text{m}}\text{TcO}_4$. Treatment with ^{131}I leads to tumor ablation. B. NIS as endogenous target is often recorded in certain cancer types. Various pharmacological modulators can be tested to improve endogenous NIS expression and/or function in these cancer cells *in vitro* or *in vivo* tumor xenograft. Radio-sensitization of tumor could further improve treatment efficacy though ^{131}I treatment.

very limited number of tissues in human body (primarily in thyroid, salivary gland, gastric mucosa and lactating breast), it is an attractive target to treat cancer in tissue types where NIS has no/aberrant expression. Among various non-thyroidal cancers NIS (isolated from both rat and human) transgene mediated targeted radioiodine therapy was studied primarily in prostate, colon, ovarian and breast cancers (BCs). Adenovirus and retrovirus mediated NIS gene transfer has been attempted specifically to these tumor cells and in some cases there *in vivo* efficacy was also tested (Fig. 3A). NIS gene delivery in prostate cancer has been thoroughly explored since last decades.

The first attempt of an *in vivo* adenovirus mediated hNIS (human NIS) delivery was done by Spitzweg et al. in non-thyroidal tumor xenograft. Significant NIS gene expression in prostate cancer xenograft was obtained followed by average reduction of tumor volume upon treatment with therapeutic ^{131}I (Spitzweg *et al.*, 2001). Later adenovirus mediated delivery of rat NIS (Ad-rNIS) was also reported in human prostate cancer tumor xenograft. The study also showed inhibition of tumor growth after intratumoral injection of Ad-rNIS followed by ^{131}I treatment (Spitzweg *et al.*, 2001). Therefore it is quite evident that rNIS has an equal potential compared to hNIS in suicide gene therapy for cancer

treatment. Further investigation has revealed that virus mediated NIS gene expression could be up-regulated through external beam radiation and DNA damage repair inhibitors (Hingorani *et al.*, 2008a; Hingorani *et al.*, 2008b). Additionally administration of radiosensitizing drugs further enhance therapeutic potential of NIS mediated ^{131}I in colorectal and head and neck cancer (Hingorani *et al.*, 2010). Baculovirus mediated NIS gene delivery in colon cancer cell line also showed efficient cell death through ^{131}I therapy (Yin *et al.*, 2010). In an *in vitro* study using ovarian cancer cell transduced with Ad/CMV/NIS and Ad/MUC1/NIS, 12- and 5-fold higher iodide uptake was reported compared to the parental cell line respectively. The same group has also shown reduction of ovarian tumor xenograft following ^{131}I treatment upon CMV/NIS infection of the tumors (Dwyer *et al.*, 2006).

On the other hand, in BC, aberrant over-expression of NIS is already reported; however the level of expression is often not sufficient for any therapeutic application. So, to achieve effective radioiodine therapy, investigators adopted strategy to selectively transfer NIS gene in breast cancer cell types. In one of such

studies engineered MCF7 cell line over expressing NIS and firefly luciferase was used to create mice xenograft and tested the effect of ^{131}I through bioluminescent imaging. The pattern of tumor regression upon ^{131}I administration in NIS expressing BC xenograft was also shown (Ghosh *et al.*, 2006). Another study has generated a conditional adenovirus vector containing MUC1 promoter driven E1a gene and a transcriptional cassette RSV promoter-hNIS in the E3 region that can replicate only in the MUC1 positive cells. Therefore, the engineered virus particles were transduced into MUC1 positive T47D breast cancer cell line showing improved radioiodine uptake (Trujillo *et al.*, 2009). Interestingly, virus mediated NIS transgene expression was shown to be further induced by treating cancer cells with retinoic acid (RA) which also showed subsequent enhancement in radioiodine uptake in MCF7 cell lines (Lim *et al.*, 2007). Further, non-replicative adenovirus was used for targeted NIS gene delivery by using promoters of human telomerase subunits RNA (hTR) and human telomerase reverse transcriptase (hTERT), which are active only in cancer cells (Riesco-Eizaguirre *et al.*, 2011).

Overcoming challenges of NIS gene delivery

Although viral mediated NIS gene delivery is mostly attempted but this approach poses some risks for human application. The virus particles having their natural tropism towards certain cell types may have a natural tendency to infect and integrate and/ or re-combine within the genome. For example, in an initial clinical trial for NIS gene therapy in X linked severe combined immuno-deficiency, an incidence of leukemia was reported due to unexpected integration of the viral DNA in the host genome (Kogai *et al.*, 2012). Therefore to avoid such unwanted DNA integration, all experimental NIS gene therapies are conducted with replication deficient viral vectors (Boland *et al.*, 2000; Hutzen *et al.*, 2012; Kim *et al.*, 2007). However, immunogenic reactions by the host, mutagenic integration (retroviral and lentiviral vectors), inflammatory toxicity (adenoviral vectors), and large scale production of the viral particles (adeno-associated vectors) are major limitations for such applications (Duarte *et al.*, 2012; Witlox *et al.*, 2007). So, current attempts are oriented towards non-viral vectors as vehicles for gene/drug delivery. Cationic

lipids, polymers, peptides and nanoparticles have commonly been used for DNA delivery into the cells in this regard (Chen *et al.*, 2015; Duarte *et al.*, 2012; Fan *et al.*, 2015;). Unlike viral vectors, targeting of non-viral vectors is a major concern and that could be improved by conjugating non-viral vectors with ligands which bind to specific receptors or antigens expressed on the cancer cells (Ogris *et al.*, 2002).

Sodium Iodide Symporter as an Endogenous Target for Cancer Therapy

Apart from introducing NIS gene in various cancer cells, ongoing active research has also exploited therapeutic potential of the gene where a specific cancer type is associated with aberrant over-expression of this gene. In such cases, since NIS protein is already present in the tumor cells, the burden of designing a vector for delivering the gene in the target cell is eliminated and thus can be spontaneously utilized for therapeutic intervention using radioiodine. Here, we have discussed various studies which have attempted ways to improve endogenous NIS expression and function for optimal therapeutic benefit (Fig. 3B). NIS is a true theranostic molecule which supports non-

invasive *in vivo* imaging using different form of radioactive iodine (PET, SPECT radiotracers) to judge sufficient iodine accumulation inside the tumor, which in turn indicates therapeutic success. Endogenous NIS gene therapy applications are primarily aimed at breast cancer. Apart from BC, modulation strategy to alter NIS aberrant expression was so far known from hepatocellular and testicular cancers (Guerrieri *et al.*, 2013; Maggisano *et al.*, 2014). In rat Leydig testicular carcinoma cells (LC540) treatment of HDAC inhibitors such as suberoylanilide hydroxamic acid (SAHA) and valproic acid (VPA) in combination showed enhanced NIS expression both in transcript and protein level with subsequent improvement of radioiodine uptake (Maggisano *et al.*, 2014). Evaluation was also done in liver cancer cells and correlated with p53 family member proteins showing increased NIS expression (Guerrieri *et al.*, 2013).

In the context of BC, of the various known subtypes, more than 80% hormone receptor positive cases (estrogen and progesterone receptor) were reported to have natural expression of NIS protein, while around 65% triple negative breast cancer (TNBC) cases showed positive

expression (Chatterjee *et al.*, 2013; Tazebay *et al.*, 2000; Wapnir *et al.*, 2003). Taking these results forward, clinical studies have been carried out to verify the translational potential of NIS in malignant BC patients. Moon *et al.* (2001) reported significant $^{99m}\text{TcO}_4^-$ uptake in 4 out of 25 breast cancer patients by scintigraphic scanning method. Another important study by Wapnir *et al.* detected radioiodine uptake in metastatic BC by scintigraphic analysis using radioiodides (^{123}I and ^{131}I) or $^{99m}\text{TcO}_4^-$. However, surprisingly iodine uptake was noted in only 25% (2 out of 8) of NIS positive metastatic breast tissues (Renier *et al.*, 2009; Wapnir *et al.*, 2004). Therefore, current focus on achieving effective NIS gene therapy is to find potent modulators that could improve endogenous NIS expression and/ or function.

Overcoming challenges of NIS gene therapy

As mentioned above, exploiting endogenous NIS as a therapeutic target is limited by its ability to pump sufficient iodine inside cells, especially in non-thyroidal cancer tissues such as breast. This discordant is primarily due to lack of NIS expression on plasma membrane,

which is critical for its iodine transporter function delivering iodine inside the cell. In this regard, epidermal growth factor (EGF) was identified to localized NIS better on the plasma membrane in non-thyroidal cancers, while reverse localization was also obtained by treatment with a MEK-1 inhibitor, suggesting involvement of MEK-ERK signaling pathway in NIS localization (). Moreover, as far as improvement of NIS endogenous expression is concerned, trans-retinoic acid (tRA) was frequently studied as the major inductor in BC. Treatment with tRA showed significant increase in ^{131}I mediated radioablation in MCF7 cell lines (Kogai *et al.*, 2012). Besides, *in vivo* study using MCF7 xenograft mouse model showed enhanced radioiodine uptake sufficient for effective cell killing (Kogai *et al.*, 2004). But due to short biological half-life, the effect of tRA exists in the system only for a limited time period and therefore requires frequent use during therapeutic intervention. However, frequent use of tRA caused cardio-respiratory distress syndrome in patients of acute promyelocytic leukemia (Warrell, 1993). 13-cis RA can be used as a prodrug which finally gets converted into tRA inside the target cells. Experimental study

has shown that although it induces NIS expression, the level of expression is much lower than tRA administration (Kogai *et al.*, 2005). RAs operate by binding through two families of nuclear receptors, retinoic acid receptors (RARs), which are activated by both all-trans RA and 9-cis-RA, and retinoid X receptors (RXRs), which are activated by 9-cis-RA only. Upon binding of RAs, receptors get activated and bind to RA responsive elements in the promoter regions of target genes and work as ligand-dependent transcription factors (Alotaibi *et al.*, 2010).

Primarily, NIS activation takes place by binding of RA to RAR α /RXR β receptor hetero-dimer followed by activation of the phosphoinositide 3-kinase (PI3K) pathway and the p38 β MAPK pathway (Kogai *et al.*, 2012). By administering PI3K inhibitor or by knock down of p85 α (a regulatory subunit of PI3K) showed decreased RA induced NIS expression in MCF7 cells. Decrease in iodide uptake was also reported upon inhibition of AKT pathway in MCF7 cell lines (Kogai *et al.*, 2012) suggesting involvement of PI3K/AKT pathway in NIS induction. Further, combination of tRA with several pharmacological compounds found to be

more effective in NIS gene induction in non-thyroidal cancers. Such combination therapies used drugs like hydrocortisone, dexamethasone (Dex) (Dohan *et al.*, 2006; Kogai *et al.*, 2005), troglitazone (a peroxisome proliferator-activated receptor γ , PPAR γ , agonist) (Tanosaki *et al.*, 2003; Wei *et al.*, 2009), various histone deacetylase (HDAC) inhibitors (trichostatin A and sodium butyrate), and carbamazepine, an agonist of pregnane X receptor (Unterholzner *et al.*, 2006; Willhauck *et al.*, 2011). Some other compounds like prolactin, insulin, and insulin growth factor (IGF)-I and II were also used to stimulate NIS mRNA expression in BC cell line (Arturi *et al.*, 2005).

Moreover, the efficiency of radioiodine therapy in various cancers not only depends on NIS protein expression and its membrane localization, the factor that can't be ignored is biological half-life and retention potential of radioiodine in the tumor bed. In thyroid gland, due to iodine organification the biological half-life of iodine is sufficiently long, making radioiodine therapeutics effective for thyroid cancer treatment (Shimura *et al.*, 1997). However in lactating mammary gland approximately 20% of the iodine

was trapped due to iodine oxidation by lactoperoxidase (LPO) expressed in alveolar cells followed by binding of iodine to various milk proteins (Etling *et al.*, 1984; Strum *et al.*, 1983). But this may be insufficient for radioiodine therapy in BC. So, studies are underway to improve iodine retention time in breast cancer cells for getting ^{131}I therapy effective. Combination treatment with all-trans RA and Dex has shown modest improvement in iodine retention in MCF7 cells. However, the exact mechanism is not known (Unterholzner *et al.*, 2006).

Furthermore, as NIS is highly expressed in normal thyroid tissues, while treating non-thyroidal cancers safeguarding thyrocytes can also be a major issue. However, this issue has already been addressed by selective down-regulation of NIS expression in thyrocytes by administering T3 and methimazole in combination (Wapnir *et al.*, 2004). Apart from thyroid and lactating mammary gland NIS is also expressed in several other normal tissues such as salivary gland, intestinal epithelium, lacrimal gland, stomach lining etc. that become barrier for NIS based imaging and treatment. Uptake in these normal tissues not only reduces therapeutic efficiency but

subsequently radioiodine damages of normal organ function as well.

Clinical Experiences with NIS

NIS based diagnosis and therapy is well known in thyroid clinics for several decades to identify and treat various thyroid diseases including cancer. In a recent case study after complete thyroidectomy in differentiated thyroid cancer patient, whole-body scans based on diagnostic or therapeutic doses of ^{131}I can visualize various distant metastatic lesions. Nonetheless, extreme precautionary measure is required while analyzing the data because several false positive signals were also obtained (Ahn *et al.*, 2011). As per the NIH database (www.clinicaltrials.gov), at present there is no ongoing or completed clinical trials on NIS based therapy in non-thyroidal cancers. The reasons are indeed due to several such issues that are discussed above. So, investigators are actively investigating effective methods of modulation to enhance NIS expression and localization on plasma membrane to achieve the optimal efficacy. However, recently few clinical trials in prostate and ovarian cancer are attempted for NIS gene therapy. In a phase I clinical trial, prostate

cancer patients received intra-prostatic injection of Ad5-yCD/mutTK(sr39)rep-hNIS followed by measuring hNIS expression over time through SPECT imaging after adenovirus injection. Positive hNIS expression was obtained in the patient's prostate gland suggesting non-invasive imaging of NIS gene is achievable and safe for humans (Barton *et al.*, 2008; 2011). Moreover this study also proved the application of ^{131}I in human for localized prostate cancer treatment. Recently a clinical study in ovarian cancer also showed promising observation upon intra-peritoneal administration of engineered measles virus to express NIS. NIS expression was confirmed in patient's tumor through ^{123}I uptake using SPECT/CT scan (Galanis *et al.*, 2015). Further clinical evaluations are expected in the coming years to achieve successful NIS gene therapy application in non-thyroidal cancers.

CONCLUSIONS

Gene therapy in cancer has high potential because of recent advancement in genetic engineering in cellular and molecular level. In this arena, NIS gene gains importance in cancer for gene therapy because of its property to serve both

diagnostic imaging and therapeutics. Moreover to improve therapeutic effectiveness, NIS gene therapy is rapidly evolving in various non-thyroidal cancers particularly in BC because of lack of appropriate therapeutic options in hormone receptor negative patients. Although NIS gene therapy is already on track for various thyroid cancers, but for non-thyroidal cancers there are still various logistics that need to be addressed before successful clinical translation. Major concerns which need to be taken care are the expression level, protein localization on cell membrane and lower retention time of radioiodine in cancer tissues. So, optimal strategies are yet to be developed to improve radioiodine uptake and retention by modulating NIS gene expression in non-thyroidal cancers.

REFERENCES

- Ahn BC, Lee SW, Lee J, Kim C. Pulmonary aspergilloma mimicking metastasis from papillary thyroid cancer. *Thyroid* 2011;21: 555–558.
- Alotaibi H, Yaman E, Salvatore D, Di Dato V, Telkoparan P, Di Lauro R, *et al.* Intronic elements in the Na⁺/I⁻ symporter gene (NIS) interact with retinoic acid receptors and mediate initiation of transcription. *Nucleic Acids Res* 2010;38:3172–3185.
- Arturi F, Ferretti E, Presta I, Mattei T, Scipioni A, Scarpelli D, *et al.* Regulation of iodide uptake and sodium/iodide symporter expression in the mcf-7 human breast cancer cell line. *J Clin Endocrinol Metab* 2005;90:2321–2326.
- Barton KN, Stricker H, Brown SL, Elshaikh M, Aref I, Lu M, *et al.*, Phase I study of noninvasive imaging of adenovirus-mediated gene expression in the human prostate. *Mol Ther* 2008;16:1761–1769.
- Barton KN, Stricker H, Elshaikh MA, Pegg J, Cheng J, Zhang Y, *et al.* Feasibility of adenovirus-mediated hNIS gene transfer and ¹³¹I radioiodine therapy as a definitive

Expectedly, the progression in recent basic research unraveling NIS biology in the field of gene therapy would develop right strategies of treatment to care devastating cancer.

ACKNOWLEDGMENTS

Funding support from TMC-Woman Cancer Initiative funding (#82) and ICMR, New Delhi (Ref. No. 5/13/25/10/NCD-III) to AD and DST-WOSA award to SD is gratefully acknowledged. For some parts of the figure materials, we also acknowledge the work done by Sushmita Chatterjee in the lab.

CONFLICT OF INTEREST

The authors claim no conflict of interest.

- treatment for localized prostate cancer. *Mol Ther* 2011;19:1353–1359.
- Black ME, Kokoris MS, Sabo P. Herpes simplex virus-1 thymidine kinase mutants created by semi-random sequence mutagenesis improve prodrug-mediated tumor cell killing. *Cancer Res* 2001;61:3022–3026.
- Boland A, Ricard M, Opolon P, Bidart JM, Yeh P, Filetti S, *et al.* Adenovirus-mediated transfer of the thyroid sodium/iodide symporter gene into tumors for a targeted radiotherapy. *Cancer Res* 2000;60:3484–3492.
- Carrere N, Vernejoul F, Souque A, Asnacios A, Vaysse N, Pradayrol L, *et al.* Characterization of the bystander effect of somatostatin receptor sst2 after in vivo gene transfer into human pancreatic cancer cells. *Hum Gene Ther* 2005;16:1175–1193.
- Chatterjee S, Malhotra R, Varghese F, Bukhari AB, Patil A, Budrukkar A, *et al.* Quantitative immunohistochemical analysis reveals association between sodium iodide symporter and estrogen receptor expression in breast cancer. *PLoS One* 2013;8:e54055.
- Chen JK, Hu LJ, Wang D, Lamborn KR, Deen DF. Cytosine deaminase/5-fluorocytosine exposure induces bystander and radiosensitization effects in hypoxic glioblastoma cells in vitro. *Int J Radiat Oncol Biol Phys* 2007;67:1538–1547.
- Chen L, Liu Q, Qin R, Le H, Xia R, Li W, *et al.* Amplification and functional characterization of MUC1 promoter and gene-virotherapy via a targeting adenoviral vector expressing hSSTR2 gene in MUC1-positive Panc-1 pancreatic cancer cells in vitro. *Int J Mol Med* 2005;15:617–626.
- Chen Y, Nan J, Lu Y, Wang C, Chu F, Gu Z. Hybrid Fe₃O₄-Poly(acrylic acid) Nanogels for Theranostic Cancer Treatment. *J Biomed Nanotechnol* 2015;11:771–779.
- Cohen JL, Boyer O, Salomon B, Onclercq R, Depetris D, Lejeune L, *et al.* Fertile homozygous transgenic mice expressing a functional truncated herpes simplex thymidine kinase delta TK gene. *Transgenic Res* 1998;7:321–330.
- De A, Lewis XZ, Gambhir SS. Noninvasive imaging of lentiviral-mediated reporter gene expression in living mice. *Mol Ther* 2003;7:681–691.
- Dohan O, De la Vieja A, Carrasco N. Hydrocortisone and purinergic signaling stimulate sodium/iodide symporter (NIS)-mediated iodide transport in breast cancer cells. *Mol Endocrinol* 2006;20:1121–37.
- Duarte S, Carle G, Faneca H, de Lima MC, Pierrefite-Carle V. Suicide gene therapy in cancer: where do we stand now? *Cancer Lett* 2012;324:160–170.
- Dwyer RM, Bergert ER, O'Connor MK, Gendler SJ, Morris JC. Sodium iodide symporter-mediated radioiodide imaging and therapy of ovarian tumor xenografts in mice. *Gene Ther* 2006;13:60–66.
- Etling N, Gehin-Fouque F. Iodinated compounds and thyroxine binding to albumin in human breast milk. *Pediatr Res* 1984;18:901–903.
- Fan Y, Moon JJ. Nanoparticle drug delivery systems designed to improve cancer vaccines and immunotherapy. *Vaccines (Basel)* 2015;3:662–685.
- Gabriel M, Decristoforo C, Kendler D, Dobrozemsky G, Heute D, Uprimny C, *et al.*

- 68Ga-DOTA-Tyr3-octreotide PET in neuroendocrine tumors: comparison with somatostatin receptor scintigraphy and CT. *J Nucl Med* 2007;48:508–518.
- Galanis E, Atherton PJ, Maurer MJ, Knutson KL, Dowdy SC, Cliby WA, *et al.* Oncolytic measles virus expressing the sodium iodide symporter to treat drug-resistant ovarian cancer. *Cancer Res* 2015;75:22–30.
- Ghosh M, Gambhir SS, De A, Nowels K, Goris M, Wapnir I. Bioluminescent monitoring of NIS-mediated (131)I ablative effects in MCF-7 xenografts. *Mol Imaging* 2006;5:76–84.
- Green NK, Youngs DJ, Neoptolemos JP, Friedlos F, Knox RJ, Springer CJ, *et al.* Sensitization of colorectal and pancreatic cancer cell lines to the prodrug 5-(aziridin-1-yl)-2,4-dinitrobenzamide (CB1954) by retroviral transduction and expression of the E. coli nitroreductase gene. *Cancer Gene Ther* 1997;4:229–238.
- Guerrieri F, Piconese S, Lacoste C, Schinzari V, Testoni B, Valogne Y, *et al.* The sodium/iodide symporter NIS is a transcriptional target of the p53-family members in liver cancer cells. *Cell Death Dis* 2013;4:e807.
- Hingorani M, White CL, Merron A, Peerlinck I, Gore ME, Slade A, *et al.* Inhibition of repair of radiation-induced DNA damage enhances gene expression from replication-defective adenoviral vectors. *Cancer Res* 2008a;68:9771–9778.
- Hingorani M, White CL, Zaidi S, Merron A, Peerlinck I, Gore ME, *et al.*, Radiation-mediated up-regulation of gene expression from replication-defective adenoviral vectors: implications for sodium iodide symporter gene therapy. *Clin Cancer Res* 2008b;14:4915–4924.
- Hingorani M, White CL, Zaidi S, Pandha HS, Melcher AA, Bhide SA, *et al.* Therapeutic effect of sodium iodide symporter gene therapy combined with external beam radiotherapy and targeted drugs that inhibit DNA repair. *Mol Ther* 2010;18:1599–1605.
- Hutzen B, Pierson CR, Russell SJ, Galanis E, Raffel C, Studebaker AW. Treatment of medulloblastoma using an oncolytic measles virus encoding the thyroidal sodium iodide symporter shows enhanced efficacy with radioiodine. *BMC Cancer* 2012;12:508.
- Jung KH, Paik JY, Ko BH, Lee KH. Mitogen-activated protein kinase signaling enhances sodium iodide symporter function and efficacy of radioiodide therapy in nonthyroidal cancer cells. *J Nucl Med* 2008;49:1966–1972.
- Kaliberova LN, Della Manna DL, Krendelchtchikova V, Black ME, Buchsbaum DJ, Kaliberov SA. Molecular chemotherapy of pancreatic cancer using novel mutant bacterial cytosine deaminase gene. *Mol Cancer Ther* 2008;7:2845–2854.
- Kam BL, Teunissen JJ, Krenning EP, de Herder WW, Khan S, van Vliet EI, *et al.* Lutetium-labelled peptides for therapy of neuroendocrine tumours. *Eur J Nucl Med Mol Imaging* 2012;39 Suppl 1:S103–112.
- Kim HJ, Jeon YH, Kang JH, Lee YJ, Kim KI, Chung HK, *et al.* In vivo long-term imaging and radioiodine therapy by sodium-iodide symporter gene expression using a lentiviral system containing ubiquitin C promoter. *Cancer Biol Ther* 2007;6:1130–1135.
- Kim KY, Kim SU, Leung PC, Jeung EB, Choi KC. Influence of the prodrugs 5-fluorocytosine and

- CPT-11 on ovarian cancer cells using genetically engineered stem cells: tumor-tropic potential and inhibition of ovarian cancer cell growth. *Cancer Sci* 2010;101:955–962.
- Kogai T, Kanamoto Y, Che LH, Taki K, Moatamed F, Schultz JJ, *et al.* Systemic retinoic acid treatment induces sodium/iodide symporter expression and radioiodide uptake in mouse breast cancer models. *Cancer Res* 2004;64:415–422.
- Kogai T, Kanamoto Y, Li AI, Che LH, Ohashi E, Taki K, *et al.* Differential regulation of sodium/iodide symporter gene expression by nuclear receptor ligands in MCF-7 breast cancer cells. *Endocrinology* 2005;146:3059–3069.
- Kogai T, Brent GA, The sodium iodide symporter (NIS): regulation and approaches to targeting for cancer therapeutics. *Pharmacol Ther* 2012;135:355–370.
- Kumar M, Liu ZR, Thapa L, Chang Q, Wang DY, Qin RY. Antiangiogenic effect of somatostatin receptor subtype 2 on pancreatic cancer cell line: Inhibition of vascular endothelial growth factor and matrix metalloproteinase-2 expression in vitro. *World J Gastroenterol* 2004;10:393–399.
- LaDuca H, Stuenkel AJ, Dolinsky JS, Keiles S, Tandy S, Pesaran T, *et al.* Utilization of multigene panels in hereditary cancer predisposition testing: analysis of more than 2,000 patients. *Genet Med* 2014;16:830–837.
- Lee YL, Lee YJ, Ahn SJ, Choi TH, Moon BS, Cheon GJ, *et al.* Combined radionuclide-chemotherapy and in vivo imaging of hepatocellular carcinoma cells after transfection of a triple-gene construct, NIS, HSV1-sr39tk, and EGFP. *Cancer Lett* 2010;290:129–138.
- Li ZS, Pan X, Xu GM, Cui L, Dai GR, Gong YF, *et al.* Killing effects of cytosine deaminase gene mediated by adenovirus vector on human pancreatic cancer cell lines in vitro. *Hepatobiliary Pancreat Dis Int* 2003;2:147–151.
- Lim SJ, Paeng JC, Kim SJ, Kim SY, Lee H, Moon DH. Enhanced expression of adenovirus-mediated sodium iodide symporter gene in MCF-7 breast cancer cells with retinoic acid treatment. *J Nucl Med* 2007;48:398–404.
- Lv SQ, Zhang KB, Zhang EE, Gao FY, Yin CL, Huang CJ, *et al.* Antitumor efficiency of the cytosine deaminase/5-fluorocytosine suicide gene therapy system on malignant gliomas: an *in vivo* study. *Med Sci Monit* 2009;15:BR13–20.
- Maggisano V, Puppini C, Celano M, D'Agostino M, Sponziello M, Micali S, *et al.* Cooperation of histone deacetylase inhibitors SAHA and valproic acid in promoting sodium/iodide symporter expression and function in rat Leydig testicular carcinoma cells. *Endocrine* 2014;45:148–152.
- McNeish IA, Green NK, Gilligan MG, Ford MJ, Mautner V, Young LS, *et al.* Virus directed enzyme prodrug therapy for ovarian and pancreatic cancer using retrovirally delivered *E. coli* nitroreductase and CB1954. *Gene Ther* 1998;5:1061–1069.
- Mitrofanova E, Unfer R, Vahanian N, Daniels W, Roberson E, Seregina T, *et al.* Rat sodium iodide symporter for radioiodide therapy of cancer. *Clin Cancer Res* 2004;10:6969–6976.
- Moon DH, Lee SJ, Park KY, Park KK, Ahn SH, Pai

- MS, *et al.* Correlation between ^{99m}Tc -pertechnetate uptakes and expressions of human sodium iodide symporter gene in breast tumor tissues. *Nucl Med Biol* 2001;28: 829–834.
- Myers CE. The pharmacology of the fluoropyrimidines. *Pharmacol Rev* 1981;33: 1–15.
- Nimesh S. Potential implications of nanoparticle characterization on in vitro and in vivo gene delivery. *Ther Deliv* 2012;3:1347–1356.
- Nishiyama T, Kawamura Y, Kawamoto K, Matsumura H, Yamamoto N, Ito T, *et al.*, Antineoplastic effects in rats of 5-fluorocytosine in combination with cytosine deaminase capsules. *Cancer Res* 1985;45: 1753–1761.
- Ogris M, Wagner E. Targeting tumors with non-viral gene delivery systems. *Drug Discov Today* 2002;7:479–485.
- Pan D. *In situ (in vivo)* gene transfer into murine bone marrow stem cells. *Methods Mol Biol* 2009;506:159–169.
- Ponomarev V, Doubrovin M, Serganova I, Beresten T, Vider J, Shavrin A, *et al.* Cytoplasmically retargeted HSV1-tk/GFP reporter gene mutants for optimization of noninvasive molecular-genetic imaging. *Neoplasia* 2003;5:245–254.
- Ramnaraine M, Pan W, Goblirsch M, Lynch C, Lewis V, Orchard P, *et al.* Direct and bystander killing of sarcomas by novel cytosine deaminase fusion gene. *Cancer Res* 2003;63: 6847–6854.
- Ray P, Wu AM, Gambhir SS. Optical bioluminescence and positron emission tomography imaging of a novel fusion reporter gene in tumor xenografts of living mice. *Cancer Res* 2003;63:1160–1165.
- Ray P, De A, Min JJ, Tsien RY, Gambhir SS. Imaging tri-fusion multimodality reporter gene expression in living subjects. *Cancer Res* 2004; 64:1323–1330.
- Renier C, Yao C, Goris M, Ghosh M, Katznelson L, Nowles K, *et al.* Endogenous NIS expression in triple-negative breast cancers. *Ann Surg Oncol* 2009;16:962–968.
- Riesco-Eizaguirre G, De la Vieja A, Rodriguez I, Miranda S, Martin-Duque P, Vassaux G, *et al.* Telomerase-driven expression of the sodium iodide symporter (NIS) for in vivo radioiodide treatment of cancer: a new broad-spectrum NIS-mediated antitumor approach. *J Clin Endocrinol Metab* 2011;96:E1435–1443.
- Ruggiero A, Brader P, Serganova I, Zanzonico P, Cai S, Lipman NS, *et al.* Different strategies for reducing intestinal background radioactivity associated with imaging HSV1-tk expression using established radionucleoside probes. *Mol Imaging* 2010;9:47–58.
- Salomon B, Maury S, Loubiere L, Caruso M, Onclercq R, Klatzmann D. A truncated herpes simplex virus thymidine kinase phosphorylates thymidine and nucleoside analogs and does not cause sterility in transgenic mice. *Mol Cell Biol* 1995;15:5322–5328.
- Schaer JC, Waser B, Mengod G, Reubi JC. Somatostatin receptor subtypes sst1, sst2, sst3 and sst5 expression in human pituitary, gastroentero-pancreatic and mammary tumors: comparison of mRNA analysis with receptor autoradiography. *Int J Cancer* 1997;70: 530–537.
- Searle PF, Chen MJ, Hu L, Race PR, Lovering AL, Grove JJ, *et al.* Nitroreductase: a prodrug-

- activating enzyme for cancer gene therapy. *Clin Exp Pharmacol Physiol* 2004;31:811–816.
- Serganova I, Mayer-Kukuck P, Huang R, Blasberg R. Molecular imaging: reporter gene imaging. *Handb Exp Pharmacol* 2008;167–223.
- Shimura H, Haraguchi K, Miyazaki A, Endo T, Onaya T. Iodide uptake and experimental ¹³¹I therapy in transplanted undifferentiated thyroid cancer cells expressing the Na⁺/I⁻ symporter gene. *Endocrinology* 1997;138:4493–4496.
- Sollini M, Erba PA, Fraternali A, Casali M, Di Paolo ML, Froio A, *et al.* PET and PET/CT with ⁶⁸Ga-labeled somatostatin analogues in Non GEP-NETs Tumors. *Scientific World Journal* 2014;2014:194123.
- Sowa-Staszczak A, Pach D, Kunikowska J, Krolicki L, Stefanska A, Tomaszuk M, *et al.* Efficacy and safety of ⁹⁰Y-DOTATATE therapy in neuroendocrine tumours. *Endokrynol Pol* 2011;62:392–400.
- Spitzweg C, Dietz AB, O'Connor MK, Bergert ER, Tindall DJ, Young CY, *et al.* *In vivo* sodium iodide symporter gene therapy of prostate cancer. *Gene Ther* 2001;8:1524–1531.
- Strum JM, Phelps PC, McAtee MM. Resting human female breast tissue produces iodinated proteins. *J Ultrastruct Res* 1983;84:130–139.
- Tanosaki S, Ikezoe T, Heaney A, Said JW, Dan K, Akashi M, *et al.* Effect of ligands of nuclear hormone receptors on sodium/iodide symporter expression and activity in breast cancer cells. *Breast Cancer Res Treat* 2003;79: 335–345.
- Tazebay UH, Wapnir IL, Levy O, Dohan O, Zuckier LS, Zhao QH, *et al.* The mammary gland iodide transporter is expressed during lactation and in breast cancer. *Nat Med* 2000;6: 871–878.
- Thorek D, Robertson R, Bacchus WA, Hahn J, Rothberg J, Beattie BJ, *et al.* Cerenkov imaging - a new modality for molecular imaging. *Am J Nucl Med Mol Imaging* 2012;2:163–173.
- Trujillo MA, Oneal MJ, Davydova J, Bergert E, Yamamoto M, Morris JC, 3rd. Construction of an MUC-1 promoter driven, conditionally replicating adenovirus that expresses the sodium iodide symporter for gene therapy of breast cancer. *Breast Cancer Res* 2009;11:R53.
- Unterholzner S, Willhauck MJ, Cengic N, Schutz M, Goke B, Morris JC, *et al.* Dexamethasone stimulation of retinoic Acid-induced sodium iodide symporter expression and cytotoxicity of ¹³¹I in breast cancer cells. *J Clin Endocrinol Metab* 2006;91:69–78.
- Wapnir IL, van de Rijn M, Nowels K, Amenta PS, Walton K, Montgomery K, *et al.* Immunohistochemical profile of the sodium/iodide symporter in thyroid, breast, and other carcinomas using high density tissue microarrays and conventional sections. *J Clin Endocrinol Metab* 2003;88:1880–1888.
- Wapnir IL, Goris M, Yudd A, Dohan O, Adelman D, Nowels K, *et al.* The Na⁺/I⁻ symporter mediates iodide uptake in breast cancer metastases and can be selectively down-regulated in the thyroid. *Clin Cancer Res* 2004; 10:4294–4302.
- Warrell RP, Jr. Retinoid resistance in acute promyelocytic leukemia: new mechanisms, strategies, and implications. *Blood* 1993;82: 1949–1953.
- Weedon SJ, Green NK, McNeish IA, Gilligan MG, Mautner V, Wrighton CJ, *et al.* Sensitisation of human carcinoma cells to the prodrug CB1954 by adenovirus vector-mediated expression of *E*.

- coli* nitroreductase. *Int J Cancer* 2000;86: 848–854.
- Wei S, Yang J, Lee SL, Kulp SK, Chen CS. PPARgamma-independent antitumor effects of thiazolidinediones. *Cancer Lett* 2009;276: 119–124.
- Willhauck MJ, DJ OK, Wunderlich N, Goke B, Spitzweg C. Stimulation of retinoic acid-induced functional sodium iodide symporter (NIS) expression and cytotoxicity of (1)(3)(1)I by carbamazepine in breast cancer cells. *Breast Cancer Res Treat* 2011;125:377–386.
- Witlox MA, Lamfers ML, Wuisman PI, Curiel DT, Siegal GP. Evolving gene therapy approaches for osteosarcoma using viral vectors: review. *Bone* 2007;40:797–812.
- Xu Y, Liu H, Chang E, Jiang H, Cheng Z. Cerenkov Luminescence Imaging (CLI) for cancer therapy monitoring. *J Vis Exp* 2012;e4341.
- Yang XP, Liu L, Wang P, Ma SL. Human Sulfatase-1 Improves the Effectiveness of Cytosine Deaminase Suicide Gene Therapy with 5-Fluorocytosine Treatment on Hepatocellular Carcinoma Cell Line HepG2 and. *Chin Med J (Engl)* 2015;128:1384–1390.
- Yin HY, Zhou X, Wu HF, Li B, Zhang YF. Baculovirus vector-mediated transfer of NIS gene into colon tumor cells for radionuclide therapy. *World J Gastroenterol* 2010;16: 5367–5374.
- Zarogoulidis P, Darwiche K, Sakkas A, Yarmus L, Huang H, Li Q, *et al.* Suicide Gene Therapy for Cancer - Current Strategies. *J Genet Syndr Gene Ther* 2013;4.

Microbiota in Immune Pathogenesis and the Prospects for Pre and Probiotic Dietetics in Psoriasis

Garima Pandey¹, Abhay Kumar Pandey^{2*}, S. S. Pandey¹, B. L. Pandey³

¹Department of Dermatology and Venereology, Institute of Medical Sciences, Banaras Hindu University, Varanasi, India

²Department of Physiology, All India Institute of Medical Sciences, Bhopal, India

³Department of Pharmacology, Institute of Medical Sciences, Banaras Hindu University, Varanasi, India

Psoriasis is a common autoimmune inflammatory disease wherein pathogenesis is advanced by fundamental genetic predisposition/s in concert with environmental triggers. Inflammation in psoriasis may represent efforts of innate immune system to target pathogens for restoring immune homeostasis. Aberrant microbiota may resist elimination efforts by sheer advantage of several fold gene pool as compared to the host. The microbes deregulate gene expression by the molecular insults targeting host immune system. Role of microbiota in autoimmunity dictates establishment of microbiome homeostasis and suppress host immune response; as a treatment approach. Dietary prebiotics and probiotics are of particular interest for prevention and amelioration of autoimmune inflammatory diseases, due to their potential to foster healthy host-microbiome relationship. The rational dietetics aims towards balancing friendly versus enemical microbes via manipulation of gut environment and modulation of immune system to improve regulation of inflammatory and autoimmune mechanisms.

INTRODUCTION

Increasing prevalence of autoimmune disorders and global epidemic of 'modern age' chronic inflammatory diseases due to changing lifestyle and environment is currently witnessed (Pandey *et al.*, 2014). A genetic predisposition underlies autoimmune diseases, with immune, hormonal and environmental factors contributing to the clinical manifestations. Clinical manifestations are restricted to susceptible individuals, although the autoimmunity processes have wider occurrence. Genetic predisposition influences the disease precipitation by exposure to environmental triggers as sunlight, diet, allergens, infectious agents and several other environmental insults. Infectious agents commonly trigger autoimmune diseases. Microbial products can aggravate T cell responses to self as well as non self antigens. Environmental factors may shift the balance of T cells

Key words: Psoriasis, Autoimmunity, Microbiota, Dysbiosis, Endotoxins, Epigenetic, Superantigens.

***Corresponding Author:** Abhay Kumar Pandey, Department of Physiology, All India Institute of Medical Sciences, Bhopal, India.

Email: abhay.physiology@gmail.com

between inflammatory interferon gamma producing Th1 cells and IL-4 and IL-5 producing Th2 cells in an individual. Lifestyle factors apparently affect immune function. Interaction between dietary factors and other exposure/s are current significant research areas of pathogenesis and therapeutics of autoimmune disorders (Pandey and Pandey, 2013).

The microbial flora in the gut interacts with diet and influences the host immune cells with mutual benefit to both the microbes and the host (Lee and Mazamania, 2010; Lee *et al.*, 2010). This aspect has major implication for prevention and mitigation of psoriasis, the most common immune mediated inflammatory disease.

The Gut Microbiome

The gut mucosal immune system is the largest lymphoid organ in our body, intricately involved in regulation of immunity and inflammation characterizing autoimmune disorders. Gut immune system is strongly subject to changes by dysbiosis or imbalance among microbial species (Maynard *et al.*, 2012). Change in population of singular species of segmented filamentous bacteria in the gut may markedly influence emergence and stability of specific T lymphocyte subsets (Prakash *et al.*, 2011). Biochemical interactions of colonic microbes and immune cells have

implications for immune homeostasis. Examples include, action of serum amyloid A on dendritic cells (Ivanov *et al.*, 2009), ATP mediated activation of dendritic cells (Atarashi *et al.*, 2008), induction of TGF-beta expression by gut epithelial cells (Atarashi *et al.*, 2011), bacillus fragilis derived polysaccharide A action on dendritic cells (Mazamania *et al.*, 2005) and TR-2 regulatory T cell function (Round *et al.*, 2011). Changes in gut microbiota in response to diet, sanitation, antibiotic use, environmental chemicals, etc. can influence and impact maturation and balance of immune responses.

Dysbiosis and Immune Disorder

Innate immune system provocation in autoimmunity involves a crucial role of the Toll like receptors. These recognize specific forms of microbial nucleic acids and bind them to induce proinflammatory signals. The TLRs also recognize certain self antigens and mediate autoimmunity against cells with parallel expression of proinflammatory cytokines (Trivedi and Greidenger, 2009). The common proinflammatory autoimmune phenotypes are due to CD4+ T helper lymphocytes. Organ specific autoimmunity is driven by cell mediated immune responses aimed to attack intracellular foreign element. The Th1 cytokines IL-2 and interferon-gamma predominance foster development of such

responses, as in psoriasis (Smith and Germolec, 1999; Street and Mossman, 1991). The Th17 lymphocyte phenotype contributes to autoimmunity through procuring neutrophil chemo-attractant and activator cytokine IL-23 (Wild *et al.*, 1994). Several intracellular signal transduction pathways in the Treg (Suppressor) lymphocytes may be deregulated leading to inappropriate activation of naïve T cells and persistence of autoreactive cells in organ (Jain *et al.*, 2010). Altered quantities of STAT-3 (signal transducers and activators of transcription) influence autoimmunity in varied ways. There is direct selection, favouring Th-17 pro-inflammatory phenotype over the immune-inhibitory Treg phenotype of lymphocyte subsets. STAT-3 also influences vital T-cell biology of growth and survival as well as transcription of pro-inflammatory genes (Egwuagh, 2009).

The microbes may initiate autoreactivity by several mechanisms (Ercolini and Miller, 2009). Infection induced general proinflammatory environment serves as a bystander to promote deregulation of immune response and modification of endogenous proteins to autoantigens. Alternatively some microbial antigens may mimic structurally homologous self peptide of the host which initiates immune response. The increasingly emphasized mechanism

however, is production of super-antigens by certain microbes. These nonspecifically crosslink MHC-II to T cells primed to other antigens including self-antigens, leading to unintended stimulation and massive release of cytokines (Freidman *et al.*, 1991; Schiffenbouer *et al.*, 1998).

Intestinal dysbiosis generates endotoxin-peptidoglycan super antigens inducing autoimmune/inflammatory pathology in psoriasis. Immune response is directed at toxins produced by microorganisms in the gut, and psoriatic patients exhibit positive skin test to gut bacterial antigens (Baker *et al.*, 2006a; 2006b; Gyurcsovics and Bertok, 2003; Karotkii and Peslyak, 2005; Qayoom and Ahmed 2003; Stenina *et al.*, 2003). Gut and skin colonization by *Staphylococcus aureus*, *malassezia*, and *candida*, etc. cause exacerbation in psoriasis (Fry and Baker 2007). Autoimmune reactions can be advanced or blocked by commensal bacteria affecting the innate and adaptive arms of immune responses and interlinking mechanisms. Whether immunity and autoimmunity is affected by specific or multiple lineages of microbes that may shift the homeostatic balance toward reduced or exaggerated reactivity in host-microbiota interaction, is yet unsettled (Chervonsky, 2013).

New genomic understanding indicates collective metagenome (Interactive

microbe-host genomes) as determinant of outcome of host-microbiome interactions. Diverse microbial metabolites may affect expression of genes associated with immune responses and autoimmunity. Microbiota accumulates incurring adaptations to persist, with the genes impacting the disease process. Success in reversing autoimmunity by reduction of microbes that have evolved capability to block vitamin-D receptors and thus evade immune-elimination supports the view (Proal *et al.*, 2009). Huge load of metabolites resulting from over million microbial genes, are juxtaposed to interact with small number of proteins made by human genes. Immune function thus is manipulated (Honda and Littman, 2012). Genetic predisposition leads to adverse consequences following interaction with other entities. Altered cytokine profile can change cellular milieu and xenobiotic exposures may vitiate the micro-world of immune cells. Altered gene expression as a result of an epigenetic change and surge of proinflammatory mediators weakens immune regulation and tolerance. The risk of autoantigen availability and consequent autoimmune disease therefore increases (Pillai, 2013).

Involvement of Gut-brain Neuro-immune axis

Dendritic cells in the gastrointestinal (GI) tract send processes throughout the gut

epithelium in the lumen to interact with microbes. The signals to humoral immune system to produce immunoglobulin A secretion prevails (Corthesy and Spertini, 1999). The secreted IgA checks microbes from penetrating gut epithelium. The dendritic cells are in close proximity to the nerves in GI tract and their function is modulated by sensory neuropeptide CGRP (Calcitonin gene related peptide) (Hosoi *et al.*, 1993). The brain is informed about microbiota via the vagus nerve (Gochler *et al.*, 1999). Bacterial endotoxins or inflammatory cytokines like IL-1 β and TNF- α may stimulate the vagus nerve. The vagal reflex in response, suppresses proinflammatory cytokine release by intestinal macrophages (Borovikova *et al.*, 2000).

The gut-brain axis modulates the feeding behavior as well (Bercik *et al.*, 2009). Peripheral afferent nerves transmit “danger” signals and elicit neural reflexes, regulating immune responses. Prototype inflammatory reflex operates through afferent sensory and efferent motor vagus nerve fibers (Tracey, 2009). The central projections regulate hypothalamic-pituitary-adrenal (HPA) neuro-humoral axis and causes glucocorticosteroid release. The afferent vagal activity triggered by endotoxins and cytokines, also sends efferent signals to thymes and the splenic nerve (Rossa-Ballima *et al.*, 2008). Spleen is the primary target for

signals in the efferent pathway of vagal antiinflammatory reflex. Over 90% of systemically released TNF- α during early endotoxaemia, is of splenic origin. Vagal stimulation attenuates TNF- α release. Vagus effect is mediated through adrenergic splenic nerve activation and β -2 adrenoceptors mediate inhibition of TNF- α release (Vida *et al.*, 2011). The mechanism of precipitation and aggravation of psoriasis lesions by β -adrenergic blocking drugs is thus explained. The innervating vagus nerve fibers coordinate with the gut microbiome via bidirectional communications (Lee and Mazmanian, 2010).

Dietetic Management of Gut Microbiota

Diverse commensal bacteria reside in the gut. Individual species appear to have distinct and opposite roles in gut immune response. Certain commensal microbes preferentially drive regulatory Treg lymphocyte development, while others promote pro-inflammatory Th1 γ cell development in gut lymphoid tissue (Kamada and Nunez 2013). Altered microbiota associates with several inflammatory diseases (Kamada *et al.*, 2013).

Microbiota in gut serves a number of nutritional health effects. The composition and performance is influenced by diet, as a key factor. Dietetic strategy attempts to

suppress harmful bacterial species while stimulating beneficial bacteria. Such a strategy implies selective consumption of probiotics and/or prebiotics and diet rich in fiber content.

Prebiotics

Most plant origin foods contain dietary fiber. The fibers undergoing bacterial degradation include polysaccharides e.g., resistant starch, pectin, inulin, guar gum and oligosaccharides. Structural polysaccharide like cellulose and lignin are insoluble and are not degraded by bacteria e.g., wheat bran. Such components have ability to hold water and thus increase mass of stool. This facilitates motility and cleansing of gut microbial mass. Soluble fibers also increase fecal output and promote bacterial biomass via fermentation. Prebiotics are non digestible food components that selectively enhance growth and/or activity of one or limited bacterial species with beneficial health consequences. Prebiotic has to remain undigested and unabsorbed in upper segment of GI tract. The majority of prebiotics are oligosaccharides, however some polysaccharides also serve as substrates to colonic bacteria and stimulate their activity. Prominent prebiotic activity is seen with non-digestible oligosaccharides including xylo-oligosaccharides, galacto-oligosaccharides and isomalt-

oligosaccharides (Van Loo *et al.*, 1999). Anaerobes constitute over 99% of fecal flora. These break down the available carbohydrate substrate to short chain fatty acids, acetate, propionate, butyrate and gas hydrogen and carbon dioxide. Propionates and acetates are absorbed and contribute to the fuel resource of the body. Butyrate is a preferred energy resource for colonic epithelium and plays a role in proliferation and differentiation (Litvak *et al.*, 1998). The hydrogen generated through fermentation reactions is primarily used by methanogenic, acetogenic or sulfate reducing microorganisms (Gibson *et al.*, 1990).

Stimulation of Bifidobacteria and Lactobacilli is advantageous due to their immuno-modulatory abilities and inhibitory potential against pathogens. These reduce ammonia formation and lower blood cholesterol, and serve to restore gut microbiota damaged by antibiotics (Goldin, 1998). Selective stimulation of indigenous beneficent microbe strains, impart antimicrobial potential of prebiotics. The beneficent microbes selectively possess exo-glycosidase enzymes that enable utilization of oligosaccharides (Perrin *et al.*, 2001). The uptake and intracellular metabolism by the microbes as an alternate strategy (even by non-beneficent microbe) is a possibility. Prebiotic selection needs refinement for avoiding the later alternate

pathways. Antimicrobial potential is particularly vested in smaller prebiotic molecules e.g., chito-oligosaccharides (Vishnukumar *et al.*, 2005). Bacterial species have different preferences for energy substrates. Diet is a strong direct means of influencing gut microbial colonization. Dietary fiber effectively causes major shifts in composition of gut microbiota and directly affects mucosal immune system. Fiber therefore improves chronic inflammatory disorders and systemic immune responses. Anti-inflammatory potential is contributed through short chain fatty acids generated upon microbial fermentation of prebiotic components (Huda-Faujan *et al.*, 2010). Butyrate is richly produced from resistance starch, soluble fiber, and inulin foods and increases the regulatory Treg lymphocyte percentage with reduced production of interferon- γ . As a consequence, there is down regulation of inflammation (Vieira *et al.*, 2013). Higher levels of butyrate causes activation of nuclear transcription factor and peroxisome proliferator activator receptor gamma PPAR- γ (Luhrs *et al.*, 2002; Schwab *et al.*, 2006). PPAR- γ activity inhibits proinflammatory pathways like STAT, AP-1 and NF κ B pathways, specifically desired in psoriasis management (Sertzing *et al.*, 2008).

Acetate is produced in greater abundance than butyrate following

fermentation. Acetate levels are raised more in circulation than in the gut due to absorption. Immune cells bear specific G-protein coupled receptors for binding the small chain fatty acid ligands. Specific GPR43 receptor mediated protection against colitis through induction of Fox P3+ IL-10 producing regulatory Treg cells has been demonstrated (Smith *et al.*, 2013). Prebiotics inhibit pathogen adherence to gut epithelium, with positive effects on lipid metabolism and stimulation of mineral (especially calcium) absorption in colon, through influencing the gut microbiota (Gibson and Roberfroid, 1995).

Polyphenolic Bioactive Food Constituents

Colonic microbiota serves as primary agents for metabolism of polyphenolic dietary constituents. These are esters, glycosides and polymers contained in fruits and vegetables and bear protective antioxidant potential. Citrus fruits, apples, grapes, berries, wine, tea, soy and many vegetables including onion are rich sources of dietary polyphenols comprising complex mixtures. The nature of the gut microbiome therefore, determines extraction of their bioactive antioxidant and anti-inflammatory principles. Polyphenolics like isoflavones and flavonones are absorbed to a small extent. Proanthocyanidines and anthocyanidines

are obligatorily metabolized by gut microbiota (Manach *et al.*, 2005). Strategic modulation of composition of gut microbiota may enhance utilization and bioavailability of polyphenols and their potential health benefits. Synergistic benefit of simultaneous oligosaccharide consumption is observed with isoflavones (Mathey *et al.*, 2004; Piazza *et al.*, 2007).

Probiotics

Two bacterial phyla, Bacterioides (bifidobacteria) and firmicutes (lactobacilli) comprise over 90% of the gut microbiota (Mariat *et al.*, 2009). These produce large number of vitamins including the B group vitamins; synthesize amino acids; and carry out biotransformation of bile; and ferment undigested fiber and mucus. Beneficial microbes produce antimicrobial substances and promote mucin secretion and directly interfere with pathogen adherence to the epithelium (Rogier *et al.*, 2014). The bifidobacteria and lactobacilli can be introduced in the gut and encouraged to multiply as probiotics by supplementing prebiotic rich diet. Administration of these probiotic strains in healthy individual enhances mucin and nonspecific IgA secretion and the phagocytotic potential of surveillance cells.

Thus, probiotic strengthens the gut barrier and opposes entry of foreign

antigens. Attenuation of proinflammatory responses adds to this. Probiotics compete for nutrients at the site of attachment to gut epithelium and inhibits colonization by pathogens with simultaneous release of antimicrobial products. Generation of lipopolysaccharides and peptidoglycans detrimental to the host is checked by probiotic mechanisms (Tlaskova-Hogenova *et al.*, 2004). A major role for peptidoglycans is emphasized in psoriasis pathogenesis (Baker *et al.*, 2006). The development of regulatory T cells, the Type1 and Type2 helper T cell and Th1 γ helper cells are all subject to signals by intestinal microbiota. The pro-inflammatory responses attenuated by probiotics include IL-8, MCP-1, MIP-1 and RANTES, proinflammatory cytokines and lipid mediators evoked by pathogens. RANTES activation is of major pathogenic significance in psoriasis (Raychaudhuri *et al.*, 1999).

Stable health promoting relationships between host gut and microbiota is crucially determined by pattern recognition receptors viz. Toll like receptors (TLRs) and Nod like receptor (NLRs) (Abreu, 2010). Microbe associated molecular patterns signal to affect epithelial cytoprotection, survival/proliferation pathways and barrier function (Rakoff-Nohoum *et al.*, 2004). TLR activation upregulates proinflammatory mediators facilitating

immune defense. The NLRs are present in the cytoplasm of immune cells and their stimulation by commensal associated signals regulate inflammatory responses, contributing to gut homeostasis (Yeretssian 2012). Disturbed interaction of microbiota with pattern recognition receptors underlies diseases with exaggerated inflammation (Lavelle *et al.*, 2010; Maynard *et al.*, 2012). An investigation of mechanism by which specific probiotic strain triggers reaction can help to indicate appropriate choice of probiotic for prophylactic use in diverse inflammatory diseases. The probiotic benefit is external to gut through complex microbe-immune system interaction in immune mediated inflammatory diseases. The gut-brain axis and inflammation reflexes have a bearing in this context.

Dysbiosis of gut microbiome may be a secondary consequence of a primary adversary that must be diagnosed and managed. The metabolic phenotypes of individuals determine the composition of gut microflora independently of dietary pattern or even genotype (Serino *et al.*, 2012). Long term consumption of high fat diet impacts the microbiota directly, and indirectly through alteration of redox state. Antioxidant dietary supplements improve gut microbiota profile (Espley *et al.*, 2014). A protective potential of gut microbiota against pathogen invasion is promoted by prebiotic combination with

other bioactive plant principles and quality proteins in the diet. Lupine fermentation is noteworthy in this regard (Berthkiene *et al.*, 2013). Probiotic administration by rectal route is far superior to oral route for successful immuno-modulation (Matthes *et al.*, 2010).

The Pathogenic Gut-skin Linkage in Psoriasis

Cytokinaemia and exaggerated inflammation in psoriasis is crucially linked to absorption of endotoxins from pathogenic gut bacteria (Gyurcsovics and Bertok, 2003). A strong evidence of defective barrier function of gut is observed (Scarpa *et al.*, 2000). Immunopathologic process in psoriasis extend from gut to skin (Michaelsson *et al.*, 1997). The evidence is compelling to address issues of gut barrier integrity and dysbiosis as a rational consideration in psoriasis therapy. Psoriasis therapies are conventionally focused on managing the consequences of immune mediated

inflammatory pathology and causes several adverse effects. Regular incorporation of prebiotic and probiotic dietetics is a rational consideration, neither too expensive nor unsafe. Its significance particularly appeals for prophylaxis in individuals with familial predisposition, and subjects bearing other heightened risk factors (Gupta *et al.*, 2013). Evidence based personalized pre- and pro-biotic dietetics in management of psoriasis has appeal. This is challenging however, microbiome management is subject to individual contexts and not amenable to ordinary laboratory means. Metagenomic investigations may be a solution to match the indications, with appropriate dietetic address. This may comprise a rational and lead to better quality management for psoriasis and autoimmune inflammatory diseases at large.

CONFLICT OF INTEREST

The authors claim no conflict of interest.

REFERENCES

- Abreu MT. Toll-like receptor signalling in the intestinal epithelium: how bacterial recognition shapes intestinal function. *Nat Rev Immunol* 2010;10:131–144.
- Angélica T Vieira, Mauro M. Teixeira, Flaviano S. Martins. The Role of Probiotics and Prebiotics in Inducing Gut Immunity. *Front Immunol* 2013;4:445.
- Atarashi K, Nishimura J, Shima T, Umesaki Y, Yamamoto M, Onoue M *et al.* ATP drives lamina propria Th-17 cell differentiation. *Nature* 2008;455(7214):808–812.
- Atarashi K, Tanoue T, Shima T, Imaoka A, Kuwahara T, Momose Y. *et al.* Induction of colonic regulatory T cells by indigenous Clostridium species. *Science* 2011;331(6015):337–341.
- Baker BS, Laman JD, Powles A, vander Fits L,

- Voerman JSA, Melief MJ, Fry L. Peptidoglycan and peptidoglycan-specific Th1 cells in psoriatic skin lesions. *J Pathol* 2006;209:174–181.
- Baker BS, Powles A, Fry L. Peptidoglycan: a major aetiological factor for psoriasis? *Trends Immunol* 2006;27:545–551.
- Bartkiene E, Jakobsonė I, Juodeikiene G, Vidmantiene D, Pugajeva I, Bartkevics V. Effect of lactic acid fermentation of lupine wholemeal on acrylamide content and quality characteristics of wheat-lupine bread. *Int J Food Sci Nutr* 2013;64:890–896.
- Bercik P, Verdu EF, Foster JA, Lu J, Scharringa A, Kean I, Wang L *et al.* Role of gut-brain axis in persistent abnormal feeding behaviour in mice following eradication of helicobacter Pylori infection. *M J Physiol Regul Integr Comp Physiol* 2009;296:R587–R594.
- Borovikova LV, Ivanova S, Jhang M, Yang H, Botchkina GI, Watkins LR *et al.* Vagus nerve stimulation attenuates the systemic inflammatory response to endotoxins. *Nature* 2000;405:458–462.
- Chervonsky AV. Microbiota and autoimmunity. *Cold Spring Harb Perspect Biol* 2013;5:a007294.
- Corthésy B, Spertini F. Secretory immunoglobulin A: from mucosal protection to vaccine development. *Biol Chem* 1999;380:1251–1262.
- Egwuagu EC. STAT3 in CD⁺ T helper cell differentiation and inflammatory diseases. *Cytokines* 2009;47(3):149–146.
- Ercolini AM, Miller SD. The role of infections in autoimmune disease. *Clin Exp Immunol* 2009; 155:1–15.
- Espley RV, Butts CA, Laing WA, Martell S, Smith H, McGhie TK *et al.* Dietary flavonoids from modified apple reduce inflammation markers and modulate gutmicrobiota in mice. *J Nutr* 2014;144:146–154.
- Friedman SM, Posnett DN, Tumang JR, Crow MK *et al.* A potential role of microbial superantigens in the pathogenesis of systemic autoimmune disease. *Arthritis Rheum* 1991; 34:468–480.
- Fry L, Baker BS. Triggering psoriasis: the role of infections and medications. *Clin Dermatol* 2007;25:606–615.
- Gibson GR, Cummings JH, Macfarlane GT, Allison C, Segal I, Vorster HH, Walker AR. Alternative pathways for hydrogen disposal during fermentation in the human colon. *Gut* 1990;31:679–683.
- Gibson GR, Roberfroid MB. Dietary modulation of the human colonic microbiota: introducing the concept of prebiotics. *J Nutr* 1995;125: 1401–1412.
- Goehler LE, Gaykema RP, Nguyen KT, Lee JE, Tilders FJ, Maier SF, Watkins LR. Interlukin 1beta in immune cells of the abdominal vagus nerve: a link between the immune and the nervous system? *J Neuroscience* 1999;19: 2799–2806.
- Goldin BR. Health benefits of probiotics. *Br J Nutr* 1998;80:S203–S207.
- Gupta AK, Pandey SS, Pandey BL. Effectiveness of conventional drug therapy of plaque psoriasis in the context of consensus guidelines: a prospective observational study in 150 patients. *Ann Dermatol* 2013;25: 156–162.
- Gyurcsovics K, Bertók L. Pathophysiology of

- psoriasis: coping endotoxins with bile acid therapy. *Pathophysiology* 2003;10:57–61.
- Gyurcsovics K, Bertók L. Pathophysiology of psoriasis: coping endotoxins with bile acid therapy. *Pathophysiology* 2003;10:57–61.
- Honda K, Littman DR. The microbiome in infectious disease and inflammation. *Annu Rev Immunol* 2012;30:759–795.
- Hosoi J, Murphy GF, Egan CL, Learner EA, Grabbe S, Asahina A *et al.* Regulation of langerhans cell function by nerves containing calcitonin gene related peptide. *Nature* 1993; 363:159–163.
- Huda-Faujan N, Abdulmir AS, Fatimah AB, Anas OM, Shuhaimi M, Yazid AM, Loong YY. The impact of the level of the intestinal short chain Fatty acids in inflammatory bowel disease patients versus healthy subjects. *Open Biochem J* 2010;4:53–58.
- Ivanov II, Atarashi K, Manel N, Brodie EL, Shima T, Karaoz V. *et al.* Induction of intestinal Th-17 cells by segmented filamentous bacteria. *Cell* 2009;139:485–498.
- Jain N, Nguyen H, Chambers C, Kang J, Kamada N, Núñez G. Role of the gut microbiota in the development and function of lymphoid cells. *J Immunol* 2013;190:1389–1395.
- Kamada N, Seo SU, Chen GY, Núñez G. Role of the gut microbiota in immunity and inflammatory disease. *Nat Rev Immunol* 2013;13:321–335.
- Karotkii NG, Peslyak MY. Psoriasis as a consequence of incorporation of beta-streptococci into the microbiocenosis of highly permeable intestine (a pathogenic concept). *Vestn Dermatol Venereol* 2005;1:9–18.
- Lavelle EC, Murphy C, O'Neill LA, Creagh EM. The role of TLRs, NLRs, and RLRs in mucosal innate immunity and homeostasis. *Mucosal Immunol* 2010;3:17–28.
- Lee YK, Mazmanian SK. Has the microbiota played a critical role in the evolution of the adaptive immune system? *Science* 2010;330: 1768–1773.
- Litvak DA, Evers BM, Hwang KO, Hellmich MR, Ko TC, Townsend CM Jr. Butyrate-induced differentiation of Caco-2 cells is associated with apoptosis and early induction of p21Waf1/Cip1 and p27Kip1. *Surgery* 1998; 124:161–169; discussion 169–170.
- Lühns H, Kudlich T, Neumann M, Schaubert J, Melcher R, Gostner A, Scheppach W, Menzel TP. Butyrate-enhanced TNFalpha-induced apoptosis is associated with inhibition of NF-kappa B. *Anticancer Res* 2002;22:1561–1568.
- Manach C, Williamson G, Morand C, Scalbert A, Rémésy C. Bioavailability and bioefficacy of polyphenols in humans. I. Review of 97 bioavailability studies. *Am J Clin Nutr* 2005; 81:230S–242S.
- Mariat D, Firmesse O, Levenez F, Guimaraes V, Sokol H, Doré J, Corthier G, Furet JP. The Firmicutes/Bacteroidetes ratio of the human microbiota changes with age. *BMC Microbiol* 2009;9:123.
- Mathey J, Puel C, Kati-Coulibaly S, Bennetau-Pelissero C, Davicco MJ, Lebecque P, Horcajada MN, Coxam V. Fructooligosaccharides maximize bone-sparing effects of soy isoflavone-enriched diet in the ovariectomized rat. *Calcif Tissue Int* 2004;75:169–179.
- Matthes H, Krummnerl T, Giensch M, Wolff C, Schulze J. Clinical trial: probiotic treatment of acute distal ulcerative colitis with rectally

- administered *Escherichia coli* Nissle 1917 (EcN). *BMC Complement Altern Med* 2010; 10:13.
- Maynard CL, Elson CO, Hatton RD, Weaver CT. Reciprocal interactions of the intestinal microbiota and the immune system. *Nature* 2012;489(7415):231–241.
- Mazmanian SK, Liu CH, Tzianabos AO and Kasper DL. An immunomodulatory molecules of symbiotic bacteria directs maturation of the host immune system. *Cell* 2005;122(1):107–108.
- Michaëlsson G, Kraaz W, Hagforsen E, Pihl-Lundin I, Löf L, Scheynius A. The skin and the gut in psoriasis: the number of mast cells and CD3+ lymphocytes is increased in non-involved skin and correlated to the number of intraepithelial lymphocytes and mast cells in the duodenum. *Acta Derm Venereol* 1997;77: 343–346.
- Pandey AK, Pandey G, Pandey SS, Pandey BL. Human biology of diet and lifestyle linked chronic inflammatory non-communicable disease epidemic. *Human Biol Rev* 2014;3:25–42.
- Pandey AK, Pandey G. Nutrition research perspectives in autoinflammatory disorders. *Indian J Rheumatol* 2013;8:30–36.
- Perrin P, Pierre F, Patry Y, Champ M, Berreur M, Pradal G, Bornet F, Meflah K, Menanteau J. Only fibres promoting a stable butyrate producing colonic ecosystem decrease the rate of aberrant crypt foci in rats. *Gut* 2001;48: 53–61.
- Piazza C, Privitera MG, Melilli B, Incognito T, Marano MR, Leggio GM, Roxas MA, Drago F. Influence of inulin on plasma isoflavone concentrations in healthy postmenopausal women. *Am J Clin Nutr* 2007;86:775–780.
- Pillai S. Rethinking mechanisms of autoimmune pathogenesis. *J Autoimmun* 2013;45:97–103.
- Prakash T, Oshima K, Morita H, Fukuda S, Imaoka A, Kumar N. *et al.* Complete Genome Sequences of Rat and Mouse Segmented Filamentous Bacteria, a Potent Inducer of Th17 Cell Differentiation. *Cell Host Microbe* 2011;10:273–284.
- Proal AD, Albert PJ, Marshall TG. Dysregulation of the Vitamin D nuclear receptor may contribute to the higher prevalence of some autoimmune disease in women. *Ann NY Acad Sci* 2009;1173:252–259.
- Qayoom S, Ahmad QM. Psoriasis and helicobacter pylori. *Indian J Dermatol Venereol Leprol* 2003;69:133–134.
- Rakoff-Nahoum S, Paglino J, Eslami-Varzaneh F, Edberg S, Medzhitov R. Recognition of commensal microflora by toll-like receptors is required for intestinal homeostasis. *Cell* 2004; 118:229–241.
- Raychaudhuri SP, Jiang WY, Farber EM, Schall TJ, Ruff MR, Pert CB. Upregulation of RANTES in psoriatic keratinocytes: a possible pathogenic mechanism for psoriasis. *Acta Derm Venereol* 1999;79:9–11.
- Rogier EW, Frantz AL, Bruno ME, Kaetzel CS. Secretory IgA is Concentrated in the Outer Layer of Colonic Mucus along with Gut Bacteria. *Pathogens* 2014;3:390–403.
- Rosas-Ballina M, Ochani M, Parrish WR, Ochani K, Harris YT, Huston JM *et al.* Splenic nerve is required for cholinergic anti-inflammatory pathway control of TNF in endotoxemia. *Proc Natl Acad Sci USA* 2008;105:11008–11013.

- Scarpa R, Manguso F, D'Arienzo A, D'Armiento FP, Astarita C, Mazzacca G, Ayala F. Microscopic inflammatory changes in colon of patients with both active psoriasis and psoriatic arthritis without bowel symptoms. *J Rheumatol* 2000;27:1241–1246.
- Schiffenbauer J, Soos J, Johnson H. The possible role of bacterial superantigens in the pathogenesis of autoimmune disorders. *Immunol Today* 1998;19:117–120.
- Schwab M, Reynders V, Ulrich S, Zahn N, Stein J, Schröder O. PPAR-gamma is a key target of butyrate-induced caspase-3 activation in the colorectal cancer cell line Caco-2. *Apoptosis* 2006;11:1801–1811.
- Serino M, Blasco-Baque V, Burcelin R. Microbes on-air: gut and tissue microbiota as targets in type 2 diabetes. *J Clin Gastroenterol*. 2012;46:S27–S28.
- Sertznig P, Seifert M, Tilgen W, Reichrath J. Peroxisome Proliferator-Activated Receptors (PPARs) and the Human Skin. *Am J Clin Dermatol*. 2008;9:15–31.
- Smith DA, Germolec DR. Introduction to immunology and autoimmunity. *Environ Health Perspect* 1999;107(5):661–665.
- Smith PM, Howitt MR, Panikov N, Michaud M, Gallini CA, Bohlooly-Y M, Glickman JN, Garrett WS. The microbial metabolites, short-chain fatty acids, regulate colonic Treg cell homeostasis. *Science* 2013;341:569–573.
- Stenina MA, Kulagin VI, Rudkovskaya ZV et al. Role of disturbances of intestine barrier function in pathogenesis of psoriasis in children. *Russian Journal of Skin and Sexually Transmitted Diseases* 2003;2:20–23.
- Street NE, Mossman TR. Functional diversity of T lymphocytes due to secretion of different cytokine patterns. *FASEB J* 1991;5:171–177.
- Tlaskalová-Hogenová H, Stepánková R, Hudcovic T, Tucková L, Cukrowska B, Lodinová-Zádníková R et al. Commensal bacteria (normal microflora), mucosal immunity and chronic inflammatory and autoimmune diseases. *Immunol Lett* 2004;93:97–108.
- Tracey KJ. Reflex control of immunity. *Nat Rev Immunol* 2009;9:418–428.
- Trivedi S, Greidinger EL. Endosomal Toll-like receptors in autoimmunity: mechanisms for clinical diversity. *Therapy* 2009;6(3):433–442.
- Van Loo J, Franck A, Roberfroid M. Functional food properties of non-digestible oligosaccharides. *Br J Nutr* 1999;82:329.
- Vida G, Peña G, Deitch EA, Ulloa L. α 7-cholinergic receptor mediates vagal induction of splenic norepinephrine. *J Immunol* 2011; 186:4340–4346.
- Vishu Kumar AB, Varadaraj MC, Gowda LR, Tharanathan RN. Characterization of chito-oligosaccharides prepared by chitosan analysis with the aid of papain and Pronase, and their bactericidal action against *Bacillus cereus* and *Escherichia coli*. *Biochem J* 2005;391:167–175.
- Wilde B, Thewissen M, Damoiseaux J, Paassen PV, Witzke O, Tervaert JWC. T cells in ANCA-associated vasculitis: what can we learn from lesional versus circulating T cells? *Arthritis Res Ther* 1994;12(1):204.
- Yeretssian G. Effector functions of NLRs in the intestine: innate sensing, cell death, and disease. *Immunol Res* 2012; 54:25–36.

List of Reviewers for Volume 2, Issue 1-2

Acknowledging Reviewers, 2015

Dr. Anirvan Chatterjee

Indian Institute of Technology Bombay, Mumbai, India

Dr. Anjali Karande

Indian Institute of Science, Bengaluru, India

Dr. Arundhati Mandal

Reliance Life Sciences, Navi Mumbai, India

Dr. Chinmay Kumar Panda

Chittaranjan National Cancer Institute, Kolkata, India

Dr. Cristina Maccalli

Azienda Ospedaliera Universitaria Senese, Siena, Italy

Dr. Dilek Demirkol

Biochemistry Department, Ege University, Izmir, Turkey

Dr. Hema Purandarey

SRL Diagnostics, Mumbai, India

Dr. Jaya Vyas

Kokilaben Dhirubai Ambani Hospital, Mumbai, India

Dr. Kayzad Nilgiriwala

The Foundation for Medical Research, Mumbai, India

Dr. Koodlur Lokesh

Vijayanagara, Sri Krishnadevaraya University, Bellary, India

Dr. Krutika Desai

Mithibai College of Arts, Chauhan Institute of Science and A. J. College of Commerce and Economics, Mumbai, India

Dr. Manuel Mari-Beffa

Department of Cell Biology, University of Málaga, Málaga, Spain

Dr. Nagraj Huilgol

Nanavati Superspeciality Hospital, Mumbai, India

Dr. Neelam Shirsat

Advanced Centre for Training, Research and Education in Cancer, Navi Mumbai, India

Dr. Nishigandha Naik

Haffkine Institute for Training, Research and Testing, Mumbai, India

Dr. Prabhudas Patel

The Gujarat Cancer and Research Institute, Ahmedabad, India

Dr. Prathibha Shetty

Reliance Life Sciences, Navi Mumbai, India

Dr. Prochi Madon

Jaslak Hospital and Research Centre, Mumbai, India

Dr. Rajani Bhisey

Advanced Centre for Training, Research and Education in Cancer, Navi Mumbai, India

Dr. Ramon Alemany

Catalan Institute of Oncology, Barcelona, Spain

Dr. Ravindran Ankathil

Rajiv Gandhi Centre for Biotechnology, Thiruvananthapuram, India

Dr. Sarita Gupta

Department of Biochemistry, The Maharaja Sayajirao University of Baroda, Vadodara, India

Dr. Sasikumar Menon

Therapeutic Drug Monitoring Laboratory,
Mumbai, India

Dr. Shreelaja Nair

Tata Institute of Fundamental Research,
Mumbai, India

Dr. Surendra Kumar Trigun

Department of Zoology, Banaras Hindu
University, Varanasi, India

Dr. Tarala Nandedkar

National Institute Research in Reproductive
Health, Mumbai, India

Dr. Vaijayanti Kale

National Centre for Cell Science, Pune, India

Dr. Vandana Patravale

Institute of Chemical Technology, Mumbai, India

Information for Authors

Biomedical Research Journal

Sunandan Divatia School of Science,
Bhaidas Sabhagriha Building, Bhaktivedanta Swami Marg,
Vile Parle (W), Mumbai - 400056, INDIA.
Email: brj.sos@nmims.edu



Biomedical Research Journal (BRJ) accepts the following article types for publication

Editorial

Authors who are considering submitting an editorial should contact either the Editors-in-Chief with a brief outline of the proposed contribution before submission. Editorials are welcome on any topic; however, they may also be related to articles previously published in the Journal. Editorials have no abstract and no keywords, and are usually restricted to 1000 words, up to 10 references and up to 2 tables or figures. The Editors-in-Chief can be contacted at brj.sos@nmims.edu.

Original Research Articles

Original research articles which have not been published previously, except in a preliminary form may be submitted as original full length research papers. Research articles must contain an abstract, a list of up to six keywords, and are limited to 3,500 words in length.

Review Articles

Review articles which are topical and are a critical assessment of any aspect of mentioned areas. Review articles must contain an abstract, a list of up to ten keywords, and are limited to 5,000 words in length. Authors whose manuscripts exceed 5,000 words are advised to contact the Editorial Office prior to submission.

Letters to the Editor

Letters to the Editor relating to published work in the journal are welcome. Letters should be closely related to the contents of the referred article.

After reading the Instructions to Authors, please visit our online submission system to submit your manuscript.

Submission checklist

It is hoped that this list will be useful during the final checking of an article prior to sending it to the journal's Editor for review. Ensure that the following items are present:

- One Author designated as corresponding Author:
 - E-mail address
 - Full postal address
 - Telephone(s) and fax numbers
- All necessary files have been uploaded
- Keywords (as comprehensive as possible)
- All figure captions
- All tables (including title, description, footnotes)
- The copyright form has been completed and uploaded

Further considerations

- Manuscript has been "spellchecked" and is written in good English
- Title is clear and unambiguous
- If the manuscript is an original research article it should contain a structured abstract, if the manuscript is a review article it should contain an unstructured abstract
- References are in the correct format for this journal
- All references mentioned in the Reference list are cited in the text, and vice versa
- Permission has been obtained for use of copyrighted material from other sources (including the Web)
- Colour figures are clearly marked as being intended for colour reproduction on the Web (free of charge), and in print or to be reproduced in colour on the Web (free of charge) and in black-and-white in print
- If only colour on the Web is required, black and white versions of the figures are also supplied for printing purposes
- The manuscript conforms to the limits imposed on original research (3,500 words); Review articles (5,000 words), excluding the abstract, keywords, references, tables and figures)

For any further information please contact the Author Support Department at brj.sos@nmims.edu

Prior to Submission

BRJ will consider manuscripts prepared according to the guidelines adopted by the International Committee of Medical Journal Editors ("Uniform requirements for manuscripts submitted to biomedical journals", available as a PDF from (3500 words) <http://www.icmje.org>). Authors are advised to read these guidelines.

Previous Publication

Submission of an article implies that the work described is:

- Not published previously (except in the form of an abstract or as part of a published lecture or academic thesis)
- Not under consideration for publication elsewhere
- The publication is approved by all authors and tacitly or explicitly by the responsible authorities where the work was carried out, and that, if accepted, it will not be published elsewhere in the same form, in English or in any other language, without the written consent of the Publisher.

Ethics

Work on human beings that is submitted to Journal should comply with the principles laid down in the Declaration of Helsinki: Recommendations guiding physicians in biomedical research involving human subjects, adopted by the 18th World Medical Assembly, Helsinki, Finland, June 1964, amended by the 29th World Medical Assembly, Tokyo, Japan, October, 1975, the 35th World Medical Assembly, Venice, Italy, October 1983, and the 41st World Medical Assembly, Hong Kong, September 1989. The manuscript should contain a statement that the work has been approved by the appropriate ethical committees related to the institution(s) in which it was performed and that subjects gave informed consent to the work. Studies involving experiments with animals must state that their care was in accordance with institution guidelines. Patients' and volunteers' names, initials and hospital numbers, should not be used and patient confidentiality must be maintained.

Conflict of Interest

By means of a "Conflict of interest statement", all authors must disclose any financial and personal relationships with other people or organizations that could inappropriately influence (bias) their work. If there are no conflicts of interest, please state "No conflict of interest declaration". This document should be uploaded as a separate file alongside the submitted manuscript.

Role of the Funding Source

All sources of funding should be declared as an acknowledgment at the end of the text.

Authorship and Acknowledgments

All authors must be accredited on the paper and all must submit a completed Author Form with their submission. The form must be signed by the corresponding author on behalf of all authors and can be scanned and uploaded.

Copyright

Upon acceptance of an article, Authors will be asked to transfer copyright. This transfer will ensure the widest possible dissemination of information. A letter will be sent to the corresponding Author confirming receipt of the manuscript. A form facilitating transfer of copyright will be provided.

If excerpts from other copyrighted works are included, the Author(s) must obtain written permission from the copyright owners and credit the source(s) in the article.

General Points

We accept Word format of submitted manuscript. Always keep a backup copy of the electronic file for reference and safety. Save your files using the default extension of the program used. It is important that the file be saved in the native format of the word processor used. The text should be in single-column format. Keep the layout of the text as simple as possible. Most formatting codes will be removed and replaced on processing

the article. In particular, do not use the word processor's options to justify text or to hyphenate words. However, do use bold face, italics, subscripts, superscripts etc. Do not embed "graphically designed" equations or tables, but prepare these using the word processor's facility. When preparing tables, if you are using a table grid, use only one grid for each individual table and not a grid for each row. If no grid is used, use tabs, not spaces, to align columns. The electronic text should be prepared in a way very similar to that of conventional manuscripts. Do not import the figures into the text file but, instead, indicate their approximate locations directly in the electronic text and on the manuscript. See also the section on Preparation of electronic illustrations.

To avoid unnecessary errors you are strongly advised to use the "spellchecker" function of your word processor.

Presentation of Manuscript

Please write your text in good English (American or British usage is accepted, but not a mixture of these). Italicize expressions of Latin origin, for example, *in vivo*, *et al.*, *per se*. Use decimal points (not commas).

Title Page

Provide the following data on the title page:

Title

Concise and informative. Titles are often used in information-retrieval systems. Avoid abbreviations and formulae where possible.

Author names and affiliations

Where the family name may be ambiguous (e.g., a double name), please indicate this clearly. Present the Authors' affiliation addresses (where the actual work was done) below the names. Indicate all affiliations with a lower-case superscript letter immediately after the Author's name and in front of the appropriate address. Provide the full postal address of each affiliation, including the country name, and, if available, the e-mail address of each Author.

Corresponding Author

Clearly indicate who is willing to handle correspondence at all stages of refereeing and publication, also post-publication. Ensure that telephone and fax numbers (with country and area code) are provided in addition to the e-mail address and the complete postal address.

Present/permanent address

If an Author has moved since the work described in the article was done, or was visiting at the time, a "Present address" (or "Permanent address") may be indicated as a footnote to that Author's name. The address at which the Author actually did the work must be retained as the main, affiliation address. Superscript Arabic numerals are used for such footnotes.

Suggestions for reviewers

Please supply the names of up to three potential reviewers for your manuscript. Please do not suggest reviewers from your own institution, previous or current collaborators. Please provide full names, addresses and email addresses of suggested reviewers. Please note: the final choice of reviewers is that of the Editor and the journal reserves the right for choice of final reviewers.

Abstract

A concise and factual abstract of no more than 250 words is required. The abstract must be structured for original research articles. The abstract should be divided by subheadings as follows: Objectives, Materials and Methods, Results, Discussion and Conclusion.

The abstract should not be structured for review articles. The abstract should state briefly the purpose of the research, the principal results and major conclusions. An abstract is often presented separate from the article, so it must be able to stand alone.

Keywords

After the abstract provide a maximum of six keywords, to be chosen from the Medical Subject Headings from Index Medicus. These keywords will be used for indexing purposes

Abbreviations

Define abbreviations or acronyms that are not standard in this field at their first occurrence in the article; in the abstract and also in the main text after it. Ensure consistency of abbreviations throughout the article.

Text

This should start on the third page and should be subdivided into the following sections: Introduction, Patients or Materials and Methods, Results, Discussion and Conclusions, Acknowledgements.

References

Responsibility for the accuracy of bibliographic citations lies entirely with the authors. Please ensure that every reference cited in the text is also present in the reference list (and vice versa). Any references cited in the abstract must be given in full. "Unpublished data" and "Personal communications" are not allowed. As an alternative, say in the text, for example, '(data not shown)' or '(Dr D. Saranath, School of Science, NMIMS (Deemed-to-be) University, Mumbai)'. Citation of a reference as "in press" implies that the item has been accepted for publication and a copy of the title page of the relevant article must be submitted.

Indicate references by (first author, year) in the text.

Examples:

Kulkarni J, Khanna A. Functional hepatocyte-like cells derived from mouse embryonic stem cells: A novel *in vitro*

hepatotoxicity model for drug screening. *Toxicol In Vitro* 2006;20:1014-1022.

Bhatnagar R, Dabholkar J, Saranath D. Genome-wide disease association study in chewing tobacco associated oral cancers. *Oral Oncol* 2012;48(9):831-835.

Molinolo AA, Hewitt S, Amornphimoltham PI, Keelawat S, Saranath D, Gutkind JS *et al.* Dissecting the Akt/mTOR signaling network: emerging results from the head and neck cancer tissue array initiative. *Clin Cancer Res* 2007;13:4964-4973.

Saranath D. Integrated Biology and Molecular Pathology of Oral Cancer. *In: Saranath D, editor. Contemporary Issues in Oral Cancer.* Oxford Press, 2001:30-71.

List all authors if the total number of authors is seven. For more than seven authors, first six authors should be listed, followed by "et al." For further details you are referred to "Uniform Requirements for Manuscripts submitted to Biomedical Journals" (*J Am Med Assoc* 1997;277:927-934).

Figure Captions, Tables, Figures and Schemes

Present these, in the given order, at the end of the article. They are described in more detail below. High-resolution graphics files must always be provided separate from the main text file.

Footnotes

Footnotes should be used sparingly. Number them consecutively throughout the article, using superscript Arabic numbers. Many word processors build footnotes into the text, and this feature may be used. Should this not be the case, indicate the position of footnotes in the text and present the footnotes themselves on a separate sheet at the end of the article. Do not include footnotes in the Reference list.

Table footnotes

Indicate each footnote in a table with a superscript lowercase letter.

Tables

Number tables consecutively in accordance with their appearance in the text. Place footnotes to tables below the table body and indicate them with superscript lowercase letters. Avoid vertical rules. Be sparing in the use of tables and ensure that the data presented in tables do not duplicate results described elsewhere in the article.

Nomenclature and Units

Follow internationally accepted rules and conventions: use the international system of units (SI). If other quantities are mentioned, give their equivalent in SI.

Preparation of Electronic Illustrations

- Make sure you use uniform lettering and sizing of your original artwork.
- Save text in illustrations as "graphics" or enclose the font.

- Only use the following fonts in your illustrations: Arial or Times Roman.
- Number the illustrations according to their sequence in the text.
- Use a logical naming convention for your artwork files.
- Provide all illustrations as separate files and as hardcopy printouts on separate sheets.
- Provide captions to illustrations separately.
- Produce images near to the desired size of the printed version.

Formats

Regardless of the application used, when your electronic artwork is finalised, please "save as" or convert the images to one of the following formats (Note the resolution requirements for line drawings, halftones, and line/halftone combinations given below):

EPS: Vector drawings. Embed the font or save the text as "graphics".

TIFF: Colour or greyscale photographs (halftones): always use a minimum of 300 dpi.

TIFF: Bitmapped line drawings: use a minimum of 1000 dpi.

TIFF: Combinations bitmapped line/half-tone (colour or greyscale): a minimum of 500 dpi is required.

DOC, XLS or PPT: If your electronic artwork is created in any of these Microsoft Office applications please supply "as is".

Please do not

- Supply embedded graphics in your wordprocessor (spreadsheet, presentation) document;
- Supply files that are optimised for screen use (like GIF, BMP, PICT, WPG); the resolution is too low;
- Supply files that are too low in resolution;
- Submit graphics that are disproportionately large for the content.

If, together with your accepted article, you submit usable colour figures then it will be ensured that at no additional charge these figures will appear in colour on the Web (e.g., ScienceDirect and other sites) in addition to colour reproduction in print.

Captions

Ensure that each illustration has a caption. Supply captions separately, not attached to the figure. A caption should comprise a brief title (not on the figure itself) and a description of the illustration. Keep text in the illustrations themselves to a minimum but explain all symbols and abbreviations used.

Line drawings

The lettering and symbols, as well as other details, should have

proportionate dimensions, so as not to become illegible or unclear after possible reduction; in general, the figures should be designed for a reduction factor of two to three. The degree of reduction will be determined by the Publisher. Illustrations will not be enlarged.

Do not use any type of shading on computer-generated illustrations.

Photographs (halftones)

Remove non-essential areas of a photograph. Do not mount photographs unless they form part of a composite figure. Where necessary, insert a scale bar in the illustration (not below it), as opposed to giving a magnification factor in the caption. Note that photocopies of photographs are not acceptable.

Preparation of supplementary data

Electronic supplementary material to support and enhance your scientific research is accepted as supplementary file. Supplementary files offer the Author additional possibilities to publish supporting applications, movies, animation sequences, high-resolution images, background datasets, sound clips and more. Supplementary files supplied will be published online alongside the electronic version of your article. In order to ensure that your submitted material is directly usable, please ensure that data is provided in one of our recommended file formats. Authors should submit the material in electronic format together with the article and supply a concise and descriptive caption for each file.

Proofs

when your manuscript is received by the Publisher it is considered to be in its final form. Proofs are not to be regarded as "drafts". One set of page proofs in PDF format will be sent by e-mail to the corresponding author, to be checked for typesetting/editing. No changes in, or additions to, the accepted (and subsequently edited) manuscript will be allowed at this stage. Proofreading is solely your responsibility.

The corrected article will be published as quickly and accurately as possible. In order to do this we need your help. When you receive the (PDF) proof of your article for correction, it is important to ensure that all of your corrections are sent back to us in one communication. Subsequent corrections will not be possible, so please ensure your first sending is complete. Note that this does not mean you have any less time to make your corrections just that only one set of corrections will be accepted.

About Sunandan Divatia School of Science

Sunandan Divatia School of Science was started in 2007 with a view to provide undergraduate and post graduate students an opportunity to be a part of the unique learning methodology of the university, which lays emphasis on academic excellence combined with industry oriented training. With the boom in information technology and more and more sophistication in instrumentation techniques, there is now a very thin dividing line between the various disciplines of science. Therefore, there is a

greater need for flexibility in scientific thought as well as training manpower on an interdisciplinary plane. With this thought in view, the SVKM's NMIMS introduced, highly innovative and unique interdisciplinary courses at the School of Science from the academic year 2007-2008. The goal of the School of Science is to be a Center of Excellence in the domain of Pure and Applied Science by providing quality education and research.

Courses Offered

Ph.D. in Biological Sciences, Chemistry (Regular and Professional) and Physiotherapy

Integrated M.Sc.-Ph.D. in Biological Sciences and Chemistry

M.Sc. in Biological Sciences, Chemistry (Analytical and Organic) and Statistics

Master of Physiotherapy [In collaboration with Nanavati Super Speciality Hospital, Mumbai, India]

Post-Graduate Diploma in Physician Assistance (2 years), Operation Theatre

Technology (1 year), Non-Invasive Cardiology (1 year) and Central Sterile Services

(1 year) [In collaboration with Asian Heart Institute and Research Centre, Mumbai, India]

Diploma in Clinical Research (Part time: 1 year) [In collaboration with C. B. Patel Research Centre, Mumbai, India]

Certificate Courses in Molecular Medicine and Molecular Oncology (Part time: 6 months) [for medical/science graduates]

Advanced Course in Clinical Data Management (Part time: 3 months) [In collaboration with C. B. Patel Research Centre, Mumbai, India]

Salient Features

Research constitutes a major thrust in all the courses offered at the School

Courses oriented to fulfill needs/demands of Research Institutions/Industry

Thrust Areas in Research

Cell Biology, Stem Cell Biology, Molecular Oncology, Reproductive Biology, Microbiology, Immunology, Pharmacology, Phytochemistry, Nanosciences, Applied Chemistry, Colloidal Chemistry and Applied Statistics

For More Information Please Contact:

Sunandan Divatia School of Science, NMIMS (Deemed-to-be) University

Tel: 91-22-42355958/59; Fax: 91-22-2611 4512; E-mail: admissions.sos@nmims.edu;

Visit us at: <http://science.nmims.edu>



The Aditya Birla Group: A Premium Global Corporation

A US \$40 billion corporation, the Aditya Birla Group is in the League of Fortune 500. Anchored by an extraordinary force of over 120,000 employees, belonging to 42 nationalities. The Group has topped the Nielsen's Corporate Image Monitor 2013-14 and emerged as the Number 1 corporate, the 'Best in Class', for the second consecutive year. Over 50 per cent of our revenues flow from our overseas operations in 36 countries.

- From being in every second can in the world, to shaping automobiles
- From building your homes to the highways that get you there
- From inks, dyes, plastics to every 5th tyre in the world
- From securing futures to empowering over 6 million dreams
- From connecting the world to enabling over a billion conversations a day
- From styling your wardrobe every three seconds, to bringing you the world of fashion

We are a big part of your life, because we know every small moment, is big for you!

Globally, we are:

- A metals powerhouse, among the world's most cost-efficient aluminium and copper producers. Hindalco-Novelis is the largest aluminium rolling company. It is one of the 3 biggest producers of primary aluminium in Asia, with the largest single location copper smelter
- No.1 in viscose staple fibre
- No.1 in carbon black
- The 4th largest producer of insulators
- The 4th largest producer of acrylic fibre
- Among the top 10 cement producers globally
- Among the best energy efficient fertilizer plants
- The largest Indian MNC with manufacturing operations in the USA

In India:

- A top fashion (branded apparel) and lifestyle player
- The 2nd largest player in viscose filament yarn

- The largest in the chlor-alkali sector
- Among the top 3 mobile telephony companies
- A leading player in life insurance and asset management
- Among the top 2 super-market chains in the retail business

Beyond business we

- Reach out to 7 million people in 3,000 villages annually in India through the Aditya Birla Centre for Community Initiatives and Rural Development, spearheaded by Mrs. Rajashree Birla
- Focus on: health-care, education, sustainable livelihood, infrastructure and espousing social reform
- Run 42 schools which provide quality education to 45,000 children. Of these 18,000 students belong to the underprivileged segment. Merit Scholarships are given to an additional 12,000 children from the interiors.
- Our 18 hospitals tend to more than a million villagers
- Ongoing education, healthcare and sustainable livelihood projects in Philippines, Thailand, Laos, Indonesia, Egypt, Korea and Brazil, lift thousands of people out of poverty.
- Set up the Aditya Birla India Centre at the London Business School

Transcending the conventional barriers of business because we believe it is our duty to facilitate inclusive growth, and we care.



ADITYA BIRLA GROUP

Big In Your Life

www.adityabirla.com

SVKM's
Narsee Monjee Institute of Management Studies

Deemed to be UNIVERSITY

V. L. Mehta Road, Vile Parle (W), Mumbai-400 056, INDIA.

Tel: 91-22-4235555 | **Fax:** 91-22-26114512

Email: nmims@nmims.edu | **Website:** www.nmims.edu